

Product Datasheet

IL-33 Antibody (1C5) NBP2-31145

Unit Size: 0.1 mg

Store at 4C short term. Aliquot and store at -20C long term. Avoid freeze-thaw cycles.

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NBP2-31145

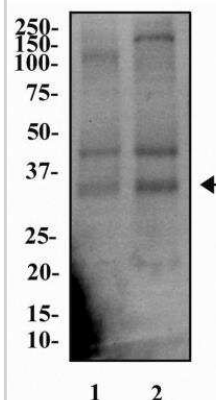
IL-33 Antibody (1C5)

Product Information	
Unit Size	0.1 mg
Concentration	1.0 mg/ml
Storage	Store at 4C short term. Aliquot and store at -20C long term. Avoid freeze-thaw cycles.
Clonality	Monoclonal
Clone	1C5
Preservative	0.05% Sodium Azide
Isotype	IgG2b Kappa
Purity	Protein A purified
Buffer	PBS
Product Description	
Host	Mouse
Gene ID	90865
Gene Symbol	IL33
Species	Human, Mouse
Immunogen	Synthetic peptide corresponding to an internal sequence of the human IL33 protein (between amino acids 100-270) [UniProt# O95760]
Product Application Details	
Applications	Western Blot, Immunohistochemistry, Immunohistochemistry-Paraffin
Recommended Dilutions	Western Blot 2 ug/ml, Immunohistochemistry 2-5 ug/ml, Immunohistochemistry-Paraffin 2-5 ug/ml
Application Notes	The canonical form of IL33 i.e. Isoform 1 is 270 amino acids long chain with predicted molecular weight of 30.8 kDa, however, this full length form gets processed via cleavage of n-terminal 111 amino acids (pro-peptide cleavage) to generate 159 amino acids long mature IL33 protein. In WB, the full length protein runs at ~30kDa, whereas, the processed mature form runs at ~18kDa position.

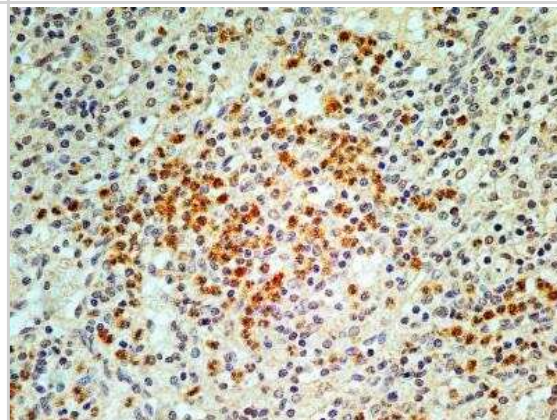


Images

Western Blot: IL-33 Antibody (1C5) [NBP2-31145] - Analysis of IL33 in (1) U2OS and (2) HUVEC cell lysates.



Immunohistochemistry-Paraffin: IL-33 Antibody (1C5) [NBP2-31145] - Analysis of IL33 protein in normal human spleen tissue sections using IL33 antibody (clone 1C5) at a concentration of 5 ug/ml. This antibody stained the nuclei as well as the cytoplasm of granulocytes and some other cell types in the red pulp of spleen tissue.



Western Blot: IL-33 Antibody (1C5) [NBP2-31145] - Analysis of IL33 recombinant protein using IL33 antibody (clone 1C5) at a concentration of 2 ug/ml.



Procedures

Western Blot Protocol for IL33 Antibody (NBP2-31145)

IL-33 Antibody (1C5):

Western Blot Protocol

1. Perform SDS-PAGE on samples to be analyzed, loading 25 ug of total protein per lane.
2. Transfer proteins to membrane according to the instructions provided by the manufacturer of the membrane and transfer apparatus.
3. Stain according to standard Ponceau S procedure (or similar product) to assess transfer success, and mark molecular weight standards where appropriate.
4. Rinse the blot.
5. Block the membrane using standard blocking buffer for at least 1 hour.
6. Wash the membrane in wash buffer three times for 10 minutes each.
7. Dilute anti-IL33 primary antibody in blocking buffer and incubate 1 hour at room temperature.
8. Wash the membrane in wash buffer three times for 10 minutes each.
9. Apply the diluted HRP conjugated secondary antibody in blocking buffer (as per manufacturers instructions) and incubate 1 hour at room temperature.
10. Wash the blot in wash buffer three times for 10 minutes each (this step can be repeated as required to reduce background).
11. Apply the detection reagent of choice in accordance with the manufacturers instructions.

Note: Tween-20 can be added to the blocking or antibody dilution buffer at a final concentration of 0.05-0.2%.



Immunohistochemistry-Paraffin Protocol for IL33 Antibody (NBP2-31145)**IL-33 Antibody (1C5):**

1. Deparaffinize the tissue sections by immersing the slides in Xylene with two changes for 10 min each. Sections should not get dried at any stage from this point.
2. Rehydrate the tissue sections by immersing the slides in decreasing grades of ethanol as follows:
 - a. Immerse in 100% ethanol with 2 changes for 5 minutes each
 - b. Immerse in 95% ethanol with 2 changes for 5 minutes each
 - c. Immerse in 90% ethanol for 5 minutes
 - d. Immerse in 70% ethanol for 5 minutes
 - e. Immerse in 50% ethanol for 5 minutes
 - f. Immerse in distilled water for 5 minutes
3. Antigen Retrieval (Microwave Method):
 - a. Immerse the slides in a microwave compatible tray containing 10 mM Sodium Citrate buffer (pH 6.0) with 0.05% Tween 20.
 - b. Boil the slides and maintain the sub-boiling temperature for 5 minutes in the microwave. Thereafter, take out the tray very carefully and cool it at room temperature (RT) for about 30 minutes.
 - c. Wash the slides 3 times, 3 minutes each by immersing them in TBST (Tris Buffered Saline having 0.05% Tween 20).
4. Quenching of Endogenous Peroxidase:
 - a. Incubate the slides in 3% hydrogen peroxide prepared in methanol for 15 minutes (at RT, in dark conditions).
 - b. Wash the slides in TBST 3 times, 3 minutes each.
5. Protein Blocking:
 - a. Incubate the sections with background sniper solution at RT for 15 minutes (Biocare Medicals, USA).
 - b. Wash the sections 3 times, 3 min each by immersing the slides in TBST.
6. Primary Antibody:
 - a. Dilute the primary antibody at 5ug/ml concentration using PBS as a diluent.
 - b. Incubate the sections with diluted primary antibody for 90 minutes at RT in a humidified chamber.
 - c. Thereafter, wash the slides 4 times, 5 minutes each with TBST.
7. Probe (Secondary Reagent):
 - a. Incubate with MACH 1 Mouse probe for 15 minutes at RT.
 - b. Incubate for 30 min at room temperature with HRP-Polymer (Biocare Medical, USA).
 - c. Wash the slides with TBST 4 times, 5 minutes each
8. Chromogen:
 - a. Mix 32ul of DAB Chromogen with 1 ml of DAB substrate buffer (Biocare Medical, USA).
 - b. Apply 200ul DAB mixture/section and incubate at RT in dark conditions (few seconds - 5 minutes).
 - c. As soon as an appropriate color develops, rinse the slides with deionized water (2-3 brief rinses).
9. Counter stain:
 - a. Counter stain with Hematoxylin for 30 seconds (Vector Labs, USA).
 - b. Wash in deionized water for 1-2 minutes to clear the extra stain.
 - c. Incubate the slides in bluing solution or Scott's water twice for 2 minutes each time.
10. Dehydrate the sections in increasing grades of alcohols:
 - a. 50% alcohol for 1 minute
 - b. 70% for 1 minute
 - c. 90% for 1 minute
 - d. 95% for 1 minute
 - e. 100% for 1 minute
 - f. Xylene with 2 changes for 2 minutes each
11. Mount with DPX mount and cover-slip glass (Fisher Scientific, USA), carefully not allowing any air bubbles to enter.

NOTE:- This protocol is provided as a reference tool only. Depending upon the type of tissues /tissue processing and reagents employed, the end user will need to optimize the final conditions for achieving an expected staining.



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