

Product Datasheet

c-Myc Antibody - BSA Free

NB600-336SS

Unit Size: 0.025 ml

Store at 4C short term. Aliquot and store at -20C long term. Avoid freeze-thaw cycles.

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NB600-336SS

c-Myc Antibody - BSA Free

Product Information

Unit Size	0.025 ml
Concentration	1 mg/ml
Storage	Store at 4C short term. Aliquot and store at -20C long term. Avoid freeze-thaw cycles.
Clonality	Polyclonal
Preservative	0.09% Sodium Azide
Isotype	IgG
Purity	Immunogen affinity purified
Buffer	Tris-Glycine, 0.15 M NaCl
Target Molecular Weight	48.8 kDa

Product Description

Host	Rabbit
Gene ID	4609
Gene Symbol	MYC
Species	Human, Mouse, Insect, S. pombe
Reactivity Notes	Insect reactivity reported in scientific literature (Martinez-Velez N et al). Use in S. pombe reported in scientific literature (PMID:18094683).
Immunogen	A synthetic peptide made to the human c-Myc Antibody (between residues 385-435) [Uniprot: P01106]

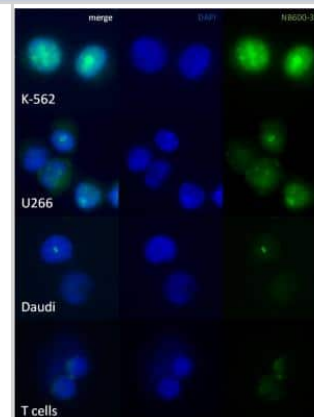
Product Application Details

Applications	Western Blot, Simple Western, ELISA, Immunocytochemistry/Immunofluorescence, Immunohistochemistry, Immunohistochemistry-Paraffin, Immunoprecipitation, Chromatin Immunoprecipitation (ChIP)
Recommended Dilutions	Western Blot 1:1000, Simple Western 1:500, ELISA 1:1000, Immunohistochemistry 1:100, Immunocytochemistry/Immunofluorescence 1:50, Immunoprecipitation 1:1000, Immunohistochemistry-Paraffin 1:100, Chromatin Immunoprecipitation (ChIP)
Application Notes	Prior to immunostaining paraffin tissues, antigen retrieval with sodium citrate buffer (pH 6.0) is recommended. In Simple Western only 10 - 15 uL of the recommended dilution is used per data point. See Simple Western Antibody Database for Simple Western validation: Tested in Jurkat lysate 0.05 mg/mL, separated by Size, antibody dilution of 1:500, apparent MW was 87 kDa. Separated by Size-Wes, Sally Sue/Peggy Sue.

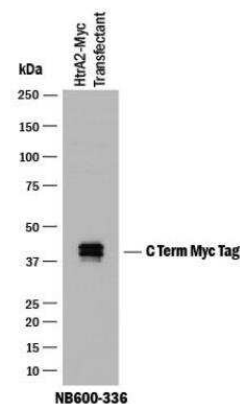


Images

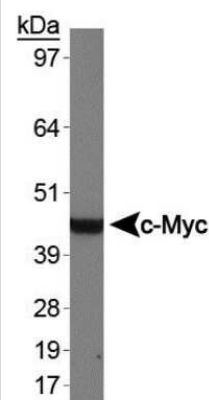
Immunocytochemistry/Immunofluorescence: c-Myc Antibody [NB600-336] - Staining of K-562, U266, Daudi and normal T cells with c-Myc antibody. ICC/IF image submitted by a verified customer review.



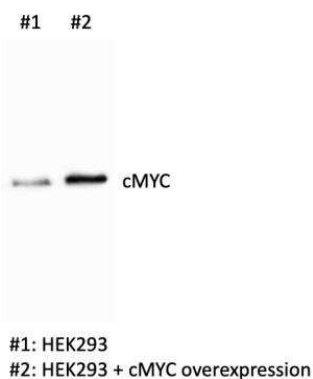
Western Blot: c-Myc Antibody [NB600-336] - Detection of c-Myc in HtrA2-Myc Transfectant lysates .



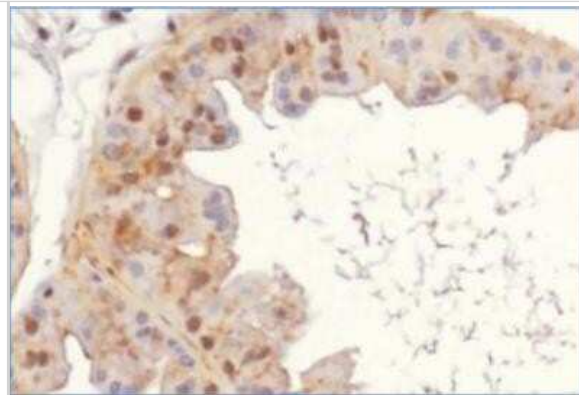
Western Blot: c-Myc Antibody [NB600-336] - Analysis of c-Myc on Jurkat whole cell extract using NB600-336.



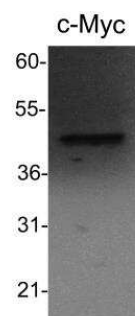
Western Blot: c-Myc Antibody [NB600-336] - Lane 1: control HEK293 cell lysate. Lane 2: CMYC over-expression in HEK293 cell lysate. WB image submitted by a verified customer review.



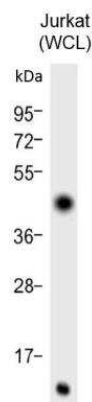
Immunohistochemistry: c-Myc Antibody [NB600-336] - Analysis of c-Myc in mouse prostate using DAB with hematoxylin counterstain.



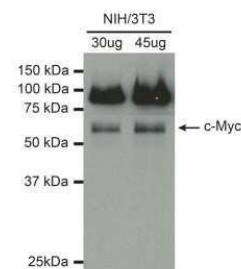
Western Blot: c-Myc Antibody [NB600-336] - c-Myc in MOLT-4 cells. WB image submitted by a verified customer review.



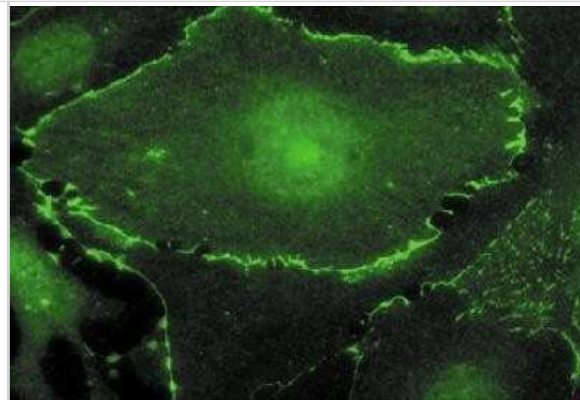
Western Blot: c-Myc Antibody [NB600-336] - Analysis of whole cell lysate (WCL) of Jurkat cells using anti-cMyc antibody (NB600-336) at 1:1000 dilution. HRP conjugated goat anti-rabbit IgG (H+L) cross adsorbed secondary antibody was used with ECL substrate for the detection of c-Myc antibody bound to the blotted protein. This c-Myc antibody detected the c-Myc specific band at its expected position (48-50kDa). The signal below 10 kDa in this blot is potentially the degraded protein and we have not characterized this band.



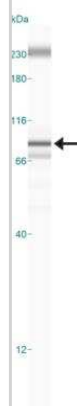
Western Blot: c-Myc Antibody [NB600-336] - Analysis using the HRP conjugate of NB600-336. Detection of c-Myc in NIH/3T3 cell lysates (30ug and 45ug per lane) using anti-c-Myc antibody. WB image submitted by a verified customer review.



Immunocytochemistry/Immunofluorescence: c-Myc Antibody [NB600-336] - Detection of c-myc Tagged Plakoglobin by Immunofluorescence. Samples: Human microvascular endothelial cells expressing c-myc tagged plakoglobin following transient transfection.



Simple Western: c-Myc Antibody [NB600-336] - Lane view shows a specific band for c-Myc in 0.05 mg/ml of Jurkat lysate. This experiment was performed under reducing conditions using the 12-230 kDa separation system.



Publications

T Shibata, DY Cao, TB Dar, F Ahmed, SA Bhat, LC Veiras, EA Bernstein, AA Khan, M Chaum, SL Shiao, WG Tourtellot, JF Giani, KE Bernstein, X Cui, E Vail, Z Khan miR766-3p and miR124-3p Dictate Drug Resistance and Clinical Outcome in HNSCC Cancers, 2022-10-27;14(21):. 2022-10-27 [PMID: 36358691]

Wells JR, Padua MB, Haaning AM et al. Non-coding cause of congenital heart defects: Abnormal RNA splicing with multiple isoforms as a mechanism for heterotaxy Human Genetics and Genomics Advances 2024-09-12 [PMID: 39275801]

Filidou E, Kandilogiannakis L, Tarapatzi G et al. A Simplified and Effective Approach for the Isolation of Small Pluripotent Stem Cells Derived from Human Peripheral Blood Biomedicines 2023-03-05 [PMID: 36979766] (Immunocytochemistry/ Immunofluorescence, Human)

Liang JH, Akhanov V, Ho A et al. Dopamine signaling from ganglion cells directs layer-specific angiogenesis in the retina Current biology : CB 2023-08-07 [PMID: 37572663]

Xiong Y, Wang L, Xu S et al. Small molecule Z363 co-regulates TAF10 and MYC via the E3 ligase TRIP12 to suppress tumour growth Clinical and Translational Medicine 2023-01-13 [PMID: 36639831] (Immunoprecipitation, Western Blot)

Park ZM, Sporer AJ, Kraft K et al. Kar4, the yeast homolog of METTL14, is required for mRNA m6A methylation and meiosis PLOS Genetics 2023-08-21 [PMID: 37603553] (Western Blot)

Park ZM, Sporer A, Kraft K et al. Kar4, the Yeast Homolog of METTL14, is Required for mRNA m (6) A Methylation and Meiosis bioRxiv 2023-08-16 [PMID: 36747717] (Western Blot)

Park ZM It Takes Two to Tetrad: Dissecting the Role of Kar4 and Vir1 in Meiosis and mRNA m6 A Methylation Thesis 2023-01-01 (WB)

Park ZM, Belnap E, Remillard M, Rose MD Vir1p, the Yeast Homolog of Virilizer, is Required for mRNA m 6 A Methylation and Meiosis bioRxiv : the preprint server for biology 2023-02-07 [PMID: 36798303] (Western Blot)

Park Z, Sporer A, Kraft K et al. Kar4, the Yeast Homolog of METTL14, is Required for mRNA m6A Methylation and Meiosis bioRxiv 2023-01-30

Alzaydi MM, Abdul-Salam VB, Whitwell HJ et al. Intracellular Chloride Channels Regulate Endothelial Metabolic Reprogramming in Pulmonary Arterial Hypertension American journal of respiratory cell and molecular biology 2022-10-20 [PMID: 36264759]

Martinez-Velez N, Garcia-Moure M, Marigil M et al. Blood feeding activates the vitellogenic stage of oogenesis in the mosquito Aedes aegypti through inhibition of glycogen synthase kinase 3 by the insulin and TOR pathways Dev Biol 2019-06-03 [PMID: 31153832]

More publications at <http://www.novusbio.com/NB600-336>



Procedures

Serum protocol for c-Myc Antibody (NB600-336)

Western Blot Protocol

1. Perform SDS-PAGE on samples to be analyzed, loading 40 ug of total protein per lane.
 2. Transfer proteins to membrane according to the instructions provided by the manufacturer of the membrane and transfer apparatus.
 3. Stain according to standard Ponceau S procedure (or similar product) to assess transfer success, and mark molecular weight standards where appropriate.
 4. Rinse the blot.
 5. Block the membrane using standard blocking buffer for at least 1 hour.
 6. Wash the membrane in wash buffer three times for 10 minutes each.
 7. Dilute primary antibody in blocking buffer and incubate 1 hour at room temperature.
 8. Wash the membrane in wash buffer three times for 10 minutes each.
 9. Apply the diluted HRP conjugated secondary antibody in blocking buffer (as per manufacturers instructions) and incubate 1 hour at room temperature.
 10. Wash the blot in wash buffer three times for 10 minutes each (this step can be repeated as required to reduce background).
 11. Apply the detection reagent of choice in accordance with the manufacturers instructions.
- Note: Tween-20 can be added to the blocking or antibody dilution buffer at a final concentration of 0.05-0.2%.

Immunohistochemistry-Paraffin Embedded Sections

Antigen Unmasking:

Bring slides to a boil in 10 mM sodium citrate buffer (pH 6.0) then maintain at a sub-boiling temperature for 10 minutes. Cool slides on bench-top for 30 minutes.

Staining:

1. Wash sections in deionized water three times for 5 minutes each.
2. Wash sections in wash buffer for 5 minutes.
3. Block each section with 100-400 ul blocking solution for 1 hour at room temperature.
4. Remove blocking solution and add 100-400 ul diluted primary antibody. Incubate overnight at 4 C.
5. Remove antibody solution and wash sections in wash buffer three times for 5 minutes each.
6. Add 100-400 ul biotinylated diluted secondary antibody. Incubate 30 minutes at room temperature.
7. Remove secondary antibody solution and wash sections three times with wash buffer for 5 minutes each.
8. Add 100-400 ul Streptavidin-HRP reagent to each section and incubate for 30 minutes at room temperature.
9. Wash sections three times in wash buffer for 5 minutes each.
10. Add 100-400 ul DAB substrate to each section and monitor staining closely.
11. As soon as the sections develop, immerse slides in deionized water.
12. Counterstain sections in hematoxylin.
13. Wash sections in deionized water two times for 5 minutes each.
14. Dehydrate sections.
15. Mount coverslips.

Immunocytochemistry Protocol

Culture cells to appropriate density in 35 mm culture dishes or 6-well plates.

1. Remove culture medium and add 10% formalin to the dish. Fix at room temperature for 30 minutes.
2. Remove the formalin and add ice cold methanol. Incubate for 5-10 minutes.
3. Remove methanol and add washing solution (i.e. PBS). Be sure to not let the specimen dry out. Wash three times for 10 minutes.
4. To block nonspecific antibody binding incubate in 10% normal goat serum from 1 hour to overnight at room temperature.
5. Add primary antibody at appropriate dilution and incubate at room temperature from 2 hours to overnight at room



temperature.

6. Remove primary antibody and replace with washing solution. Wash three times for 10 minutes.
7. Add secondary antibody at appropriate dilution. Incubate for 1 hour at room temperature.
8. Remove antibody and replace with wash solution, then wash for 10 minutes. Add Hoechst 33258 to wash solution at 1:25,000 and incubate for 10 minutes. Wash a third time for 10 minutes.
9. Cells can be viewed directly after washing. The plates can also be stored in PBS containing Azide covered in Parafilm (TM). Cells can also be cover-slipped using Fluoromount, with appropriate sealing.

*The above information is only intended as a guide. The researcher should determine what protocol best meets their needs. Please follow safe laboratory procedures.



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