

Product Datasheet

Histone H2AX [p Ser139] Antibody - BSA Free NB100-384

Unit Size: 0.1 ml

Store at 4C. Do not freeze.

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NB100-384

Histone H2AX [p Ser139] Antibody - BSA Free

| Product Information | |
|-------------------------|---------------------------------------|
| Unit Size | 0.1 ml |
| Concentration | 1.0 mg/ml |
| Storage | Store at 4C. Do not freeze. |
| Clonality | Polyclonal |
| Preservative | 0.09% Sodium Azide |
| Isotype | IgG |
| Purity | Immunogen affinity purified |
| Buffer | Tris-Citrate/Phosphate (pH 7.0 - 8.0) |
| Target Molecular Weight | 15 kDa |

| Product Description | |
|-------------------------|--|
| Host | Rabbit |
| Gene ID | 3014 |
| Gene Symbol | H2AX |
| Species | Human, Mouse, Rat, Canine |
| Reactivity Notes | Rat reactivity reported in scientific literature (PMID: 27102221), Canine reactivity reported in scientific literature (PMID: 23365434). |
| Marker | DNA Double-strand break marker |
| Specificity/Sensitivity | The epitope maps to a region surrounding phosphorylated serine 139 of human histone H2AX. |
| Immunogen | This Histone H2AX [p Ser139] Antibody was developed against to a region surrounding phosphorylated serine 139 of human histone H2AX [Swiss-Prot entry P16104] (GeneID 3014). |
| Notes | Licensed to Novus Biologicals LLC under U.S. Patent Nos. 6,362,317 and 6,884,873. |

| Product Application Details | |
|-----------------------------|--|
| Applications | Western Blot, Simple Western, Flow Cytometry, Immunocytochemistry/Immunofluorescence, Immunohistochemistry, Immunohistochemistry-Frozen, Immunohistochemistry-Paraffin, Chromatin Immunoprecipitation (ChIP), Knockout Validated |
| Recommended Dilutions | Western Blot 1:10000-1:25000, Simple Western 5 ug/ml, Flow Cytometry 5 ug per 1 million cells, Immunohistochemistry 1:2000 - 1:10000, Immunocytochemistry/ Immunofluorescence 1:500 to 1:5000, Immunohistochemistry-Paraffin 1:2000 - 1:10000, Immunohistochemistry-Frozen 1:1000 - 1:5000, Chromatin Immunoprecipitation (ChIP), Knockout Validated |



Application Notes

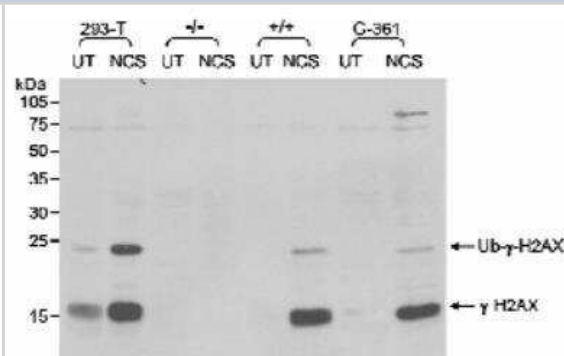
For IHC, epitope retrieval with citrate buffer pH6.0 is recommended for FFPE tissue sections. Formaldehyde fixation is recommended. Permeabilization with Triton-X 100 is recommended for formaldehydefixed cells. Immunoprecipitation is not recommended.

In Simple Western only 10 - 15 uL of the recommended dilution is used per data point.

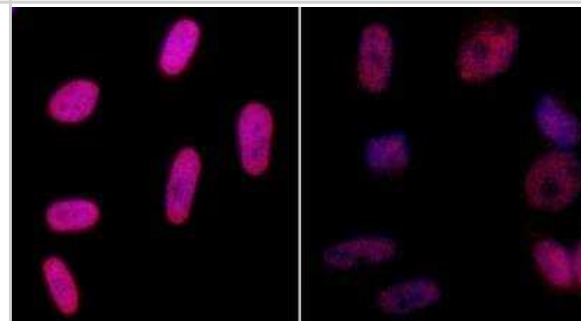
See [Simple Western Antibody Database](#) for Simple Western validation: Tested in Jurkat lysate, separated by Size, antibody dilution of 5 ug/mL, apparent MW was 29 kDa. Separated by Size-Wes, Sally Sue/Peggy Sue. Use in chromatin immunoprecipitation reported in scientific literature (PMID: 30049290).

Images

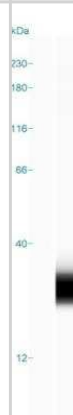
Detection of Human and Mouse Histone H2AX [p Ser139] by Western Blot. Samples: Nuclear extract (50 ug) from human HEK293, human melanoma (G361), mouse wildtype embryonic fibroblasts (+/+) or mouse H2AX knockout embryonic fibroblasts (-/-). Antibody: Affinity purified rabbit Histone H2AX [p Ser139] antibody NB100-384 used at 0.1 ug/ml. Detection: Chemiluminescence with 30 second exposure. (NCS, neocarzinostatin - 200 ng/ml, 30 min). Bands appear at an observed molecular weight of ~15 kDa.



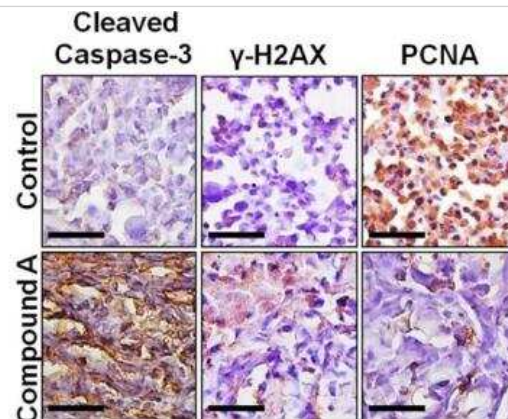
Samples: Neocarzinostatin treated asynchronous HeLa cells (left) and untreated asynchronous HeLa cells (right). Antibody: Affinity purified rabbit Histone H2AX [p Ser139] used at a dilution of 1:5,000 (0.2ug/ml). Detection: Red fluorescent Anti-rabbit IgG-DyLight 594 used at a dilution of 1:100.



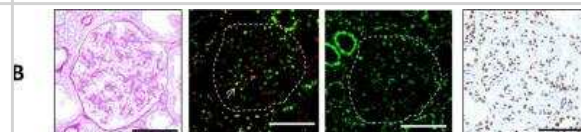
Simple Western lane view shows a specific band for Histone H2AX [p Ser139] in 0.2 mg/ml of Jurkat lysate(s). This experiment was performed under reducing conditions using the 12 - 230 kDa separation system.



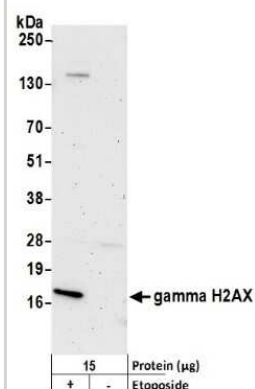
Immunocompromised mice were subcutaneously injected with cancerous cell lines and tumors were allowed to establish. Treatments occurred every other day and the studied compound or the equivalent vehicle control administered intraperitoneally for five weeks. Tumor volume and mass were measured two times per week. IHC analysis of sectioned tumor tissues from the MDA-MB-231 study. Each section was subjected to the specified antibody followed by a biotinylated secondary antibody. Detection was done using a DAB Peroxidase HRP Substrate Kit (brown) followed by Hematoxylin counterstaining (purple). Images were obtained using inverted bright field microscopy. Sectioning results are representative of three individual tumors. Scale bar is 50 microns. Image collected and cropped by CiteAb from the following publication ([nature.com/articles/s41598-017-01230-4](https://www.nature.com/articles/s41598-017-01230-4)), licensed under a CC-BY license.



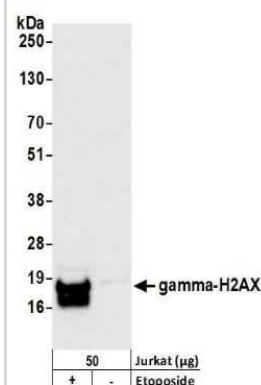
Immunostaining of Histone H2AX [p Ser139] WT1 and 5mC in patients with IgA nephropathy and controls. Examples of PAS staining and immunostaining with Histone H2AX [p Ser139] (green) and WT1 (red), pATM and 5mC in glomeruli of IgA nephropathy and controls. A kidney sample of a 65-year-old male of IgA nephropathy without podocytopathic features. Arrows indicate Histone H2AX [p Ser139] and WT1 double-positive cells. Scale bars: 50um. Image collected and cropped by CiteAb from the following publication (<https://www.nature.com/articles/s41598-019-57140-0>) licensed under a CC-BY license.



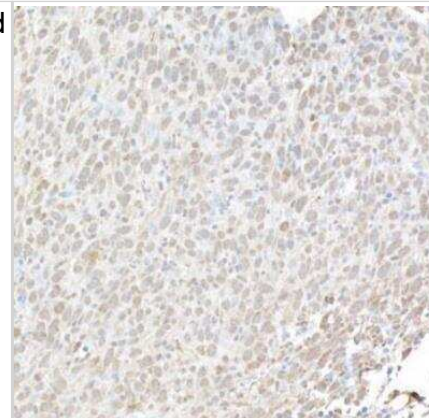
Samples: Nuclear extract from HeLa cells treated with 100 uM EPE for 4 hours (+) or mock treated (-). Antibody: Affinity purified rabbit Histone H2AX [p Ser139] antibody used at 0.1 ug/ml. Detection: Chemiluminescence with an exposure time of 3 minutes. Band appears at an observed molecular weight of ~17 kDa.



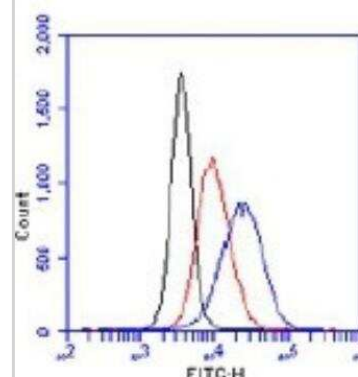
Detection of human Histone H2AX [p Ser139] by western blot. Samples: Whole cell lysate (50 ug) from Jurkat cells treated with 100 uM EPE for 4 hours (+) or mock treated (-). Antibody: Affinity purified rabbit Histone H2AX [p Ser139] antibody NB100-384 used for WB at 0.1 ug/ml. Detection: Chemiluminescence with an exposure time of 3 seconds. Band appears at an observed molecular weight of ~18 kDa.



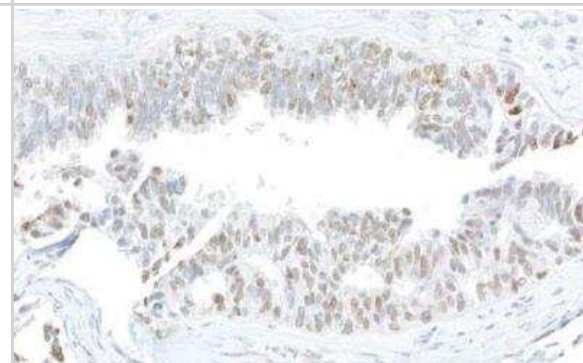
FFPE section of mouse CT26 colon carcinoma. Antibody: Affinity purified rabbit Histone H2AX [p Ser139] antibody used at a dilution of 1:1,000 (1 ug/ml). Detection: DAB.



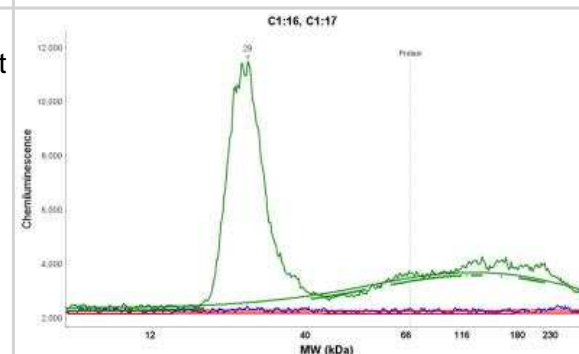
Analysis of Histone H2AX [p Ser139] in EPE Treated Jurkat Cells. Cells were treated for 3 hrs in 5ug/ml etoposide, fixed in 1.5% PFA, and permeabilized in 90% Methanol. 1 million cells were stained with 0.5 ug anti-KLH or anti-H2AX NB100-384 and secondary FITC-conjugated goat anti-rabbit (in a 150ul reaction). Black- etoposide treated, anti-KLH; Red- untreated, anti-Histone H2AX [p Ser139]; Blue- etoposide treated , anti-Histone H2AX [p Ser139].



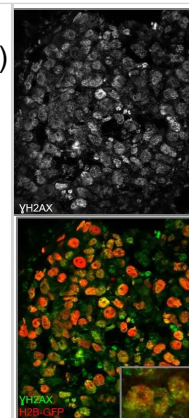
FFPE section of human ovarian carcinoma. Antibody: Affinity purified rabbit Histone H2AX [p Ser139] antibody used at a dilution of 1:5,000 (0.2 ug/ml). Detection: DAB.



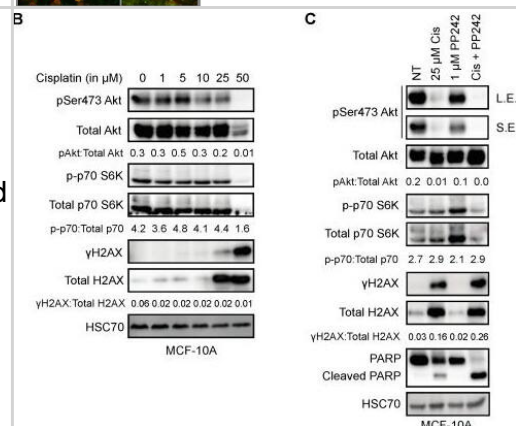
Electropherogram image(s) of corresponding Simple Western lane view. Histone H2AX [p Ser139] antibody was used at 5 ug/ml dilution on Jurkat lysate(s).



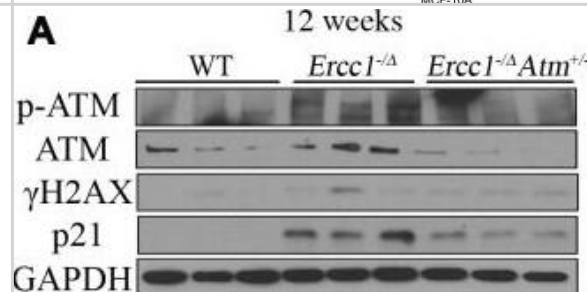
Immunohistochemistry: Rabbit Polyclonal Histone H2AX [p Ser139] Antibody - Histone H2AX Antibody on mouse cancer tissue. H2AX (Gray) and H2BGFP(Green). Primary antibody dilution: 1:1000 in a 10um slice.



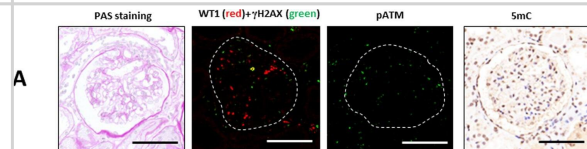
mTORC1/2 activity prevents Cisplatin-induced cell death in MCF-10A cells. (A) Western blot displaying effects on mTOR signaling during a dose escalation of PP242 treatment in MCF-10A cells; (B) Western blot displaying effects of mTOR signaling on a dose escalation of cisplatin treatment in MCF-10A cells; (C) Western blot displaying effects on mTOR signaling and cell death during non-treated, Cisplatin, PP242, and Cisplatin + PP242-treated MCF-10A cells.



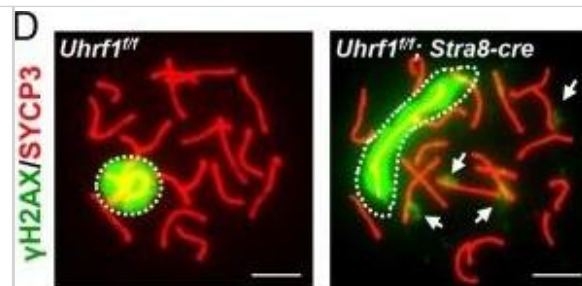
Western Blot: Histone H2AX [p Ser139] Antibody [NB100-384] - ATM & NF-κB activation are downregulated in Ercc1-Δ mice heterozygous for Atm. (A) Livers were collected at 12 weeks of age from WT, Ercc1-Δ & Ercc1-ΔAtm+/- mic (n=3 per genotype) & lysates analyzed by western blot for activation of ATM & its downstream effectors. (B) Same liver lysates were used to measure phosphorylation of p65 & IκBα. (C) Western blot analysis of livers from 16-week-old WT, Ercc1-Δ & Ercc1-ΔAtm+/-mice (n=3 per genotype) probed for activation of ATM. GAPDH was used as a loading control. (D) Same liver lysates used to measure activation of NF-κB. (E) Fourteen-week-old livers from Ercc1-Δ & Ercc1-Δp65+/- mice (n=3 per genotype) were analyzed by western blot for activation of ATM (F) & NF-κB. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/32201398>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



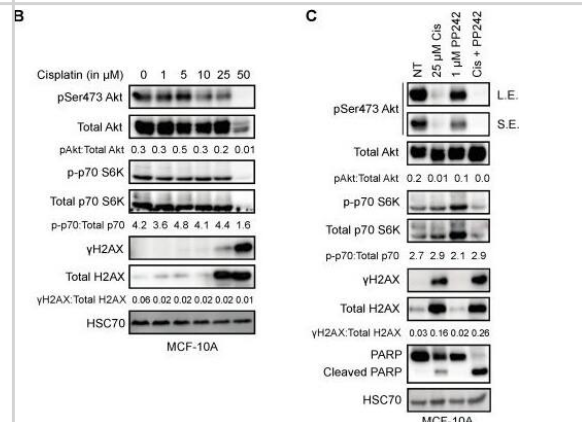
Immunocytochemistry/ Immunofluorescence: Histone H2AX [p Ser139] Antibody [NB100-384] - Immunostaining of γH2AX, WT1 & 5mC in patients with IgA nephropathy & controls. Examples of PAS staining & immunostaining with γH2AX (green) & WT1 (red), pATM & 5mC in glomeruli of IgA nephropathy & controls. (A) A control kidney sample of 44-year-old female, (B) 65-year-old male of IgA nephropathy without podocytopathic features & (C) 55-year-old male of IgA nephropathy with podocytopathic features. Arrows indicate γH2AX & WT1 double-positive cells. Scale bars: 50 μm. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/31937846>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



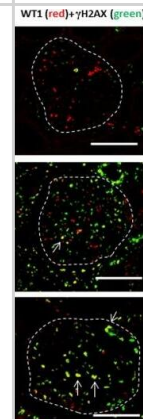
Immunocytochemistry/ Immunofluorescence: Histone H2AX [p Ser139] Antibody [NB100-384] - UHRF1 deficiency resulted in impaired meiotic recombination & defective pachynema. a Double immunofluorescence of SYCP3 (green) & DMC1 (red) in testicular spread preparations. b, c The number of DMC1 foci in zygotene stage (b) & pachytene stage (c). d Immunostaining for SYCP3 (red) & γ H2AX (green). e The percentage of abnormal γ H2AX foci in the pachytene stage. f Immunostaining for SYCP3 (red) & MLH1 (green). g The number of MLH1 foci in pachynema. h Immunostaining for SYCP3 (red) & H1t (green). i The percentage of spermatocytes with H1T staining. *** $p \leq 0.001$; * $p \leq 0.05$. Scale bar, 5 μ m in a, d, f, h. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/32081844>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



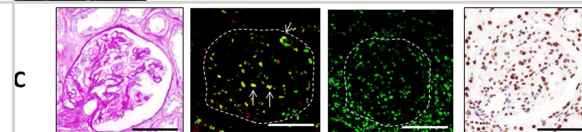
Western Blot: Histone H2AX [p Ser139] Antibody [NB100-384] - mTORC1/2 activity prevents Cisplatin-induced cell death in MCF-10A cells. (A) Western blot displaying effects on mTOR signaling during a dose escalation of PP242 treatment in MCF-10A cells; (B) Western blot displaying effects of mTOR signaling on a dose escalation of cisplatin treatment in MCF-10A cells; (C) Western blot displaying effects on mTOR signaling & cell death during non-treated, Cisplatin, PP242, & Cisplatin + PP242-treated MCF-10A cells. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/31771139>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



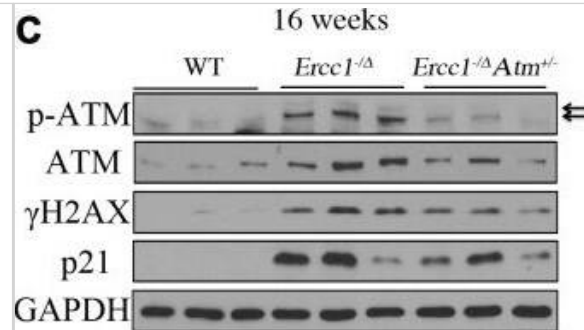
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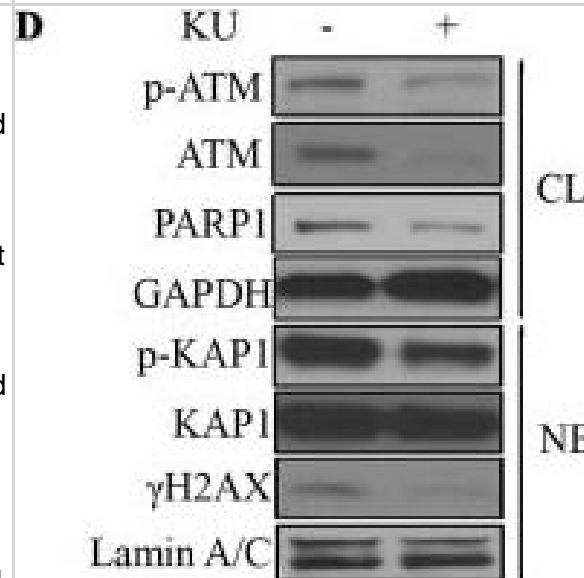
Immunocytochemistry/ Immunofluorescence: Histone H2AX [p Ser139] Antibody [NB100-384] - Immunostaining of γ H2AX, WT1 & 5mC in patients with IgA nephropathy & controls. Examples of PAS staining & immunostaining with γ H2AX (green) & WT1 (red), pATM & 5mC in glomeruli of IgA nephropathy & controls. (A) A control kidney sample of 44-year-old female, (B) 65-year-old male of IgA nephropathy without podocytopathic features & (C) 55-year-old male of IgA nephropathy with podocytopathic features. Arrows indicate γ H2AX & WT1 double-positive cells. Scale bars: 50 μ m. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/31937846>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



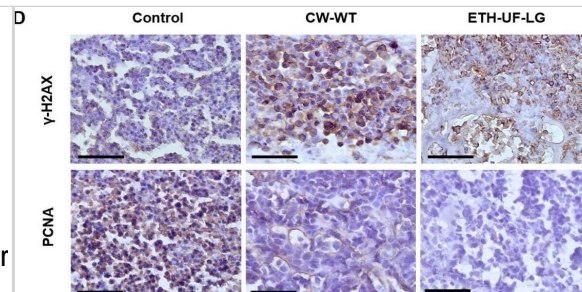
Western Blot: Histone H2AX [p Ser139] Antibody [NB100-384] - ATM & NF- κ B activation are downregulated in *Ercc1*- Δ mice heterozygous for *Atm*. (A) Livers were collected at 12 weeks of age from WT, *Ercc1*- Δ & *Ercc1*- Δ *Atm*^{+/+} mice (n=3 per genotype) & lysates analyzed by western blot for activation of ATM & its downstream effectors. (B) Same liver lysates were used to measure phosphorylation of p65 & I κ B α . (C) Western blot analysis of livers from 16-week-old WT, *Ercc1*- Δ & *Ercc1*- Δ *Atm*^{+/+} mice (n=3 per genotype) probed for activation of ATM. GAPDH was used as a loading control. (D) Same liver lysates used to measure activation of NF- κ B. (E) Fourteen-week-old livers from *Ercc1*- Δ & *Ercc1*- Δ *p65*^{+/+} mice (n=3 per genotype) were analyzed by western blot for activation of ATM (F) & NF- κ B. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/32201398>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



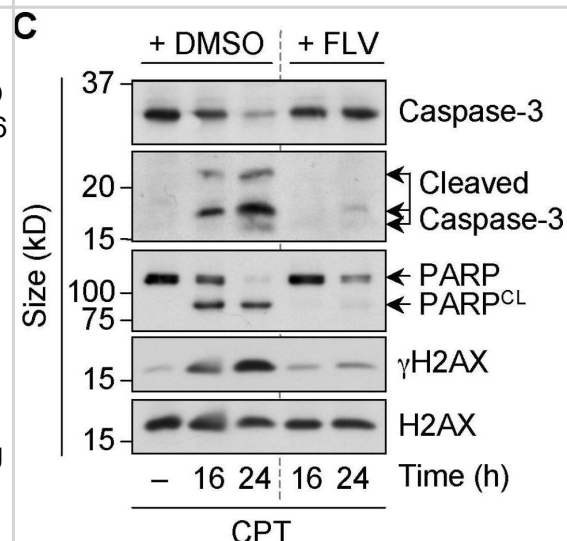
Western Blot: Histone H2AX [p Ser139] Antibody [NB100-384] - Pharmacologic inhibition of ATM rescues oxidative stress-induced senescence by suppressing ATM- & NEMO-mediated NF- κ B activation. (A) Representative images of primary WT & *Ercc1*^{-/-} MEFs were induced to undergo senescence by serial passaging at 20% oxygen. At passage 5, MEFs were grown in the presence or absence of KU-55933 (10 μ M) for 72 hrs. Senescence was determined by SA- β gal staining. Images were obtained at the magnification of 10x. (B) Quantitation of the percent SA- β gal positive cells. Graph represents the mean \pm s.e.m. of three independent experiments. Student's t-test, ***p < 0.001, ****p < 0.0001. (C) Passage 5 *Ercc1*^{-/-} MEFs treated with vehicle or KU-55933 (10 μ M) for 72 hours were collected & levels of p21 & p16INK4a were determined by western blotting. (D) Passage 5 *Ercc1*^{-/-} MEFs were treated with KU-55933 (10 μ M) for 72 hours & whole cell lysate (CL) & nuclear extracts (NE) were analyzed by immunoblotting for expression of proteins involved in the DNA damage response. (E) Whole cell lysate (CL) & nuclear extract (NE) were extracted from *Ercc1*^{-/-} MEFs treated with 10 μ M of KU-55933 for analysis of nuclear NEMO & p65. GAPDH was used as a loading control of total proteins & LaminA/C as a loading control of nuclear protein. (F) Passage 5 WT & *Ercc1*^{-/-} MEFs transfected with a NF- κ B-luciferase reporter construct were cultured in the presence or absence of KU-55933 (10 μ M) & were collected for luciferase assays after 72 hours. (G) qRT-PCR analysis of mRNA expression in passage 5 WT & *Ercc1*^{-/-} MEFs treated with or without of KU-55933 (10 μ M) for 72 hrs. P values were determined using a Student's t-test. *p < 0.05, **p < 0.01, ***p < 0.001. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/32201398>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



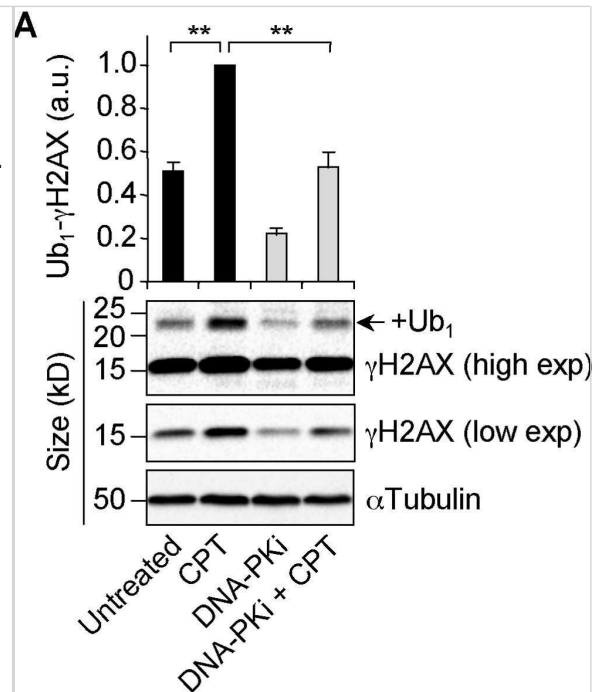
Immunohistochemistry: Histone H2AX [p Ser139] Antibody [NB100-384] - Orally administered lemongrass & white tea extract reduce tumor size in lymphoma xenograft model in immunocompromised mice. Immunocompromised mice were subcutaneously injected with cancerous cells & tumors were allowed to establish. Treatments occurred every other day & the studied compound or the equivalent vehicle control administered orally for three weeks. (A, B) The tumors were photographed before & after extraction from the animals. (C) Tumor volume & mass were measured two times per week. (D) Immunohistochemistry analysis of sectioned tumor tissues from the lymphoma study. Each section was subjected to the specified antibody followed by a biotinylated secondary antibody. Detection was done using a DAB Peroxidase HRP Substrate Kit (brown) followed by Hematoxylin counterstaining (purple). Images were obtained using inverted bright field microscopy. Sectioning results are representative of three individual tumors. Scale bar is 50 microns. Statistical analysis using One-Way ANOVA. * $p < 0.05$ vs tumor volume of the control. Image collected & cropped by CiteAb from the following publication (<https://www.oncotarget.com/lookup/doi/10.18632/oncotarget.22502>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



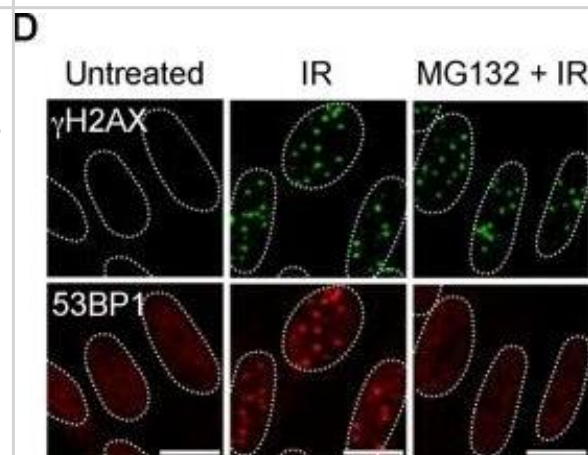
Western Blot: Histone H2AX [p Ser139] Antibody [NB100-384] - CPT induces transcription- & proteasome-dependent apoptosis in quiescent WI38 hTERT cells. (A & B) Serum-starved cells were treated with DMSO or FLV (1 μ M) for 1 h before the addition of DMSO or CPT (25 μ M) for 16 & 24 h. (A) Percentages of cells that remained attached to culture flasks (means \pm SD of quadruplicates). (B) Cell survival was analyzed by a CellTiter-Blue assay (means \pm SD of triplicates). (C–G) Western blot of the indicated proteins in serum-starved cells treated for 1 h with DMSO or with FLV (1 μ M) (C), lactacystin (10 μ M) (D), veliparib (5 μ M) or olaparib (10 μ M) (E), ATMi (10 μ M) (F) or DNA-PKi (10 μ M) (G) before the addition of DMSO ('-', 24 h in panels C & F) or CPT (25 μ M) for the indicated times. Data shown are representatives from two to three experiments. PARPCL: cleaved PARP. H2AX & α Tubulin were the loading controls. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/26578593>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



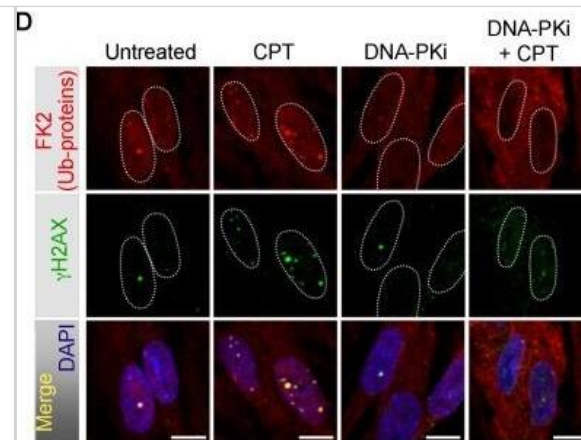
Western Blot: Histone H2AX [p Ser139] Antibody [NB100-384] - Inhibition of DNA-PK prevents monoubiquitination of H2AX & H2A in CPT-treated quiescent WI38 hTERT cells. (A–E) Serum-starved cells were treated with DMSO or DNA-PKi (10 μ M) for 1 h before the addition of DMSO (untreated) or CPT (25 μ M) for 1 h. (A) Western blot of γ H2AX. +Ub1 indicates γ H2AX monoubiquitinated. The top panel shows quantification of Ub1- γ H2AX normalized to α Tubulin (means \pm SEM, $n = 4$). ** $P < 0.01$. (B & C) Cells were pre-extracted with CSK buffer before co-staining for Ub-H2A (red) & 53BP1 phosphorylated on S1778 (p53BP1) (green). (B) Representative pictures. Images were merged to determine colocalization (yellow). The large Ub-H2AX focus at the periphery of the nuclei of untreated & CPT-treated cells may marks the inactive X chromosome as reported (91). (C) Percentages of nuclei with at least 5 Ub-H2A foci (means \pm SEM, $n = 3$, 100 nuclei were analyzed for each treatment in each experiment). *** $P < 0.001$. (D & E) Cells were co-stained for ubiquitinated proteins (FK2, red) & γ H2AX (green). (D) Representative pictures. Images were merged to determine colocalization (yellow). (E) Number of FK2 foci per nucleus from one representative experiment (76–111 nuclei were analyzed for each treatment) out of three. **** $P < 0.0001$. In the microscopic images, nuclear contours, identified by DAPI staining (blue in the merge images at bottom), are indicated by dashed lines. Bars: 10 μ m. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/26578593>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



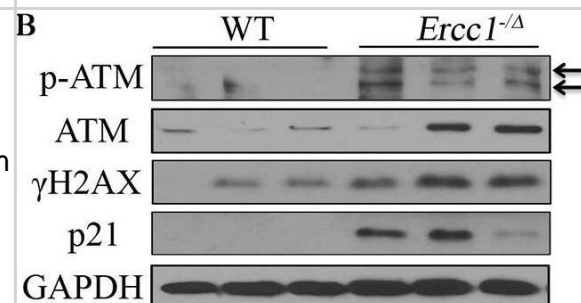
Immunocytochemistry/ Immunofluorescence: Histone H2AX [p Ser139] Antibody [NB100-384] - The production of DSBs depends on Top1 degradation in CPT-treated quiescent cells. (A–C) Serum-starved WI38 hTERT cells were co-transfected with siRNAs against cullin 3 & cullin 4B or against a control sequence & then treated with DMSO (–CPT) or 25 μ M CPT (+CPT) for 1 h. (A & B) Western blotting of the indicated proteins. α Tubulin: loading control. (C) Number of γ H2AX foci per nucleus from one representative experiment (246–348 nuclei were analyzed for each treatment) out of three. *** $P < 0.001$. (D & E) Serum-starved WI38 hTERT cells were treated with DMSO or MG132 (50 μ M) for 1 h before exposure to 0.8 Gy IR. One hour post-irradiation, cells were co-stained for γ H2AX (green) & 53BP1 (red). (D) Representative pictures. (E) Number of γ H2AX foci per nucleus from one representative experiment (162–180 nuclei were analyzed for each treatment) out of three. Ns: not significant. (F & G) U2OS EV28 cells were treated with DMSO or MG132 (10 μ M) for 1 h before the addition of ethanol (untreated) or 300 nM 4-hydroxitamoxifen (4OHT) for 4 h to express AsiSI in the nucleus (42). (F) Representative pictures of cells co-stained for γ H2AX (green) & 53BP1 (red). (G) ChIP analysis using an anti- γ H2AX antibody (black) or a non-immune antibody (IgG, gray). Enrichment was assessed by QPCR amplification using primers proximal to two AsiSI sites located inside two genes (Gene I: SFRS6, Gene II: CCD47) & primers distal to an AsiSI site (Control). Enrichment was normalized to the maximum recovery for each experiment (means \pm SEM, $n = 3$). Ns: not significant; * $P < 0.05$. In the microscopic images, nuclear contours, identified by DAPI staining (not shown), are indicated by dashed lines. Bars: 10 μ m. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/26578593>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



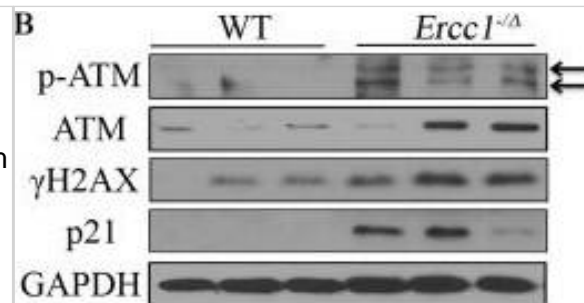
Immunocytochemistry/ Immunofluorescence: Histone H2AX [p Ser139] Antibody [NB100-384] - Inhibition of DNA-PK prevents monoubiquitination of H2AX & H2A in CPT-treated quiescent WI38 hTERT cells. (A–E) Serum-starved cells were treated with DMSO or DNA-PKi (10 μ M) for 1 h before the addition of DMSO (untreated) or CPT (25 μ M) for 1 h. (A) Western blot of γ H2AX. +Ub1 indicates γ H2AX monoubiquitinated. The top panel shows quantification of Ub1- γ H2AX normalized to α Tubulin (means \pm SEM, n = 4). **P < 0.01. (B & C) Cells were pre-extracted with CSK buffer before co-staining for Ub-H2A (red) & 53BP1 phosphorylated on S1778 (p53BP1) (green). (B) Representative pictures. Images were merged to determine colocalization (yellow). The large Ub-H2AX focus at the periphery of the nuclei of untreated & CPT-treated cells may marks the inactive X chromosome as reported (91). (C) Percentages of nuclei with at least 5 Ub-H2A foci (means \pm SEM, n = 3, 100 nuclei were analyzed for each treatment in each experiment). ***P < 0.001. (D & E) Cells were co-stained for ubiquitinated proteins (FK2, red) & γ H2AX (green). (D) Representative pictures. Images were merged to determine colocalization (yellow). (E) Number of FK2 foci per nucleus from one representative experiment (76–111 nuclei were analyzed for each treatment) out of three. ****P < 0.0001. In the microscopic images, nuclear contours, identified by DAPI staining (blue in the merge images at bottom), are indicated by dashed lines. Bars: 10 μ m. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/26578593>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



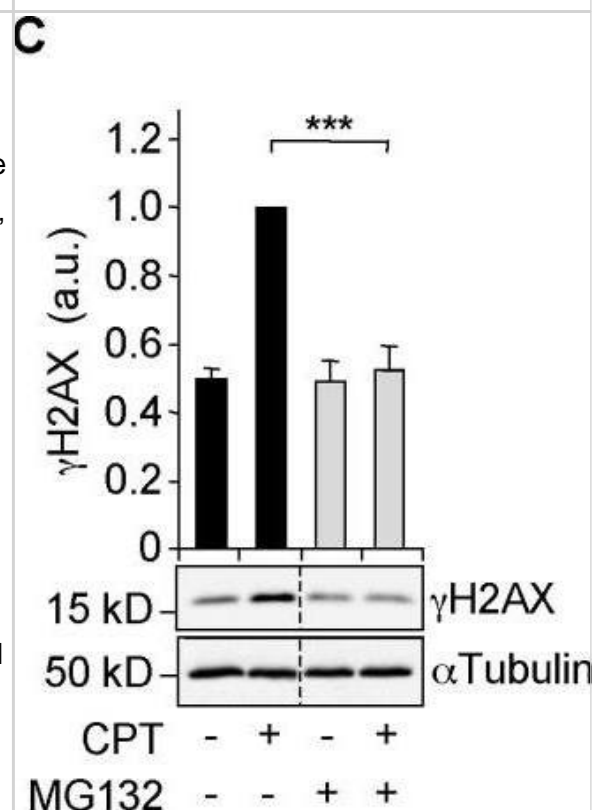
Western Blot: Histone H2AX [p Ser139] Antibody [NB100-384] - DDR & NF- κ B are activated concomitantly in senescent MEFs & aged tissues. (A) Immunoblot detection of p-p65 & total p65 in liver tissue from 16-week-old WT (n=3) & *Ercc1* $^{-/\Delta}$ (n=3) mice. (B) Immunoblot detection of phosphorylation of ATM & downstream targets γ H2AX & p21 in liver from 16-week-old WT & *Ercc1* $^{-/\Delta}$ mice. (C) Immunoblot detection of phosphorylation of NF- κ B & I κ B α in liver lysates from 3, 12 & 24 month-old WT mice. n=3 mice per group. (D) Immunoblot detection of p-ATM, ATM & p21 in the same liver lysates. (E) Immunoblot detection of DDR effectors in nuclear extracts from passage 5 WT & *Ercc1* $^{-/-}$ MEFs, grown at 20% oxygen. (F) Level of NF- κ B activation is higher in *Ercc1* $^{-/-}$ MEFs compared to WT MEFs at passage 5, as measured by Immunoblot detection of p-p65 & total p65 in WT & *Ercc1* $^{-/-}$ MEFs at passage 5 after culturing in 20% oxygen. (G) Representative images of immunofluorescent detection of p65 & NEMO in passage 4 WT & *Ercc1* $^{-/-}$ MEFs grown at 20% oxygen. Blue: DAPI staining; Green: p65 (top panel) or NEMO (bottom panel). Images were taken at the magnification of 60x. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/32201398>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



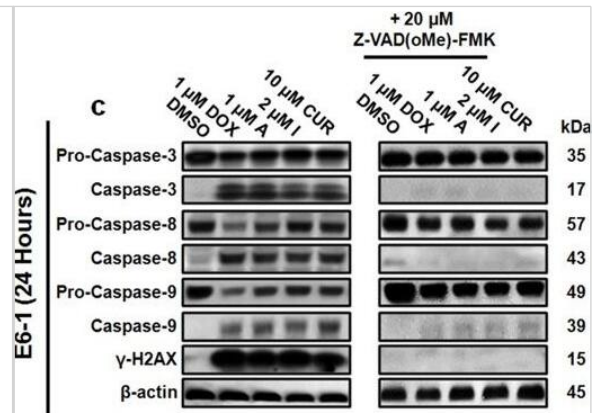
Western Blot: Histone H2AX [p Ser139] Antibody [NB100-384] - DDR & NF- κ B are activated concomitantly in senescent MEFs & aged tissues. (A) Immunoblot detection of p-p65 & total p65 in liver tissue from 16-week-old WT (n=3) & *Ercc1* Δ (n=3) mice. (B) Immunoblot detection of phosphorylation of ATM & downstream targets γ H2AX & p21 in liver from 16-week-old WT & *Ercc1* Δ mice. (C) Immunoblot detection of phosphorylation of NF- κ B & I κ B α in liver lysates from 3, 12 & 24 month-old WT mice. n=3 mice per group. (D) Immunoblot detection of p-ATM, ATM & p21 in the same liver lysates. (E) Immunoblot detection of DDR effectors in nuclear extracts from passage 5 WT & *Ercc1* Δ MEFs, grown at 20% oxygen. (F) Level of NF- κ B activation is higher in *Ercc1* Δ MEFs compared to WT MEFs at passage 5, as measured by Immunoblot detection of p-p65 & total p65 in WT & *Ercc1* Δ MEFs at passage 5 after culturing in 20% oxygen. (G) Representative images of immunofluorescent detection of p65 & NEMO in passage 4 WT & *Ercc1* Δ MEFs grown at 20% oxygen. Blue: DAPI staining; Green: p65 (top panel) or NEMO (bottom panel). Images were taken at the magnification of 60x. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/32201398>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



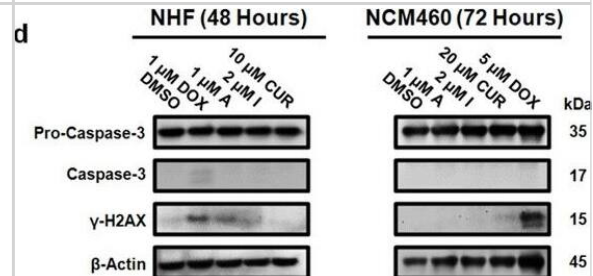
Western Blot: Histone H2AX [p Ser139] Antibody [NB100-384] - Induction of ubiquitin/proteasome-dependent DSBs in CPT-treated quiescent WI38 hTERT cells. (A–F) Serum-starved cells were treated with DMSO (1 h) or with MG132 (50 μ M, 1 h), lactacystin (10 μ M, 1 h), bortezomib (1 μ M, 4 h), G5 (1.5 μ M, 0.5 h) or Pyr-41 (9 μ M, 0.5 h) before the addition of DMSO (untreated) or 25 μ M CPT for 1 h & then co-stained for γ H2AX (green) & 53BP1 (red) or analyzed by Western blot. ‘-’ in panels C & F means cells treated with DMSO. (A & D) Representative pictures. Nuclear contours, identified by DAPI staining (not shown), are indicated by dashed lines. Bars: 10 μ m. (B & E) Number of γ H2AX foci per nucleus from two independent experiments (147–153 nuclei were analyzed for each treatment). ****P < 0.0001. (C & F) Western blot of γ H2AX. α Tubulin: loading control. Dashed lines indicate that intervening wells have been spliced out. The top panels show quantification of γ H2AX normalized to α Tubulin (means \pm SEM, n = 4 in panel C, n = 3 in panel F). ***P < 0.001; **P < 0.01. (G & H) Detection of DSBs by neutral Comet assays in serum-starved cells treated with DMSO or MG132 (25 μ M) for 1 h before the addition of DMSO (untreated) or CPT for 1 h (7.5 μ M for experiments (Exp) I & II; 5 & 7.5 μ M for Exp III). (G) Representative pictures of nuclei from Exp I. (H) Quantification of neutral Comet tail moments for three independent experiments (95–133 nuclei were analyzed for each treatment in each experiment). ***P < 0.001; ****P < 0.0001. The untreated & CPT data from Exp I are from the same experiment as that of Supplementary Figure S3D. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/26578593>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



Western Blot: Histone H2AX [p Ser139] Antibody [NB100-384] - Curcumin Analogs Induce Apoptosis in Cancerous Cells by Several Pathways. (a) E6-1 (Jurkat), dominant negative FADD (dnFADD) Jurkat, & overexpressing BCL-2 Jurkat were treated for 48 hours then stained for Annexin V & PI (b) E6-1 cells were plated & treated with or without the broad spectrum caspase inhibitor ZVAD(oMe)-FMK for 48 hours. Cells were stained for Annexin V & PI. Results were obtained using image-based cytometry with the Y-axis representative of percent of cells positive for Annexin V (green), PI (red), Annexin V & PI (yellow), or negative for both Annexin V & PI (blue). Values are expressed as a mean \pm SD from three independent experiments. (c) E6-1 cells were treated for 24 hours with or without the broad spectrum caspase inhibitor ZVAD(oMe)-FMK & the studied compounds, lysed & subjected to Western blot analysis. (d) NHF & NCM460 cells were treated for 48 hours & 72 hours respectively, lysed & subjected to Western blot analysis. Bands were visualized with a chemiluminescence reagent. Images are representative of three independent experiments. Statistical calculations were performed using Two-Way ANOVA multiple comparison. * $p < 0.05$ vs % viable of Control (DMSO); # $p < 0.05$ vs % viable cells for groups without Z-VAD(oMe)FMK. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/28439094>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



Western Blot: Histone H2AX [p Ser139] Antibody [NB100-384] - Curcumin Analogs Induce Apoptosis in Cancerous Cells by Several Pathways. (a) E6-1 (Jurkat), dominant negative FADD (dnFADD) Jurkat, & overexpressing BCL-2 Jurkat were treated for 48 hours then stained for Annexin V & PI (b) E6-1 cells were plated & treated with or without the broad spectrum caspase inhibitor ZVAD(oMe)-FMK for 48 hours. Cells were stained for Annexin V & PI. Results were obtained using image-based cytometry with the Y-axis representative of percent of cells positive for Annexin V (green), PI (red), Annexin V & PI (yellow), or negative for both Annexin V & PI (blue). Values are expressed as a mean \pm SD from three independent experiments. (c) E6-1 cells were treated for 24 hours with or without the broad spectrum caspase inhibitor ZVAD(oMe)-FMK & the studied compounds, lysed & subjected to Western blot analysis. (d) NHF & NCM460 cells were treated for 48 hours & 72 hours respectively, lysed & subjected to Western blot analysis. Bands were visualized with a chemiluminescence reagent. Images are representative of three independent experiments. Statistical calculations were performed using Two-Way ANOVA multiple comparison. * $p < 0.05$ vs % viable of Control (DMSO); # $p < 0.05$ vs % viable cells for groups without Z-VAD(oMe)FMK. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/28439094>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



Publications

Li Q, Yang G, Ren B, Liu X et Al. ZC3H14 facilitates backsplicing by binding to exon-intron boundary and 3' UTR Mol Cell 2024-10-26 [PMID: 39461343]

Herrera FG, Ronet C, Ochoa de Olza M, Barras D et Al. Low-Dose Radiotherapy Reverses Tumor Immune Desertification and Resistance to Immunotherapy Cancer Discov 2021-09-04 [PMID: 34479871]

Zeng F, Parker K, Zhan Y et al. Upregulated DNA Damage-Linked Biomarkers in Parkinson's Disease Model Mice ASN Neuro 2023-01-23 [PMID: 36683340]

S Min, HS Lee, JH Ji, Y Heo, Y Kim, S Chae, YW Choi, HC Kang, M Nakanishi, H Cho The chromatin remodeler RSF1 coordinates epigenetic marks for transcriptional repression and DSB repair Nucleic Acids Research, 2021-12-02;49(21):12268-12283. 2021-12-02 [PMID: 34850117]

Chen L, Gai X, Yu X. et Al. Pre-rRNA facilitates the recruitment of RAD51AP1 to DNA double-strand breaks J Biol Chem 2024-02-24 [PMID: 38403248]

Pignanelli C, Ma D, Noel M et al. Selective Targeting of Cancer Cells by Oxidative Vulnerabilities with Novel Curcumin Analogs. Sci Rep 2017-04-24 [PMID: 28439094]

DR Whelan, WTC Lee, F Marks, YT Kong, Y Yin, E Rothenberg Super-resolution visualization of distinct stalled and broken replication fork structures PloS Genetics, 2020-12-28;16(12):e1009256. 2020-12-28 [PMID: 33370257]

Fancy NN, Smith AM, Caramello A et Al. Characterisation of premature cell senescence in Alzheimer's disease using single nuclear transcriptomics Acta Neuropathol 2024-05-02 [PMID: 38695952]

Whelan DR, Rothenberg E., et Al. Super-resolution mapping of cellular double-strand break resection complexes during homologous recombination Proc Natl Acad Sci U S A 2021-03-16 [PMID: 33707212]

Idrees SM, Waite SL, Granados Aparici S et Al. Nicotine exposure is associated with targeted impairments in primordial follicle phenotype in cultured neonatal mouse ovaries Ecotoxicol Environ Saf 2024-11-14 [PMID: 39546863]

Fried W, Tyagi M, Minakhin L et Al. Discovery of a small-molecule inhibitor that traps Pol θ on DNA and synergizes with PARP inhibitors Nat Commun 2024-04-05 [PMID: 38580648]

Yu L, Wu D., et Al. SMARCA2 and SMARCA4 Participate in DNA Damage Repair Front Biosci (Landmark Ed) 2024-07-31 [PMID: 39082357]

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