Product Datasheet

LC3A Antibody - BSA Free NB100-2331

Unit Size: 0.1 ml

Aliquot and store at -20C or -80C. Avoid freeze-thaw cycles.

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NB100-2331

LC3A Antibody - BSA Free

Product Information		
Unit Size	0.1 ml	
Concentration	1.0 mg/ml	
Storage	Aliquot and store at -20C or -80C. Avoid freeze-thaw cycles.	
Clonality	Polyclonal	
Preservative	0.02% Sodium Azide	
Isotype	IgG	
Purity	Immunogen affinity purified	
Buffer	PBS	
Product Description		
Host	Rabbit	
Gene ID	84557	
Gene Symbol	MAP1LC3A	
Species	Human, Mouse, Rat, Amphibian, Canine, Fish, Plant, Zebrafish	
Reactivity Notes	Use in Rat reported in scientific literature (PMID:33678798). Use in Amphibian reported in scientific literature (PMID:29777142).	
Marker	Autophagosome Marker	
Specificity/Sensitivity	This LC3A Antibody detects both LC3A and LC3B.	
Immunogen	This LC3A Antibody was prepared from a synthetic peptide made to an internal portion of the human LC3 protein sequence (between residues 25-121). [Uniprot: Q9H492].	
Product Application Details		
Applications	Western Blot, Simple Western, Chromatin Immunoprecipitation, ELISA, Flow Cytometry, Immunoblotting, Immunocytochemistry/ Immunofluorescence, Immunohistochemistry, Immunohistochemistry-Frozen, Immunohistochemistry- Paraffin, Immunoprecipitation, Southern Blot, Chromatin Immunoprecipitation (ChIP), Immunohistochemistry Whole-Mount	
Recommended Dilutions	Western Blot 2.0 ug/ml, Simple Western 1:50, Chromatin Immunoprecipitation reported in scientific literature (PMID 33035707), Flow Cytometry reported in scientific literature (PMID 24419333), ELISA reported in scientific literature (PMID 20930550), Immunohistochemistry 1:200-1:400, Immunocytochemistry/ Immunofluorescence 1:100-1:300. Use reported in scientific literature (PMID 21545732), Immunoprecipitation 20 ug / 500 ug of lysate, Immunohistochemistry-Paraffin 1:200-1:400. Use reported in scientific literature (PMID 26571030), Immunohistochemistry-Frozen, Immunoblotting reported in scientific literature (PMID 28253371), Southern Blot, Immunohistochemistry Whole-Mount reported in scientific literature (PMID 31783118), Chromatin Immunoprecipitation (ChIP)	



Application	Notes
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ItesWestern blot bands are seen at ~19 kDa, representing LC3-I, and ~17 kDa,
representing LC3-II. The observed molecular weight of the protein may vary from
the listed predicted molecular weight due to post translational modifications, post
translation cleavages, relative charges, and other experimental factors. Use in
Southern blot reported in scientific literature (PMID: 21262964). In ICC,
cytoplasmic staining was observed in HeLa cells. In Simple Western only 10 - 15
uL of the recommended dilution is used per data point.
See Simple Western Antibody Database
for Simple Western validation: Tested in
Neuro2A lysate 0.5 mg/mL, separated by Size, antibody dilution of 1:50,
apparent MW was 16 kDa

Images

Western Blot: LC3A Antibody - BSA Free [NB100-2331] - LC3A Antibody [NB100-2331] - High autophagosome concentration is consumed during early immortalized human mesenchymal stem cell differentiation. Immortalized human mesenchymal stem cells were differentiated under osteogenic conditions (see Materials and methods) and assayed for changes in LC3I and LC3II during a 72-hour window. Cells were differentiated under standard conditions (top) or with addition of 5 uM rapamycin (middle) or 5 nM bafilomycin (bottom) for the first 3 hours of differentiation to modulate autophagy. Immunoblots were performed for LC3 at the indicated time points to assess autophagosome degradation via relative changes in LC3II (lower band; 17 kDa). Studies were repeated three times with similar trends seen consistently. Image collected and cropped by CiteAb from the following publication (https://stemcellres.com/content/5/6/140), licensed under a CC-BY license.

Immunocytochemistry/Immunofluorescence: LC3A Antibody - BSA Free [NB100-2331] - LC3A Antibody [NB100-2331] - Analysis in PFA fixed NIH/3T3 cells using anti-LC3A antibody. Image from verified customer review.

Immunohistochemistry: LC3A Antibody - BSA Free [NB100-2331] - LC3A Antibody [NB100-2331] - Staining in mouse meniscus and cartilage. Image from verified customer review.















hippocampal CA1 in control- and rapamycin-fed transgenic PDAPP mice stained with an anti-LC3 antibody. An increase in LC3-immunoreactive puncta was observed in CA1 projections of transgenic PDAPP mice following rapamycin administration. Image collected and cropped by CiteAb from the following publication (https://dx.plos.org/10.1371/journal.pone.0009979) licensed under a CC-BY license.

LC3 DAPI Overlay



Immunohistochemistry: LC3A Antibody - BSA Free [NB100-2331] - LC3A Antibody [NB100-2331] - Analysis in mouse renal tissue. Image from verifed customer review.



Simple Western: LC3A Antibody - BSA Free [NB100-2331] - LC3A Antibody [NB100-2331] - Image shows a specific band for LC3 in 0.5 mg/mL of Neuro2A lysate. This experiment was performed under reducing conditions using the 12-230 kDa separation system. P-AMPKa (Thr172) Western Blot: LC3A Antibody - BSA Free [NB100-2331] - Chronic, third GAPDH window RIC increases the expression of autophagosome proteins, P-mTOR (Ser2481) (289kDa) LC3I/II & Atg5.(A) Western blots for autophagy related signaling proteins. Beclin-1 (60kDa) (B) Quantification of the protein fold change in 3W RIC compared to 3W GAPDH controls. Values are means ± S.E.M. n=6-8 per group. An (*) denotes a Atg5 (55kDa) statistically significant difference (P<0.05) compared to control. (P-: GAPDH phospho-). Image collected & cropped by CiteAb from the following LC3 I (19kDa LC3 II (17kDa publication (https://pubmed.ncbi.nlm.nih.gov/25347774), licensed under GAPDH a CC-BY license. Not internally tested by Novus Biologicals. P62 (60kDa) GAPDH Cathepsin L (23kda) GAPDH -Contro BAF Ctrl Western Blot: LC3A Antibody - BSA Free [NB100-2331] - PRDX3 Α BPH-1 **RWPE-1** BPH-1 **RWPE-1** expression & its association with autophagy flux in cultured prostate cells KD (A) Representative immunoblot showing the levels of PRDX3, TOM20 & PRDX3 28 LC3-II in in lysates prepared from three different cultures of BPH-1 & **TOM20** 20 RWPE-1 cells in the absence (Ctrl) or presence of bafilomycin A1 (BAF). 17 (B–D) The quantification of the relative levels of PRDX3 (B), TOM20 (C) LC3-I 14 LC3-II & LC3-II (D) to β-Actin as shown in (A). Data are mean & standard deviation of three repeats & differences are tested with Student's T-test. **B-Actin** *P \leq 0.05; **P \leq 0.01. Image collected & cropped by CiteAb from the following publication (https://www.oncotarget.com/lookup/doi/10.18632/oncotarget.17927), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



Western Blot: LC3A Antibody - BSA Free [NB100-2331] - Acute, first window RIC activates autophagy signaling via p-AMPK upregulation & concomitant downregulation of mTOR.(A) Western blots for autophagy related signaling proteins. (B) Quantification of the protein fold change in 1W RIC compared to 1W controls. Values are means ± S.E.M. n=6–8 per group. An (*) denotes a statistically significant difference (P<0.05) compared to control. (P-: phospho-). Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/25347774), licensed under a CC-BY license. Not internally tested by Novus Biologicals.	A P-AMPKa (Thr172) \rightarrow (62k0a) (62k0a) P-mTOR (Ser281) \rightarrow (289k0a) GAPDH \rightarrow (289k0a) Beclin-1 (60kDa) \rightarrow (60kDa) \rightarrow GAPDH \rightarrow (GAPDH \rightarrow LC3 I (19kDa) \Rightarrow (GAPDH \rightarrow LC3 I (19kDa) \Rightarrow (GAPDH \rightarrow C3 (19kDa) \Rightarrow (GAPDH \rightarrow C3 (19kDa) \Rightarrow (GAPDH \rightarrow Cathepsin L (28kda) \rightarrow (Cathepsin L (28kda) \rightarrow
Western Blot: LC3A Antibody - BSA Free [NB100-2331] - LiCl administered via 0.5% LiCl food pellets for 4 wks does not increase markers for autophagy in Gfap-R236H/+ mice.Each lane of the immunoblots is tissue from one mouse. Immunoblots for LC3-I & LC3-II in (A) did not detect a change in parietal cortex (and underlying white matter) with LiCl treatment. LC3-II bands normalized to LC3-I are quantified in B (N = 3–4 mice from 3–4 cages per genotype, & is representative of 3 blots). LC3-II normalized to GAPDH gave similar results & is not shown. P62 was increased in control diet R236H/+ mouse olfactory bulb compared with control diet +/+, but LiCl did not change P62 levels in GFAP+/+ or R236H/+ mice (C-D). P62 was normalized to total protein loaded. Error bars are SEM. ****P < 0.0001. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/26378915), licensed under a CC-BY license. Not internally tested by Novus Biologicals.	A +/+ R236H/+ +/+ R236H/+ +/+ R236H/+ B G G G G G G G
Western Blot: LC3A Antibody - BSA Free [NB100-2331] - MIR376A overexpression blocked autophagy in Huh-7 cells.(A) MIR376A blocked GFP-LC3 dot formation under starvation condition. (B) Quantitative analysis of experiments in A (mean \pm S.D. of independent experiments, n =3, ***p<0,01). (C) Overexpression of MIR376A resulted in decreased autophagic flux in Huh-7 cells. Starvation-induced conversion of LC3-I to LC3-II was analyzed. Tests were performed in the presence or absence of E64d (10 µg/ml) & Pepstatin A (10 µg/ml) (E+P). LC3-II/LC3-I densitometric ratios were marked. ACTB was used as a loading control. Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/24358205), licensed under a CC-BY license. Not internally tested by Novus Biologicals.	C <u>MIR-CNT</u> <u>MIR376A</u> STV: - + + - + + E+P: + + LC3-I LC3-II ACTB
Immunohistochemistry: LC3A Antibody - BSA Free [NB100-2331] - Elevated levels of additional proteins & GM3 ganglioside in the MEC of MPS IIIB brain.Staining performed with antibodies to the indicated substances was observed in the MEC region of 3 month-old MPS IIIB mice (for total ubiquitin & polyubiquitin) & 6 months for all others. Staining was not seen in the MEC region of age-matched control mice (Naglu +/-) nor in the LEC region of MPS IIIB mice (the latter not shown). Image collected & cropped by CiteAb from the following publication (https://dx.plos.org/10.1371/journal.pone.0027461), licensed under a CC-BY license. Not internally tested by Novus Biologicals.	-/- MEC +/- MEC



Page 6 of 20 v.20.1 Updated 4/13/2025

Western Blot: LC3A Antibody - BSA Free [NB100-2331] - Autophagy responds rapidly to changing glucose conditions. Immortalized MSCs were cultured in physiologic (also called low in culture parlance; 1 g/L; 5.5 mM) or high (4.5 g/L; 25 mM) glucose media for 2 days & then changed to the corresponding opposite glucose concentration for up to 96 h. Myosin light chain 3 (LC3) levels were probed via immunoblot to assess autophagic response (a). The role of oxygen in the glucose response was also assessed by culturing the MSCs in a Biospherix hypoxic chamber at 4% & 1% oxygen in high & low glucose media for 4 days, followed by comparable LC3 blots (b). Shown are representative blots of three repeated studies. α -Actinin was used as a housekeeping control for all blots Image collected & cropped by CiteAb from the following publication

(https://stemcellres.biomedcentral.com/articles/10.1186/s13287-016-0436-7), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Immunohistochemistry: LC3A Antibody - BSA Free [NB100-2331] -Elevated levels of various proteins in MEC & in dentate gyrus of MPS IIIA brain. Staining was performed with antibodies against the proteins shown in a 7 months-old MPS IIIA mouse brain. The first 3 rows are for the MEC region & the bottom row for dentate gyrus. Age-matched C57BL6 mice, used as controls, showed no staining in the MEC region (not shown). The dentate gyrus showed AT270 inclusions in MPS IIIA (-/ -) mouse brain (arrows) but not in the C57BL/6 (control) brain (bottom row). Image collected & cropped by CiteAb from the following publication (https://dx.plos.org/10.1371/journal.pone.0027461), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: LC3A Antibody - BSA Free [NB100-2331] - Autophagy responds rapidly to changing glucose conditions. Immortalized MSCs were cultured in physiologic (also called low in culture parlance; 1 g/L; 5.5 mM) or high (4.5 g/L; 25 mM) glucose media for 2 days & then changed to the corresponding opposite glucose concentration for up to 96 h. Myosin light chain 3 (LC3) levels were probed via immunoblot to assess autophagic response (a). The role of oxygen in the glucose response was also assessed by culturing the MSCs in a Biospherix hypoxic chamber at 4% & 1% oxygen in high & low glucose media for 4 days, followed by comparable LC3 blots (b). Shown are representative blots of three repeated studies. α -Actinin was used as a housekeeping control for all blots Image collected & cropped by CiteAb from the following publication

(https://stemcellres.biomedcentral.com/articles/10.1186/s13287-016-0436-7), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: LC3A Antibody - BSA Free [NB100-2331] - Delayed, second window RIC maintains mTOR inhibition without activating autophagosome machinery.(A) Western blots for autophagy related signaling proteins. (B) Quantification of the protein fold change in 2W RIC compared to 2W controls. Values are means \pm S.E.M. n=6–8 per group. An (*) denotes a statistically significant difference (P<0.05) compared to control. (P-: phospho-). Image collected & cropped by CiteAb from the following publication

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Control

Page 8 of 20 v.20.1 Updated 4/13/2025 LC3 Immunohistochemistry: LC3A Antibody - BSA Free [NB100-2331] -Elevated levels of various proteins in MEC & in dentate gyrus of MPS IIIA brain. Staining was performed with antibodies against the proteins shown in a 7 months-old MPS IIIA mouse brain. The first 3 rows are for the MEC region & the bottom row for dentate gyrus. Age-matched C57BL6 mice, used as controls, showed no staining in the MEC region (not shown). The dentate gyrus showed AT270 inclusions in MPS IIIA (-/ -) mouse brain (arrows) but not in the C57BL/6 (control) brain (bottom row). Image collected & cropped by CiteAb from the following publication (https://dx.plos.org/10.1371/journal.pone.0027461), licensed under a CC-BY license. Not internally tested by Novus Biologicals. Western Blot: LC3A Antibody - BSA Free [NB100-2331] - Modulation of J Clutch 1 Clutch 2 Clutch 3 A152T-tau clearance & pathology by upregulation of autophagy. (J–N) Injection of an expression vector encoding zebrafish atg5 into A152T-tau Ctrl Atg5 Ctrl Atg5 Ctrl Atg5 fish embryos resulted in over-expression of Atg5 protein at 2 dpf (J & K) Low (high & low exposure of same blot presented; mean \pm SD, n = 6 independent clutches; two-tailed t-test, *P < 0.05 versus control). (J & L) ā High 40 KDa The increase in Atg5 protein correlated with increase in LC3II, a wellcharacterized reporter for autophagosome number, demonstrating that 17 KDa LC3II autophagy was upregulated in Atg5-injected fish (mean \pm SEM, n = 8 independent clutches; two-tailed t-test, ***P < 0.001 versus control). 42 KDa Actin Image collected & cropped by CiteAb from the following publication (https://academic.oup.com/brain/article/140/4/1128/2980948), licensed under a CC-BY license. Not internally tested by Novus Biologicals. LC3 Immunohistochemistry: LC3A Antibody - BSA Free [NB100-2331] --/- MEC +/- MEC Elevated levels of additional proteins & GM3 ganglioside in the MEC of MPS IIIB brain. Staining performed with antibodies to the indicated substances was observed in the MEC region of 3 month-old MPS IIIB mice (for total ubiquitin & polyubiquitin) & 6 months for all others. Staining was not seen in the MEC region of age-matched control mice (Naglu +/-) nor in the LEC region of MPS IIIB mice (the latter not shown). Image collected & cropped by CiteAb from the following publication (https://dx.plos.org/10.1371/journal.pone.0027461), licensed under a CC-BY license. Not internally tested by Novus Biologicals. Ctrl BAF С Western Blot: LC3A Antibody - BSA Free [NB100-2331] - Impacts of Α Ctrl BAF **PRDX3** PRDX3 PRDX3 protein on autophagy flux(A–D) Representative immunoblot (A, Plasmid Mock Mock KD siRNA C) & quantification (B, D) showing the levels of LC3-II in BPH-1 cells PRDX3 KD treated with random (MOCK) or PRDX3-specific siRNA (PRDX3) (A, B) PRDX3 28 LC3-I 17 LC3-I 17 or RWPE-1 cells transiently expressing different amount of PRDX3 (C, LC3-II 14 LC3-II D) in the absence (Ctrl) or presence of bafilomycin A1 (BAF). Data are 42 **B-Actin** β-Actin mean & standard deviation of three repeats & differences are tested with Student's T-test. * $P \le 0.05$; ** $P \le 0.01$; *** $P \le 0.001$. (E–G) Representative immunoblot (E) & quantification (F, G) showing the levels of Beclin 1 (F) & PI3KCIII (G) in BPH-1 cells treated with random (MOCK) or PRDX3-specific siRNA (PRDX3). Ns, not significant; *P ≤ 0.05. Image collected & cropped by CiteAb from the following publication (https://www.oncotarget.com/lookup/doi/10.18632/oncotarget.17927), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



Page 9 of 20 v.20.1 Updated 4/13/2025





Western Blot: LC3A Antibody - BSA Free [NB100-2331] - Optn or p62 deficiency affects autophagosome formation.(A) Workflow of the experiments shown in (B-G). Larvae were treated with 100 nM of Baf A1 for 12 h from 3.5 dpf. The GPF-Lc3 negative larvae were selected to assay autophagy activity by Western blot, the GFP-Lc3 positive larvae were collected to monitor autophagic activity using confocal imaging. The red square indicates the region for confocal imaging. (B) Level of basal autophagy in WT & mutant embryos in absence or presence of Baf A1. Protein samples were extracted from 4 dpf WT & mutant larvae (>10 embryos/sample). The blots were probed with antibodies against Lc3 & Actin as a loading control. Western blots were repeated at least three times with independent extracts. (C) Quantification of Lc3-II fold changes in WT & mutant embryos in absence or presence of Baf A1. Western blot band intensities were quantified by Lab Image. Data is combined from three independent experiments. (D) Representative confocal micrographs of GFP-Lc3 puncta present in the tail fin of optn+/+, optn Δ 5n/ Δ 5n, p62+/+ & p62 Δ 37n/ Δ 37n at 4 dpf. Scale bars, 10 µm. (E). Quantification of the number of GFP-Lc3 puncta in optn+/+, optn Δ 5n/ Δ 5n, p62+/+ & p62 Δ 37n/ Δ 37n larvae with & without Baf A1 treatment. Each larva was imaged at a pre-defined region of the tail fin (as indicated by the red boxed area in Fig3 A) (\geq 11 larvae/group). Results are accumulated from two independent experiments, ns. nonsignificant, *p<0.05, **p<0.01, ***p<0.001. Image collected & cropped by CiteAb from the following publication

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Immunohistochemistry: LC3A Antibody - BSA Free [NB100-2331] -Rapamycin increases autophagy in brains of PDAPP mice.a, f & h, representative immunoblots of hippocampal lysates from control- & rapamycin-treated transgenic PDAPP mice & non-transgenic littermate controls, b, g & i, quantitative analyses, a & b, LC3-II levels are decreased in hippocampi of rapamycin-treated transgenic PDAPP mice (*, P=0.0009), but not in hippocampi of rapamycin-treated nontransgenic littermates. c & d, representative epifluorescent (c, 200×) & higher-magnification confocal (d, 600×) images of hippocampal CA1 (e, green box, region of epifluorescent images; blue box, region of confocal images) in control- & rapamycin-fed transgenic PDAPP mice stained with an anti-LC3 antibody. An increase in LC3-immunoreactive puncta was observed in CA1 projections of transgenic PDAPP mice following rapamycin administration. f & g, levels of the autophagic substrate p62SQSTM are decreased (*, P=0.0015) in hippocampi of rapamycintreated PDAPP transgenic mice. f, representative Western blots; g, quantitative analyses of p62SQSTM levels. h & i, Levels of phosphorylated (activated) p70 were decreased in brains of rapamycintreated PDAPP & non-transgenic mice (*, P=0.001 & P=0.04 respectively). Significance of differences between group means were determined using two-tailed unpaired Student's t test. Data are means ± SEM. Image collected & cropped by CiteAb from the following publication (https://dx.plos.org/10.1371/journal.pone.0009979), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Page 10 of 20 v.20.1 Updated 4/13/2025







Immunocytochemistry/ Immunofluorescence: LC3A Antibody - BSA Free [NB100-2331] - Macroautophagy is a major mechanism in the rapid disposal of insulin precursor in β -cells. The Ins2+/+ β -cells were cultured under the 5.5 mM glucose concentration for a 24-hour pre-experimental period until treatment. (A) The $lns2+/+\beta$ -cells were treated with cycloheximide (Chx; 100 µg/mL), Chx (100 µg/mL) & 3-MA (5 mM), Baf A1 (5 μ M), or chloroquine (Chl; 100 μ g/mL) for 30 minutes with an untreated control. Cellular proteins (30 µg) were separated by 16.5% non-reduced (upper panels) or reduced (lower panels) tricine SDS-PAGE & then examined by immunoblotting. (B) The upper panel, the immunoreactive LC3-I/II (%) in individual treatments in (A); the lower panel, the percentages of proinsulin levels on reduced gels (that were normalized by tubulin) in individual treatments compared to the untreated controls. (C) The Ins2+/+ β -cells subjected to the same treatments described in (A) were immunostained with antibodies against LC3, Cpeptide, & insulin as described in the Materials & Methods. Fluorescent Cv2 (for LC3), Cv3 (for C-peptide & insulin), & their merged images were shown. The scale of bar in (C), 10 μ m. (D) The relative levels (%) of the LC3 & (pro)insulin and/or C-peptide positive dots per cell of individual treatments in (C). The data in (B) or (D) were reported as mean ± SD. *P<0.05; **P<0.01, n=4. Image collected & cropped by CiteAb from the following publication (https://dx.plos.org/10.1371/journal.pone.0027647), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: LC3A Antibody - BSA Free [NB100-2331] - DBA mutations induce autophagy.(A) Immunofluorescence with LC3 antibodies in LCLs derived from a normal control or DBA patients. Higher magnifications are represented in the lower panel. Arrows denote puncta indicative of LC3 recruitment to autophagosomes, or accumulation in autolysosomes. Size bars=10 µM. (B) Quantification of the percent of cells revealing LC3 puncta compared to the total number of cells in the 60x shots. (C) Western blot analysis of LC3 in DBA LCLs compared to normal controls. The LC3II/actin ratio is determined by densitometer analysis. (D) Representative western blot analysis of p62 levels in normal control & DBA patient LCLs. (E) Densitometer analysis of p62 protein expression from western blots (N=3) represented in (D). (F) Immunofluorescence with p62 antibodies of LCLs derived from a normal control or DBA patients. Size bars=10 µM. (G) ImageJ measurements of p62 expression in (F) per total cell area. (H) Representative electron micrographs of LCLs derived from a normal control & RPS17 cells. Control cells have small typically dense lysosomes (*). The much larger autolysosomes (A) are only detected in RPS17 LCLs. The boxed area in the upper right panel is shown at higher magnification in the lower right panel. N=nucleus, ECS=extracellular space. Bars in top panels=1 μ M, bottom panels=200 nM. Image collected & cropped by CiteAb from the following publication (https://dx.plos.org/10.1371/journal.pgen.1004371), licensed under a CC-BY license. Not internally tested by Novus Biologicals.





Western Blot: LC3A Antibody - BSA Free [NB100-2331] - UBC9 is degraded by autophagy in epithelial cells.(A) Immunolocalization of UBC9 in ultrathin sections of HKs transduced with empty or HPV16 E6/E7 vectors. Original view (left) & higher magnification (right) of boxed regions. Gold particles are selectively enriched in autophagic structures highlighted by arrows. Scale bar = 1 μ m. (B) Top: Representative WB of HaCaT cells treated with the indicated autophagic activators (left) & inhibitors (right). Activation of autophagy was monitored by the conversion of LC3 (LC3-I) to the lipidated LC3 (LC3-II) form, a marker of autophagosome production induced by autophagic stimuli [41]. LC3-II accumulation was used to verify autophagic impairment [41]. Bottom: normalized UBC9 expression. Data are expressed as fold over untreated cells. Bars represent means ± SEM of n = 4 different biological replicates. ns: not significant (Kruskal-Wallis one-way ANOVA with Dunn's post hoc test) compared to vehicle control groups. (C) Representative WB analysis of U-2 OS or MCF7 cells treated with chloroquine. n = 3 different biological replicates. (D) Representative WB analysis of MCF7 cells transduced with scramble or ATG5 shRNA. The observed ATG5 band represents the ATG5-ATG12 conjugated form. LC3-I accumulation is reported to evidence autophagic deficiencies promoted by shATG5. n = 3 replicates of a single transduction. Image collected & cropped by CiteAb from the following publication (https://dx.plos.org/10.1371/journal.ppat.1006262), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Western Blot: LC3A Antibody - BSA Free [NB100-2331] -Macroautophagy is a major mechanism in the rapid disposal of insulin precursor in β -cells.The Ins2+/+ β -cells were cultured under the 5.5 mM glucose concentration for a 24-hour pre-experimental period until treatment. (A) The Ins2+/+ β -cells were treated with cycloheximide (Chx; 100 µg/mL), Chx (100 µg/mL) & 3-MA (5 mM), Baf A1 (5 µM), or chloroquine (Chl; 100 µg/mL) for 30 minutes with an untreated control. Cellular proteins (30 µg) were separated by 16.5% non-reduced (upper panels) or reduced (lower panels) tricine SDS-PAGE & then examined by immunoblotting. (B) The upper panel, the immunoreactive LC3-I/II (%) in individual treatments in (A); the lower panel, the percentages of proinsulin levels on reduced gels (that were normalized by tubulin) in individual treatments compared to the untreated controls. (C) The Ins2+/ + β-cells subjected to the same treatments described in (A) were immunostained with antibodies against LC3, C-peptide, & insulin as described in the Materials & Methods. Fluorescent Cy2 (for LC3), Cy3 (for C-peptide & insulin), & their merged images were shown. The scale of bar in (C), 10 µm. (D) The relative levels (%) of the LC3 & (pro)insulin and/or C-peptide positive dots per cell of individual treatments in (C). The data in (B) or (D) were reported as mean \pm SD. *P<0.05; **P<0.01, n=4. Image collected & cropped by CiteAb from the following publication (https://dx.plos.org/10.1371/journal.pone.0027647), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Page 12 of 20 v.20.1 Updated 4/13/2025









highlighted by arrows. Scale bar = 1 μ m. (B) Top: Representative WB of HaCaT cells treated with the indicated autophagic activators (left) & inhibitors (right). Activation of autophagy was monitored by the conversion of LC3 (LC3-I) to the lipidated LC3 (LC3-II) form, a marker of autophagosome production induced by autophagic stimuli [41]. LC3-II accumulation was used to verify autophagic impairment [41]. Bottom: normalized UBC9 expression. Data are expressed as fold over untreated cells. Bars represent means ± SEM of n = 4 different biological replicates. ns: not significant (Kruskal–Wallis one-way ANOVA with Dunn's post hoc test) compared to vehicle control groups. (C) Representative WB analysis of U-2 OS or MCF7 cells treated with chloroquine. n = 3 different biological replicates. (D) Representative WB analysis of MCF7 cells transduced with scramble or ATG5 shRNA. The observed ATG5 band represents the ATG5-ATG12 conjugated form. LC3-I accumulation is reported to evidence autophagic deficiencies promoted by shATG5. n = 3 replicates of a single transduction. Image collected & cropped by CiteAb from the following publication (https://dx.plos.org/10.1371/journal.ppat.1006262), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Page 13 of 20 v.20.1 Updated 4/13/2025





Western Blot: LC3A Antibody - BSA Free [NB100-2331] - Dram1 is required for GFP-Lc3 targeting to Mm clusters.a, b Representative confocal micrographs & quantification of GFP-Lc3 puncta in dram1 Δ 19n/ Δ 19n & dram1+/+ larvae in an unstimulated situation (basal autophagy, a) & following BafA1 treatment b. Each larva was imaged at a pre-defined region of the tail fin (\geq 11 larvae/group). Results are accumulated from two independent experiments & represented by scatter & boxplots as detailed in the "Methods" section. ns nonsignificant, *p < 0.05,**p < 0.01,***p < 0.001. Scale bars, 10 µm. The intensity calibration bar for the Lookup table (LUT) is displayed in panel a. c–e Western blot analysis of autophagy. Protein samples were obtained from 4 dpf dram1 Δ 19n/ Δ 19n & dram1+/+ larvae (>10 larvae/sample). Lc3 c & e, or p62 & Optineurin d protein levels were detected in absence or presence of BafA1, c & d, or in the presence or absence of Mm e. Actin was used as a loading control. Western Blots were repeated three, c & d, or two e times with protein extracts derived from independent experiments. The Lc3II/Actin or p62/Actin & Optineurin/Actin ratio, normalized to the control sample, is indicated below the blots. f-g Representative confocal micrographs & quantification of GFP-Lc3 co-localization with Mm clusters in infected dram1 Δ 19n/ Δ 19n & dram1+/+ larvae. The top images f show the entire region of imaging, while the bottom images f' & f'' show details of GFP-Lc3 colocalization of Mm clusters in dram1 Δ 19n/ Δ 19n & dram1+/+ larvae. The arrowheads indicate GFP-Lc3-positive Mm clusters. The data is accumulated from two independent experiments (≥15 larvae/group) & represented by scatter & boxplots as detailed in the "Methods" section. Scale bars, 10 µm. Image collected & cropped by CiteAb from the following publication

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Western Blot: LC3A Antibody - BSA Free [NB100-2331] - Knockdown of FUT1 is associated with an increase in autophagic flux. (a) Immunoblot analysis of LC3-II & p62 levels in control & FUT1 knockdown cells. Total cell lysates from MCF-7 & T47D cells transfected with control or FUT1 siRNAs were collected at 120 & 96 h post-transfection, respectively. Equal amounts of cell lysates were then loaded in each lane & separated by SDS-PAGE. Immunoblot analysis was performed with LC3 & p62 antibodies. Actin was used as a loading control. The intensity of LC3-II & p62 protein bands on immunoblot were quantified & normalized to actin, & the relative levels of protein expression were expressed as fold change by setting the control group value to 1. Values shown are mean±S.E.M. of three independent experiments (***P<0.001; **P<0.01; *P<0.05). (b) Downregulation of FUT1 enhanced the fusion of autophagosome & lysosomes in MCF-7 cells. Cells were co-stained with anti-LAMP-1 (green) & anti-LC3 (red) & nuclei stained with Hoechst (blue). Representative colocalization signals (referred to as autolysosomes) were shown in yellow in the merged. Magnification \times 63, zoom: \times 3. Scale bars, 10 µm. Histogram shows the percentages of autolysosomes (LC3+/LAMP-1+) to autophagosomes (LC3+/LAMP-1-). Data are mean \pm S.E.M. of three independent experiments of >100 cells per group (*P<0.05) Image collected & cropped by CiteAb from the following publication (https://pubmed.ncbi.nlm.nih.gov/27560716), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

Page 14 of 20 v.20.1 Updated 4/13/2025





Page 15 of 20 v.20.1 Updated 4/13/2025





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More publications at http://www.novusbio.com/NB100-2331



Procedures

Western Blot protocol specific for LC3 Antibody (NB100-2331)

Protocol: Inhibition of Autophagy and LC3 Antibody (NB100-2331) Western Blot

Materials

Chloroquine diphosphate (CQ) (10 mM) in dH2O 1X PBS Sample buffer, 2X Laemmli buffer: 4% SDS, 5% 2-mercaptoethanol (BME), 20% glycerol, 0.004% bromophenol blue, 0.125 M Tris HCl, pH 6.8 RIPA buffer: 150 mM NaCl, 1% NP-40 or Triton X-100, 0.5% sodium deoxycholate, 0.1% SDS, 50 mM Tris-HCl, pH 8.0, 20 mM Tris-HCl, pH 7.5 1X Running Buffer: 25 mM Tris-base, 192 mM glycine, 0.1% SDS. Adjust to pH 8.3 1X Transfer buffer (wet): 25 mM Tris-base, 192 mM glycine, 20% methanol, Adjust to pH 8.3 TBS TBST, TBS and 0.1% Tween Blocking solution: TBST, 5% non-fat dry milk rabbit anti-LC3 primary antibody (NB100-2331) in blocking buffer (~2 ug/mL)

Tip: For more information on Western Blotting, see our Western Blot handbook.

1. Grow cells (e.g. HeLa or Neuro2A) in vitro to semi-confluency (70-75%).

2. Add CQ to culture dishes to a final concentration of 50 uM and incubate overnight (16 hours). Remember to include an untreated sample as a negative control. Note: Validated autophagy inducers should be included as positive controls.

3. Rinse cells with ice-cold 1X PBS and lyse cells with sample buffer.

Note: LC3-I and LC3-II are sensitive to degradation, although LC3-I is more labile. These proteins are sensitive to freeze-thaw cycles and SDS sample buffers. Fresh samples should be analyzed quickly to prevent protein degradation.

4. Sonicate and incubate cells for 5 minutes at 95oC.

Tip: Cells are lysed directly in sample buffer or may be lysed in RIPA buffer.

5. Load samples of Chloroquine-treated and -untreated cell lysates 40 ug/lane on a 4-20% polyacrylamide gradient gel (SDS-PAGE).

Tip: For detection of LC3 it is particularly important to monitor the progress of the gel as this protein is relatively small (~14kDa).

Tip: Alternatively, for non-gradient gels, use a 20% polyacrylamide gel.

6. Transfer proteins to a 0.2 um PVDF membrane for 30 minutes at 100V.

7. After transfer, rinse the membrane with dH2O and stain with Ponceau S for 1-2 minutes to confirm efficiency of protein transfer.

8. Rinse the membrane in dH2O to remove excess stain and mark the loaded lanes and molecular weight markers using a pencil.

9. Block the membrane using blocking buffer solution (5% non-fat dry milk in TBST) for 1 hour at room temperature.

10. Rinse the membrane with TBST for 5 minutes.

11.Dilute the rabbit anti-LC3 primary antibody (NB100-2331) (~2 ug/mL) in blocking buffer and incubate the



membrane for 1 hour at room temperature.

12.Rinse the membrane with dH2O.

13. Rinse the membrane with TBST, 3 times for 10 minutes each.

14.Incubate the membrane with diluted secondary antibody, according with product's specifications, (e.g. anti-rabbit-IgG HRP-conjugated) in blocking buffer for 1 hour at room temperature.

Note: Tween-20 may be added to the blocking or antibody dilution buffer at a final concentration of 0.05-0.2%, provided it does not interfere with antibody-antigen binding.

15. Rinse the membrane with TBST, 3 times for 10 minutes each.

16.Apply the detection reagent of choice (e.g. BioFX Super Plus ECL) in accordance with the manufacturer's instructions.

17.Image the blot.

Tip: LC3-I and it's lipidated form LC3-II have different electrophoretic mobility properties, with the lipidated form moving faster in an SDS-PAGE gel, albeit its larger molecular weight. LC3-II runs at 14-16 kDa while LC3-I runs at 16 -18kDa.

Note: This assay measures the difference in the LC3-II signal in the presence and absence of inhibitors (e.g., lysosomotropic agents). When autophagic flux is present or induced in a system an increase in the LC3-II signal should be observed with the inhibitor.

Immunohistochemistry-Paraffin protocol for LC3A Antibody (NB100-2331)

LC3A Antibody:

I. Deparaffinization:

A. Treat slides with Xylene: 3 changes for 5 minutes each. Drain slides for 10 seconds between changes.

B. Treat slides with 100% Reagent Alcohol: 3 changes for 5 minutes each. Drain slides for 10 seconds between changes.

II. Quench Endogenous Peroxidase:

To Prepare 200 ml of Quenching Solution: Add 3 ml of 30% Hydrogen Peroxide to 200 ml of Methanol. **Use within 4 hours of preparation

A.Place slides in peroxidase guenching solution: 15-30 minutes.

III. Retrieve Epitopes:

A. Preheat Citrate Buffer. Place 200 ml of Citrate Buffer Working Solution into container, cover and place into steamer. Heat to 90-96C.

B. Place rack of slides into hot Citrate Buffer for 20 minutes. Cover.

C. Carefully remove container with slides from steamer and cool on bench, uncovered, for 20 minutes.

D. Slowly add distilled water to further cool for 5 minutes.

E. Rinse slides with distilled water. 2 changes for 2 minutes each.

IV. Immunostaining Procedure:

A. Remove each slide from rack and circle tissue section with a hydrophobic barrier pen (e.g. Liquid Blocker-Super Pap Pen).

B. Flood slide with Wash Solution.

**Do not allow tissue sections to dry for the rest of the procedure.

C. Drain wash solution and apply 4 drops of Blocking Reagent to each slide and incubate for 15 minutes.

D. Drain Blocking Reagent (do not wash off the Blocking Reagent), apply 200 ul of Primary Antibody solution to each slide, and incubate for 1 hour.

E. Wash slides with Wash Solution: 3 changes for 5 minutes each.

F. Drain wash solution, apply 4 drops of Secondary antibody to each slide and incubate for 1 hour.

G. Wash slides with Wash Solution: 3 changes for 5 minutes each.

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H. Drain wash solution, apply 4 drops of DAB Substrate to each slide and develop for 5-10 minutes.

Check development with microscope.



I. Wash slides with Wash Solution: 3 changes for 5 minutes each.

J. Drain wash solution, apply 4 drops of Hematoxylin to each slide and stain for 1-3 minutes.

Increase time if darker counterstaining is desired.

K. Wash slides with Wash Solution: 2-3 changes for 2 minutes each.

L. Drain wash solution and apply 4 drops of Bluing Solution to each slide for 1-2 minutes.

M. Rinse slides in distilled water.

N. Soak slides in 70% reagent alcohol: 3 minutes with intermittent agitation.

O. Soak slides in 95% reagent alcohol: 2 changes for 3 minutes each with intermittent agitation.

P. Soak slides in 100% reagent alcohol: 3 changes for 3 minutes each with intermittent agitation. Drain slides for 10 seconds between each change.

Q. Soak slides in Xylene: 3 changes for 3 minutes each with intermittent agitation. Drain slides for 10 seconds between each change.

R. Apply 2-3 drops of non-aqueous mounting media to each slide and mount coverslip.

S. Lay slides on a flat surface to dry prior to viewing under microscope.

NOTES:

- Use treated slides (e.g. HistoBond) to assure adherence of FFPE sections to slide.

- Prior to deparaffinization, heat slides overnight in a 60C oven.

- All steps in which Xylene is used should be performed in a fume hood.

- For Epitope Retrieval, a microwave or pressure cooker may be substituted for the steamer method. Adjust times as necessary depending on conditions.

- For the initial IHC run with a new primary antibody, test tissues with and without Epitope Retrieval. In some instances, Epitope Retrieval may not be necessary.

- 200 ul is the recommended maximum volume to apply to a slide for full coverage. Using more than 200 ul may allow solutions to wick off the slide and create drying artifacts.

- For small tissue sections less than 200 ul may be used.

- 5 minutes of development with DAB Substrate should be sufficient. Do not develop for more than 10 minutes. If 5 minutes of development causes background staining,

further dilution of the primary antibody may be necessary.

-Hematoxylin should produce a light nuclear counterstain so as not to obscure the DAB staining. Counterstain for 1-1.5 minutes for nuclear antigens. Counterstain for 2-3 minutes for cytoplasmic and membranous antigens. If darker counterstaining is desired increase time (up to 10 minutes).





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