

# Product Datasheet

## SHP/NR0B2/Nuclear Receptor SHP Antibody NLS5393

Unit Size: 0.05 ml

Store at 4C short term. Aliquot and store at -80C long term. Avoid freeze-thaw cycles.

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**NLS5393****SHP/NR0B2/Nuclear Receptor SHP Antibody**

<b>Product Information</b>	
<b>Unit Size</b>	0.05 ml
<b>Concentration</b>	1.0 mg/ml
<b>Storage</b>	Store at 4C short term. Aliquot and store at -80C long term. Avoid freeze-thaw cycles.
<b>Clonality</b>	Polyclonal
<b>Preservative</b>	0.01% Sodium Azide
<b>Purity</b>	Immunogen affinity purified
<b>Buffer</b>	PBS
<b>Product Description</b>	
<b>Host</b>	Rabbit
<b>Gene ID</b>	8431
<b>Gene Symbol</b>	NR0B2
<b>Species</b>	Human
<b>Reactivity Notes</b>	Human.
<b>Specificity/Sensitivity</b>	Nuclear Receptor SHP
<b>Immunogen</b>	Synthetic peptide [KLH conjugated] made to the ligand binding domain
<b>Product Application Details</b>	
<b>Applications</b>	Immunohistochemistry, Immunohistochemistry-Paraffin
<b>Recommended Dilutions</b>	Immunohistochemistry, Immunohistochemistry-Paraffin assay dependent

## Procedures

### Immunohistochemistry Protocol for Nuclear Receptor SHP Antibody (NLS5393)

#### Immunohistochemistry

1. Prepare tissue with formalin fixation and by embedding it in paraffin wax.
2. Make 4-um sections and place on pre-cleaned and charged microscope slides.
3. Heat in a tissue-drying oven for 45 minutes at 60 degrees Celcius.
4. Deparaffinize the tissues by wash drying the slides in 3 changes of xylene approximately 5 minutes each @ RT.
5. Rehydrate the tissues by washing the slides in 3 changes of 100% alcohol approximately 3 minutes each @ RT.
6. Wash the slides in 2 changes of 95% alcohol approximately 3 minutes each @ RT.
7. Wash the slides in 1 change of 80% alcohol approximately 3 minutes @ RT.
8. Rinse the slides in gentle running distilled water approximately 5 minutes @ RT.
9. Perform antigen retrieval by steaming the slides in 0.01M sodium citrate buffer (pH 6.0) @ 99-100 degrees Celcius for 20 minutes.
10. Remove the slides from the heat and let stand in buffer @ RT for 20 minutes.
11. Rinse the slides in 1X TBS-T for 1 minute @ RT.

**\*\*Do not allow the tissues to dry at any time during the staining procedure\*\***

12. Begin the immunostaining by applying a universal protein block approximately 20 minutes @ RT.
13. Drain protein block from the slides and apply the diluted primary antibody approximately 45 minutes @ RT.
14. Rinse the slide in 1X TBS-T approximately 1 minute @ RT.
15. Apply a biotinylated anti-rabbit IgG (H+L) secondary approximately 30 minutes @ RT.
16. Rinse the slide in 1X TBS-T approximately 1 minute at RT.
17. Apply an alkaline phosphatase streptavidin approximately 30 minutes at RT.
18. Rinse the slide in 1X TBS-T approximately 1 minute at RT.
19. Apply an alkaline phosphatase chromagen substrate approximately 30 minutes at RT.
20. Rinse the slide in distilled water approximately 1 minute @ RT.

**\*\*This method should only be used if the chromagen substrate is alcohol insoluble (ie: Vector Red, DAB)\*\***

21. Dehydrate the tissue by washing the slides in 2 changes of 80% alcohol approximately 1 minute each @ RT.
22. Wash the slides in 2 changes of 95% alcohol approximately 1 minute each @ RT.
23. Wash the slides in 3 changes of 100% alcohol approximately 1 minute each @ RT.
24. Wash the slides in 3 changes of xylene approximately 1 minute each @ RT.
25. Apply cover slip.



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### **Limitations**

This product is for research use only and is not approved for use in humans or in clinical diagnosis. Primary Antibodies are guaranteed for 1 year from date of receipt.

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