Product Datasheet

NCOR1 Antibody (7A7A9) - BSA Free
NBP1-28863

Unit Size: 0.1 ml

Store at 4C short term. Aliquot and store at -20C long term. Avoid freeze-thaw cycles.

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Publications: 3

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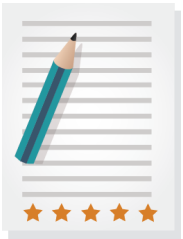
Updated 10/23/2024 v.20.1



NBP1-28863

NCOR1 Antibody (7A7A9) - BSA Free

Product Information

Unit Size	0.1 ml
Concentration	This product is unpurified. The exact concentration of antibody is not quantifiable.
Storage	Store at 4C short term. Aliquot and store at -20C long term. Avoid freeze-thaw cycles.
Clonality	Monoclonal
Clone	7A7A9
Preservative	0.03% Sodium Azide
Isotype	IgG1
Purity	Unpurified
Buffer	Ascites

Product Description

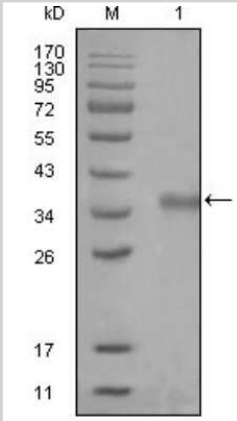
Host	Mouse
Gene ID	9611
Gene Symbol	NCOR1
Species	Human
Specificity/Sensitivity	Specific for NCOR1.
Immunogen	Purified recombinant fragment of NCOR1 (aa1-192) expressed in E. Coli.

Product Application Details

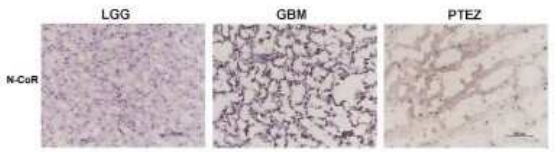
Applications	Western Blot, ELISA, Immunocytochemistry/ Immunofluorescence, Immunohistochemistry, Immunohistochemistry-Paraffin
Recommended Dilutions	Western Blot 1:500 - 1:2000, ELISA 1:10000, Immunohistochemistry 1:10 - 1:500, Immunocytochemistry/ Immunofluorescence, Immunohistochemistry-Paraffin 1:200 - 1:1000
Application Notes	Use in ICC/IF was reported in scientific literature (PMID: 31661545).

Images

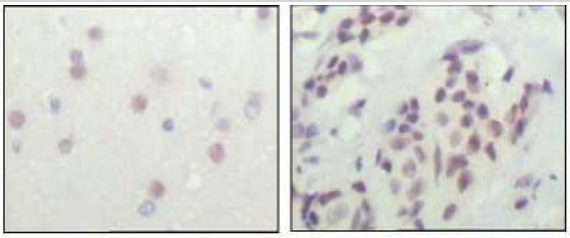
Western Blot: NCOR1 Antibody (7A7A9) [NBP1-28863] - Analysis using NCOR1 mouse mAb against truncated Trx-NCOR1 recombinant protein (1).



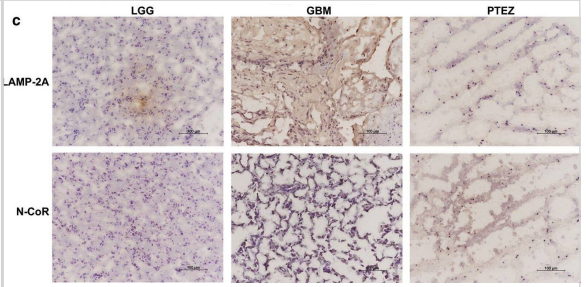
Immunohistochemistry: NCOR1 Antibody (7A7A9) [NBP1-28863] - Analysis of NCOR1 (brown signal) in glioma clinical samples. Nucleus (blue signal) was stained with hematoxylin. Study displayed downregulation of NCOR1 in GBM centers. Image collected and cropped by CiteAb from the following publication (//pubmed.ncbi.nlm.nih.gov/33761934/) licensed under a CC-BY license.



Immunohistochemistry-Paraffin: NCOR1 Antibody (7A7A9) [NBP1-28863] - Analysis of human cerebra (left) and breast carcinoma tissue (right), showing nuclear location with DAB staining using NCOR1 mouse mAb.

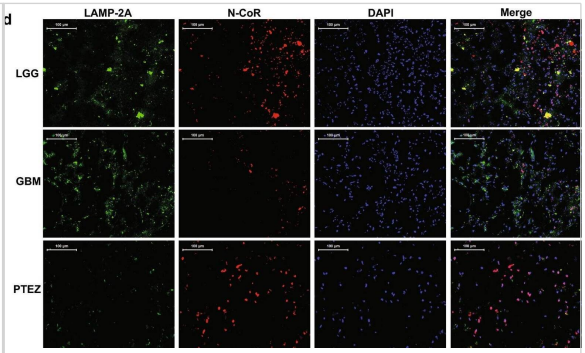


Autophagy degrades key postsynaptic proteins during LTD.a Confocal images of dendrites immunolabeled with an antibody against the extracellular region of GluA2 under control conditions or 15 min after LTD induction and in the absence or presence of Dynamin-1 inhibitory peptide (50 µM) or SBI-0206965 (500 nM), a selective inhibitor of the ULK1 kinase activity. Inhibitors were applied 25 min before, during and 15 min after the pulses. Scale bar: 10 µm. Graph showing the surface labeling of GluA2, normalized to dendritic length under the aforementioned conditions. Bars represent mean values ± SEM. N = 9 independent experiments. Statistical analysis was performed using one-way ANOVA ($F(8, 72) = 7.411$, $P < 0.0001$) (Tukey's test Pcontrol-control/D > 0.99, Pcontrol-control/S = 0.9971, PNMDA-NMDA/D = 0.0451, PNMDA-NMDA/S = 0.0008, PDHPG-DHPG/D = 0.0017, PDHPG-DHPG/S = 0.0002). b Confocal images of dendrites of neurons expressing 4 scrambled sequences (sh-scramble), or 4 sh-RNAs against atg5 (sh-atg5), immunolabeled with an antibody against the extracellular region of GluA2 under control conditions or 15 min after LTD induction. Graph showing the surface labeling of GluA2, normalized to dendritic length under the aforementioned conditions. Bars represent mean values ± SEM. N = 10 independent experiments. Statistical analysis was performed using one-way ANOVA ($F(5, 54) = 30.02$, $P < 0.0001$) (Tukey's test, Pcontrol/scr-control/atg5 = 0.0626, PNMDA/scr-NMDA/atg5 < 0.0001, PDHPG/scr-DHPG/atg5 < 0.0001, Pcontrol/atg5-NMDA/atg5 > 0.99, P control/atg5-DHPG/atg5 = 0.8602, Pcontrol/scr-NMDA/scr = 0.0008, Pcontrol/scr-DHPG/scr < 0.0001). c Representative images of consecutive confocal z-planes of cultured neurons immunostained with antibodies against PSD95, LC3, and MAP2 to label the dendrites, 15 min after cLTD. Note the colocalization of PSD95 and LC3 in dendritic spines (yellow arrows) and in the dendritic shaft (white arrows), in consecutive z-planes. Scale bar: 10 µm. Graph showing the percentage of PSD95 puncta co-localizing with LC3 in consecutive confocal z-planes in dendritic spines and shafts in control neurons or 15 min after chemically induced NMDAR- or mGluR-LTD. Bars represent

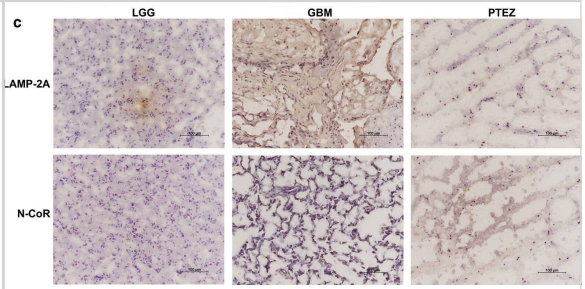


mean values ± SEM. N = 8 independent experiments. Statistical analysis was performed by one-way ANOVA ($F(5,42) = 48.43$, $P < 0.0001$) (Tukey's test for dendritic shaft, Pcontrol-NMDA = 0.0569, Pcontrol-DHPG = 0.1948, for dendritic spines, Pcontrol-NMDA < 0.0001, Pcontrol-DHPG < 0.0001). d Western blot analysis for GluA2 and PSD95 in lysates of cultured neurons in control conditions or 15 min after NMDAR- and mGluR-LTD and in the presence or absence of Bafilomycin A1 (50 µM) for 15 min before, during, and 15 min after the NMDA and DHPG pulses. e Western blot analysis for GluA2 and PSD95 in lysates of cultured neurons in control conditions or 15 min after NMDAR- and mGluR-LTD and in the presence or absence of SBI-0206965 (500 nM) for 30 min before, during, and 15 min after the NMDA and DHPG pulses. f Western blot analysis for GluA2 and PSD95 in lysates of cultured shscrambled or sh-atg5 expressing neurons in control conditions or 15 min after NMDAR- and mGluR-LTD. d–f Graphs showing the levels of PSD95 and GluA2 levels in the indicated conditions, normalized to total protein levels. Bars represent mean values ± SEM. Statistical analysis was performed by one-way ANOVA. d (N = 9 independent experiments) PSD95: $F(5,48) = 15.08$, $P < 0.0001$ (Tukey's test Pcontrol-control/Baf = 0.7566, Pcontrol-NMDA = 0.0016, Pcontrol-DHPG = 0.0081, PNMDA-NMDA/Baf < 0.0001, PDHPG-DHPG/Baf = 0.0013. GluA2: $F(5,48)=6.627$, $P < 0.0001$ (Tukey's test Pcontrol-control/Baf = 0.9692, Pcontrol-NMDA = 0.0014, Pcontrol-DHPG = 0.0067, PNMDA-NMDA/Baf = 0.0421, PDHPG-DHPG/Baf = 0.0127. e (N = 7 independent experiments) PSD95: $F(5,36) = 23.80$, $P < 0.0001$. (Tukey's test Pcontrol-control/SBI > 0.99, PNMDA-NMDA/SBI < 0.0001, PDHPG-DHPG/SBI < 0.0001, Pcontrol-NMDA < 0.0001, Pcontrol-DHPG < 0.0001, Pcontrol/SBI-NMDA/SBI = 0.9764, Pcontrol/SBI-DHPG/SBI = 0.6286). Panel e, GluA2: $F(5,36)=11.73$, $P < 0.0001$. (Tukey's test Pcontrol-control/SBI = 0.9179, PNMDA-NMDA/SBI = 0.0001, PDHPG-DHPG/SBI = 0.0002, Pcontrol-NMDA = 0.0099, Pcontrol-DHPG = 0.0323, Pcontrol/SBI-NMDA/SBI = 0.9959, Pcontrol/SBI-DHPG/SBI = 0.9407). f (N = 7 independent experiments) PSD95: $F(5,36) = 10.93$, $P < 0.0001$. (Tukey's test Pcontrol/scr-control/atg5 = 0.7927, PNMDA/scr-NMDA/atg5 = 0.0045, PDHPG/scr-DHPG/atg5 = 0.0003, Pcontrol/scr-NMDA/scr = 0.0134, Pcontrol/scr-DHPG/scr = 0.0030, Pcontrol/atg5-NMDA/atg5 = 0.9488, Pcontrol/atg5-DHPG/atg5 = 0.9976). GluA2: $F(5,36) = 10.79$, $P < 0.0001$. (Tukey's test Pcontrol/scr-control/atg5 > 0.99, PNMDA/scr-NMDA/atg5 = 0,0001, PDHPG/scr-DHPG/atg5 = 0.0019, Pcontrol/scr-NMDA/scr = 0.0134, Pcontrol/scr-DHPG/scr = 0.0021, Pcontrol/atg5-NMDA/atg5 = 0.5844, Pcontrol/atg5-DHPG/atg5 > 0.99). Image collected and cropped by CiteAb from the following open publication (https://pubmed.ncbi.nlm.nih.gov/35115539), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Immunocytochemistry/ Immunofluorescence: NCOR1 Antibody (7A7A9) - BSA Free [NBP1-28863] - Expressive analysis of LAMP-2A & N-CoR in clinical samples. a mRNA levels of LAMP-2A & N-CoR were measured by qRT-PCR. b Protein levels of LAMP-2A & N-CoR were measured by western blot. LAMP-2A mRNA & protein levels were significantly increased in GBM center (n = 8) in comparison with peri-tumor edema zone (PTEZ, n = 8) ($p < 0.0001$), while increasing trend was observed as compared with low grade glioma (LGG, n = 8). The protein level of N-CoR, but not mRNA level was significantly decreased in GBM center as compared with PTEZ ($p < 0.0001$). Linear regression analysis incorporating data from LGG, GBM center & PTEZ revealed moderate negative correlation between protein expression of LAMP-2A & that of N-CoR ($r = -0.6001$, $p = 0.0019$). c Immunohistochemistry (IHC) analysis of LAMP-2A & N-CoR (brown signal) in glioma clinical samples. Nucleus (blue signal) was stained with hematoxylin; D. immunofluorescence (IF) analysis of LAMP-2A (green signal) & N-CoR (red signal) in glioma clinical samples. DNA (blue signal) was stained with DAPI. Both IHC & IF studies displayed upregulation of LAMP-2A & downregulation of N-CoR in GBM centers. The data are mean ± SEM from 8 tissue specimens as a group. mRNA or protein levels are expressed relative to LGG set as 1. Significant changes are set as $p < 0.05$ & represented by asterisk (One-Way ANOVA; Bonferroni's test) Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/33761934), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



Immunocytochemistry/ Immunofluorescence: NCOR1 Antibody (7A7A9) - BSA Free [NBP1-28863] - Expressive analysis of LAMP-2A & N-CoR in clinical samples. a mRNA levels of LAMP-2A & N-CoR were measured by qRT-PCR. b Protein levels of LAMP-2A & N-CoR were measured by western blot. LAMP-2A mRNA & protein levels were significantly increased in GBM center (n = 8) in comparison with peri-tumor edema zone (PTEZ, n = 8) ($p < 0.0001$), while increasing trend was observed as compared with low grade glioma (LGG, n = 8). The protein level of N-CoR, but not mRNA level was significantly decreased in GBM center as compared with PTEZ ($p < 0.0001$). Linear regression analysis incorporating data from LGG, GBM center & PTEZ revealed moderate negative correlation between protein expression of LAMP-2A & that of N-CoR ($r = -0.6001$, $p = 0.0019$). c Immunohistochemistry (IHC) analysis of LAMP-2A & N-CoR (brown signal) in glioma clinical samples. Nucleus (blue signal) was stained with hematoxylin; D. immunofluorescence (IF) analysis of LAMP-2A (green signal) & N-CoR (red signal) in glioma clinical samples. DNA (blue signal) was stained with DAPI. Both IHC & IF studies displayed upregulation of LAMP-2A & downregulation of N-CoR in GBM centers. The data are mean ± SEM from 8 tissue specimens as a group. mRNA or protein levels are expressed relative to LGG set as 1. Significant changes are set as $p < 0.05$ & represented by asterisk (One-Way ANOVA; Bonferroni's test) Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/33761934), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



Publications

Wang, Y, Zhang, B Et al. Discovery of LAMP-2A as potential biomarkers for glioblastoma development by modulating apoptosis through N-CoR degradation. Cell Commun Signal 2021-03-24 [PMID: 33761934]

Nomura A, Yokoe S, Tomoda K et al. Fluctuation in O-GlcNAcylation inactivates STIM1 to reduce store-operated calcium ion entry via downregulation of Ser621 phosphorylation J Biol Chem 2020-10-06 [PMID: 33023909] (IF/IHC, Human)

Tan J, Zhang S, Li L et al. Abnormal localized DLK1 interacts with NCOR1 in non-small cell lung cancer cell nuclear Biosci. Rep. 2019-10-29 [PMID: 31661545] (ICC/IF, Human)

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NB720-B	Rabbit anti-Mouse IgG (H+L) Secondary Antibody [Biotin]
NBP1-97005-0.5mg	Mouse IgG1 Isotype Control (MG1)
H00009611-Q01-10ug	Recombinant Human NCOR1 GST (N-Term) Protein

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This product is for research use only and is not approved for use in humans or in clinical diagnosis. Primary Antibodies are guaranteed for 1 year from date of receipt.

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