Product Datasheet

Arginase 1/ARG1/liver Arginase Antibody
NBP1-32731

Unit Size: 0.1 mg
Aliquot and store at -20C or -80C. Avoid freeze-thaw cycles.

Publications: 2
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**NBP1-32731**  
Arginase 1/ARG1/liver Arginase Antibody

### Product Information

<table>
<thead>
<tr>
<th><strong>Unit Size</strong></th>
<th>0.1 mg</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Concentration</strong></td>
<td>1.0 mg/ml</td>
</tr>
<tr>
<td><strong>Storage</strong></td>
<td>Aliquot and store at -20C or -80C. Avoid freeze-thaw cycles.</td>
</tr>
<tr>
<td><strong>Clonality</strong></td>
<td>Polyclonal</td>
</tr>
<tr>
<td><strong>Preservative</strong></td>
<td>0.02% Sodium Azide</td>
</tr>
<tr>
<td><strong>Isotype</strong></td>
<td>IgG</td>
</tr>
<tr>
<td><strong>Purity</strong></td>
<td>Immunogen affinity purified</td>
</tr>
<tr>
<td><strong>Buffer</strong></td>
<td>PBS</td>
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</tbody>
</table>

### Product Description

| **Host** | Rabbit |
| **Gene ID** | 383 |
| **Gene Symbol** | ARG1 |
| **Species** | Human, Mouse, Rat |
| **Reactivity Notes** | Expected cross reactivity based on sequence homology: Porcine (89%). |
| **Immunogen** | Full length human Arginase 1 Recombinant protein. |

### Product Application Details

| **Applications** | Western Blot, Flow Cytometry, Immunocytochemistry/Immunofluorescence, Immunohistochemistry, Immunohistochemistry-Frozen, Immunohistochemistry-Paraffin, Immunoprecipitation |
| **Recommended Dilutions** | Western Blot 0.1 - 0.5 ug/ml, Flow Cytometry 5 - 10 ug/ml, Immunohistochemistry 1:200 - 1:500, Immunocytochemistry/Immunofluorescence 1:100-1:1000, Immunoprecipitation 1:100-1:1000, Immunohistochemistry-Paraffin 1:200 - 1:500, Immunohistochemistry-Frozen 1:200 - 1:500 |

### Images

Western Blot: Arginase 1/ARG1/liver Arginase Antibody [NBP1-32731] -  
Non-transfected (-) and transfected (+) HepG2 whole cell extracts (30 ug) were separated by 10% SDS-PAGE, and the membrane was blotted with Arginase 1 antibody.
Immunocytochemistry/Immunofluorescence: Arginase 1/ARG1/liver
Arginase Antibody [NBP1-32731] - HepG2 cells were fixed for 10 minutes using 10% formalin and then permeabilized for 5 minutes using 1X PBS + 0.05% Triton X-100. The cells were incubated with anti-ARG1 at 10 ng/ml overnight at 4C and detected with an anti-rabbit DyLight 488 (Green) at a 1:500 dilution. Nuclei were counterstained with DAPI (Blue). Cells were imaged using a 40X objective.


Flow Cytometry: Arginase 1/ARG1/liver Arginase Antibody [NBP1-32731] - An intracellular stain was performed on HepG2 with NBP1-32731 and a matched isotype control. Cells were fixed with 4% PFA and then permeabilized with 0.1% saponin. Cells were incubated in an antibody dilution of 5 ug/mL for 30 minutes at room temperature, followed by Rabbit IgG (H+L) Cross-Adsorbed Secondary Antibody, DyLight 550 (SA5-10033, Thermo Fisher).

Western Blot: Arginase 1/ARG1/liver Arginase Antibody [NBP1-32731] - Arginase 1 antibody detects Arginase 1 protein. Various whole cell extracts (30ug) were separated by 12% SDS-PAGE, and the membrane was blotted with Arginase 1 antibody diluted at 1:1000.
Western Blot: Arginase 1/ARG1/liver Arginase Antibody [NBP1-32731] - Mouse tissue extract (50 ug) was separated by 12% SDS-PAGE, and the membrane was blotted with Arginase 1 antibody diluted at 1:10000.

Western Blot: Arginase 1/ARG1/liver Arginase Antibody [NBP1-32731] - Rat tissue extract (50 ug) was separated by 12% SDS-PAGE, and the membrane was blotted with Arginase 1 antibody diluted at 1:10000.


Immunohistochemistry-Paraffin: Arginase 1/ARG1/liver Arginase Antibody [NBP1-32731] - Tissue section of the mouse liver using 1:200 dilution of ARG1 antibody (NBP1-32731). The signal was developed using HRP-DAB method which followed counterstaining of the cells with hematoxylin.

Immunoprecipitation: Arginase 1/ARG1/liver Arginase Antibody [NBP1-32731] - Analysis was performed using Arginase 1 antibody EasyBlot anti-Rabbit IgG was used as a secondary reagent. Arginase 1 protein from HepG2 whole cell extracts using 5 ug of Arginase 1 antibody.

Publications


Procedures

Western Blot Protocol for Arginase 1/ARG1/liver Arginase Antibody (NBP1-32731)

Western Blot Protocol

1. Perform SDS-PAGE on samples to be analyzed, loading 10-25 ug of total protein per lane.
2. Transfer proteins to PVDF membrane according to the instructions provided by the manufacturer of the membrane and transfer apparatus.
3. Stain the membrane with Ponceau S (or similar product) to assess transfer success, and mark molecular weight standards where appropriate.
4. Rinse the blot TBS - 0.05% Tween 20 (TBST).
5. Block the membrane in 5% Non-fat milk in TBST (blocking buffer) for at least 1 hour.
6. Wash the membrane in TBST three times for 10 minutes each.
7. Dilute anti-ARG1 primary antibody in blocking buffer and incubate overnight at 4C with gentle rocking.
8. Wash the membrane in TBST three times for 10 minutes each.
9. Incubate the membrane in diluted HRP conjugated secondary antibody in blocking buffer (as per manufacturer’s instructions) for 1 hour at room temperature.
10. Wash the blot in TBST three times for 10 minutes each (this step can be repeated as required to reduce background).
11. Apply the detection reagent of choice in accordance with the manufacturers instructions.

Immunohistochemistry-Paraffin Protocol for Arginase 1/ARG1/liver Arginase Antibody (NBP1-32731)

Immunohistochemistry-Paraffin Embedded Sections

Antigen Unmasking:
Bring slides to a boil in 10 mM sodium citrate buffer (pH 6.0) then maintain at a sub-boiling temperature for 10 minutes. Cool slides on bench-top for 30 minutes.

Staining:
1. Wash sections in deionized water three times for 5 minutes each.
2. Wash sections in wash buffer for 5 minutes.
3. Block each section with 100-400 ul blocking solution for 1 hour at room temperature.
4. Remove blocking solution and add 100-400 ul diluted primary antibody. Incubate overnight at 4 C.
5. Remove antibody solution and wash sections in wash buffer three times for 5 minutes each.
6. Add 100-400 ul biotinylated diluted secondary antibody. Incubate 30 minutes at room temperature.
7. Remove secondary antibody solution and wash sections three times with wash buffer for 5 minutes each.
8. Add 100-400 ul Streptavidin-HRP reagent to each section and incubate for 30 minutes at room temperature.
9. Wash sections three times in wash buffer for 5 minutes each.
10. Add 100-400 ul DAB substrate to each section and monitor staining closely.
11. As soon as the sections develop, immerse slides in deionized water.
12. Counterstain sections in hematoxylin.
13. Wash sections in deionized water two times for 5 minutes each.
14. Dehydrate sections.
15. Mount coverslips.
Immunocytochemistry/Immunofluorescence Protocol for Arginase 1/ARG1/liver Arginase Antibody (NBP1-32731)

Immunocytochemistry Protocol

Culture cells to appropriate density in 35 mm culture dishes or 6-well plates.

1. Remove culture medium and wash the cells briefly in PBS. Add 10% formalin to the dish and fix at room temperature for 10 minutes.
2. Remove the formalin and wash the cells in PBS.
3. Permeabilize the cells with 0.1% Triton X100 or other suitable detergent for 10 min.
4. Remove the permeabilization buffer and wash three times for 10 minutes each in PBS. Be sure to not let the specimen dry out.
5. To block nonspecific antibody binding, incubate in 10% normal goat serum from 1 hour to overnight at room temperature.
6. Add primary antibody at appropriate dilution and incubate overnight at 4C.
7. Remove primary antibody and replace with PBS. Wash three times for 10 minutes each.
8. Add secondary antibody at appropriate dilution. Incubate for 1 hour at room temperature.
9. Remove secondary antibody and replace with PBS. Wash three times for 10 minutes each.
10. Counter stain DNA with DAPI if required.

*The above information is only intended as a guide. The researcher should determine what protocol best meets their needs. Please follow safe laboratory procedures.*
Flow (Intracellular) Protocol for Arginase 1/ARG1/liver Arginase Antibody (NBP1-32731)
Protocol for Flow Cytometry Intracellular Staining

Sample Preparation.
1. Grow cells to 60-85% confluency. Flow cytometry requires between 2 x 105 and 1 x 106 cells for optimal performance.
2. If cells are adherent, harvest gently by washing once with staining buffer and then scraping. Avoid using trypsin as this can disrupt certain epitopes of interest. If enzymatic harvest is required, use Accutase, Collagenase, or TrypLE Express for a less damaging option.
3. Reserve 100 μL for counting, then transfer cell volume into a 50 mL conical tube and centrifuge for 8 minutes at 400 RCF.
   a. Count cells using a hemocytometer and a 1:1 trypan blue exclusion stain to determine cell viability before starting the flow protocol. If cells appear blue, do not proceed.
4. Re-suspend cells to a concentration of 1 x 106 cells/mL in staining buffer (NBP2-26247).
5. Aliquot out 1 mL samples in accordance with your experimental samples.

Tip: When cell surface and intracellular staining are required in the same sample, it is advisable that the cell surface staining be performed first since the fixation and permeabilization steps might reduce the availability of surface antigens.

Intracellular Staining.
Tip: When performing intracellular staining, it is important to use appropriate fixation and permeabilization reagents based upon the target and its subcellular location. Generally, our Intracellular Flow Assay Kit (NBP2-29450) is a good place to start as it contains an optimized combination of reagents for intracellular staining as well as an inhibitor of intracellular protein transport (necessary if staining secreted proteins). Certain targets may require more gentle or transient permeabilization protocols such as the commonly employed methanol or saponin-based methods.

Protocol for Cytoplasmic Targets:
Optional: Perform cell surface staining as described in the previous section.
1. Fix the cells by adding 100 μL fixation solution (such as 4% PFA) to each sample for 10-15 minutes.
2. Permeabilize cells by adding 100 μL of a permeabilization buffer to every 1 x 106 cells present in the sample. Mix well and incubate at room temperature for 15 minutes.
   a. For cytoplasmic targets, use a gentle permeabilization solution such as 1X PBS + 0.5% Saponin or 1X PBS + 0.5% Tween-20.
   b. To maintain the permeabilized state throughout your experiment, use staining buffer + 0.1% of the permeabilization reagent (i.e. 0.1% Tween-20 or 0.1% Saponin).
3. Following the 15 minute incubation, add 2 mL of the staining buffer + 0.1% permeabilizer to each sample.
4. Centrifuge for 5 minutes at 400 RCF.
5. Discard supernatant and re-suspend in 1 mL of staining buffer + 0.1% permeabilizer.
6. Stain each sample at 1 μL/ 1 x 106 cells of primary antibody or 1-3 μL/ 1 x 106 cells for directly conjugated antibodies. Mix well and incubate at room temperature for 30 minutes- 1 hour. Gently mix samples every 10-15 minutes.
7. Following the primary/conjugate incubation, add 2 mL/sample of staining buffer +0.1% permeabilizer and centrifuge for 5 minutes at 400 RCF.
8. Remove supernatant and re-suspend each sample in 2 mL staining buffer + 0.1% permeabilizer, repeat wash for 5 minutes at 400 RCF.
9. If using a directly conjugated antibody, after the second wash, re-suspend cell pellet to a final volume of 500 μL per sample and proceed with flow analysis.
Limitations
This product is for research use only and is not approved for use in humans or in clinical diagnosis. Primary Antibodies are guaranteed for 1 year from date of receipt.

For more information on our 100% guarantee, please visit www.novusbio.com/guarantee

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