

# Product Datasheet

## c-Myc Antibody (Y69) NB110-57172

Unit Size: 0.1 ml

Aliquot and store at -20C or -80C. Avoid freeze-thaw cycles.

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**NB110-57172**

c-Myc Antibody (Y69)

<b>Product Information</b>	
<b>Unit Size</b>	0.1 ml
<b>Concentration</b>	Please see the vial label for concentration. If unlisted please contact technical services.
<b>Storage</b>	Aliquot and store at -20C or -80C. Avoid freeze-thaw cycles.
<b>Clonality</b>	Monoclonal
<b>Clone</b>	Y69
<b>Preservative</b>	0.01% Sodium Azide
<b>Isotype</b>	IgG
<b>Purity</b>	Protein A or G purified
<b>Buffer</b>	49% PBS, 0.05% BSA and 50% Glycerol
<b>Product Description</b>	
<b>Host</b>	Rabbit
<b>Gene ID</b>	4609
<b>Gene Symbol</b>	MYC
<b>Species</b>	Human, Mouse, Rat
<b>Reactivity Notes</b>	Reacts with: Mouse, Rat, Human
<b>Specificity/Sensitivity</b>	A synthetic peptide corresponding to residues in N-terminus of human c-Myc was used as immunogen.
<b>Immunogen</b>	Synthetic peptide corresponding to residues in the N terminus of Human c-Myc
<b>Notes</b>	Produced using Abcam's RabMab® technology. RabMab® technology is covered by the following U.S. Patents, No. 5,675,063 and/or 7,429,487.
<b>Product Application Details</b>	
<b>Applications</b>	Western Blot, Simple Western, Immunocytochemistry/ Immunofluorescence, Immunohistochemistry, Immunohistochemistry-Paraffin, Immunoprecipitation
<b>Recommended Dilutions</b>	Western Blot 1:1000 - 10000, Simple Western, Immunohistochemistry, Immunocytochemistry/ Immunofluorescence 1:250 - 500, Immunoprecipitation 1:150, Immunohistochemistry-Paraffin
<b>Application Notes</b>	This antibody works in immunohistochemistry-Paraffin, immunoprecipitation, Western Blot and Immunocytochemistry



## Procedures

### Protocol specific for c-Myc Antibody (NB110-57172)

Immunohistochemistry Protocol for Paraffin-embedded Tissues

**1. Solutions and reagents**

**1.1.** Xylene

**1.2.** Ethanol, anhydrous denatured, histological grade (100%, 95%, 70%)

**1.3.** Washing buffer: TBST washing buffer: 1XTBS/0.1% Tween-20

To prepare stock solution of 10X TBS: add 24.2 g Trizma base and 80 g sodium chloride to 1L of dH<sub>2</sub>O. Adjust pH to 7.6.

Working solution. 1XTBST/0.1% Tween-20: add 100ml 10XTBS to 900 ml dH<sub>2</sub>O. Add 1 ml Tween-20 and mix well.

**1.4.** Distilled water (dH<sub>2</sub>O)

**1.5.** Antigen Retrieval Solution:

0.01M Sodium Citrate Buffer, pH 6.0

To prepare stock solutions:

**Solution A.** 0.1 M citric acid solution: dissolve 21.0 g of citric acid, monohydrate (C<sub>6</sub>H<sub>8</sub>O<sub>7</sub>.H<sub>2</sub>O) in 1 liter of dH<sub>2</sub>O.

**Solution B.** 0.1M sodium citrate solution: dissolve 29.4 g trisodium citrate dihydrate (C<sub>6</sub>H<sub>5</sub>Na<sub>3</sub>O<sub>7</sub>.2H<sub>2</sub>O) in 1 liter of dH<sub>2</sub>O.

Working solution: Add 9 ml of Stock solution A and 41 ml of stock solution B to 450 ml of dH<sub>2</sub>O. Adjust pH to 6.0.

**1.6.** 3% Hydrogene Peroxide

**1.7.** Blocking buffer: PBS (Dulbeccos Phosphate Buffered Salts, 1X, catalog #21-031-CV from Mediatech, Inc.) + 10% serum (serum origin depends on the host of the secondary antibody)

**1.8.** Hematoxylin QS (catalog #H-3404 from Vector Laboratories, Inc.)

**1.9.** Permanent Mounting medium (VectaMount, catalog# H-5000 Vector Laboratories, Inc.)

**2. Protocol**

**2.1.** Deparaffinization/Rehydration

**2.1.1.** Heat slides in an oven at 65C for 1 hour.

**2.1.2.** De-paraffinize/hydrate using the following series of washes: two Xylene washes (5 min each), followed by two 100% ethanol rinses (5 min each), followed by 95% ethanol, 70% ethanol, 50% ethanol, 30% ethanol, followed by H<sub>2</sub>O and a TBST wash for 5 min on a shaker.

**2.2.** Antigen Retrieval

**2.2.1.** Immerse slides into staining dish containing Antigen Retrieval Solution.

**2.2.2.** Place covered staining dish into the rice cooker. Add 120 ml of dH<sub>2</sub>O.

**2.2.3.** When cook is turned to warm (about 20 to 30 min), unplug the cooker and remove the staining dish to the bench top.

**2.2.4.** Allow to cool down, without cover, for 20 min.

**2.3.** Staining

**2.3.1.** Wash slides with TBST for 5 min on a shaker.

**2.3.2.** Inactivate endogenous peroxidase by covering tissue with 3% hydrogen peroxide for 10 min.

**2.3.3.** Wash slides three times with TBST (3 min each on a shaker).

**2.3.4.** Block slides with the blocking solution for 1 hour.

**2.3.5.** Dilute primary antibody in the blocking buffer per recommendation on the data sheet.

**2.3.6.** Apply primary antibody to each section and incubate overnight in the humidified chamber (4C).

**2.3.7.** Wash slides three times with TBST (3 min each on a shaker).

**2.3.8.** Apply to each section secondary HRP-conjugated anti-rabbit antibody diluted in the blocking solution per manufacturers recommendation; incubate for 1 hour at room temperature.

**2.3.9.** Wash slides three times with TBST (3 min each on a shaker).

**2.3.10.** Add freshly prepared DAB substrate to the sections.

**2.3.11.** Incubate tissue sections with the substrate at room temperature until suitable staining develops (generally 2 to 5 min).

**2.3.12.** Rinse sections with water.

**2.3.13.** Counterstain with Hematoxylin.

**2.3.14.** Rinse sections with water.

**2.3.15.** Dehydrate samples using two rinses with 100% Ethanol (20 dips per rinse) followed by two rinses with Xylene (30 dips per rinse).

**2.3.16.** Mount coverslips on slides using Permount medium.



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### **Limitations**

This product is for research use only and is not approved for use in humans or in clinical diagnosis. Primary Antibodies are guaranteed for 1 year from date of receipt.

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