

Product Datasheet

ATG5 Antibody - BSA Free NB110-53818SS

Unit Size: 0.025 ml

Store at 4C short term. Aliquot and store at -20C long term. Avoid freeze-thaw cycles.

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NB110-53818SS

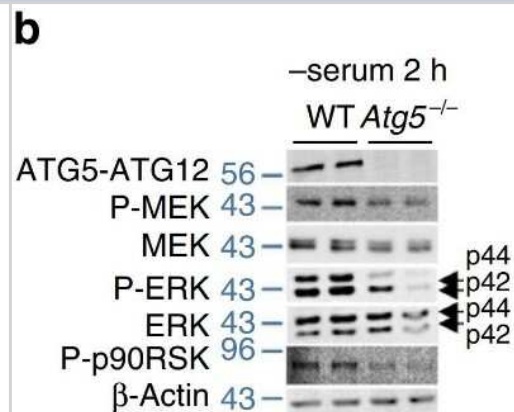
ATG5 Antibody - BSA Free

Product Information	
Unit Size	0.025 ml
Concentration	1.0 mg/ml
Storage	Store at 4C short term. Aliquot and store at -20C long term. Avoid freeze-thaw cycles.
Clonality	Polyclonal
Preservative	0.02% Sodium Azide
Isotype	IgG
Purity	Immunogen affinity purified
Buffer	PBS
Target Molecular Weight	32 kDa
Product Description	
Description	Novus Biologicals Knockout (KO) Validated Rabbit ATG5 Antibody - BSA Free (NB110-53818) is a polyclonal antibody validated for use in IHC, WB, ELISA, Flow, ICC/IF, Simple Western and IP. Anti-ATG5 Antibody: Cited in 284 publications. All Novus Biologicals antibodies are covered by our 100% guarantee.
Host	Rabbit
Gene ID	9474
Gene Symbol	ATG5
Species	Human, Mouse, Rat, Porcine, Alligator, Bovine, Drosophila, Fish, Guinea Pig, Primate, Xenopus, Zebrafish
Reactivity Notes	Drosophila reactivity reported in scientific literature (PMID:33221768) Fish reactivity reported in scientific literature (PMID: 26183773). Guinea Pig reactivity reported in scientific literature (PMID: 30766882). Use in Alligator reported in scientific literature (PMID:32061056).
Specificity/Sensitivity	This is selective for the full-length and calpain cleaved isoform proteins. The short isoform is missing amino acids 1-79. The calpain cleaved form of ATG5 is missing amino acids 195-275.
Immunogen	This ATG5 Antibody was made to a synthetic peptide of an N-terminal region of the human ATG5 protein (within residues 1-50) [Swiss-Prot Q9H1Y0].
Product Application Details	
Applications	Western Blot, Simple Western, Immunohistochemistry-Paraffin, ELISA, Electron Microscopy, Flow Cytometry, Immunoblotting, Immunocytochemistry/ Immunofluorescence, Immunohistochemistry, Immunoprecipitation, Proximity Ligation Assay, Radioimmunoassay, Knockdown Validated, Knockout Validated
Recommended Dilutions	Western Blot 1:500, Simple Western 1:50, Flow Cytometry, ELISA, Immunohistochemistry 1:400, Immunocytochemistry/ Immunofluorescence 1:250, Immunoprecipitation 1:10 - 1:500, Immunohistochemistry-Paraffin 1:400, Immunoblotting, Radioimmunoassay, Proximity Ligation Assay, Electron Microscopy, Knockout Validated, Knockdown Validated



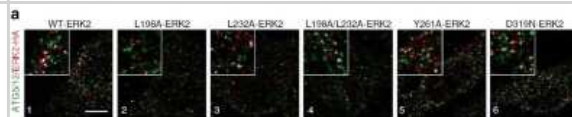
Images

Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - ERK phosphorylation in *Atg5*^{-/-} cells depends on nutrient availability. Serum-deprived *Atg5*^{-/-} MEFs display decreased ERK and p90RSK phosphorylation. Immunoblots for the indicated proteins in total lysates from 2 h serum-deprived WT and *Atg5*^{-/-} MEFs. The bars represent mean \pm s.e.m. **P* < 0.05, ****P* < 0.001 compared with Con; Student's t-test, *n* = 3. Image collected and cropped by CiteAb from the following publication (<https://www.nature.com/articles/ncomms3799>), licensed under a CC-BY license.

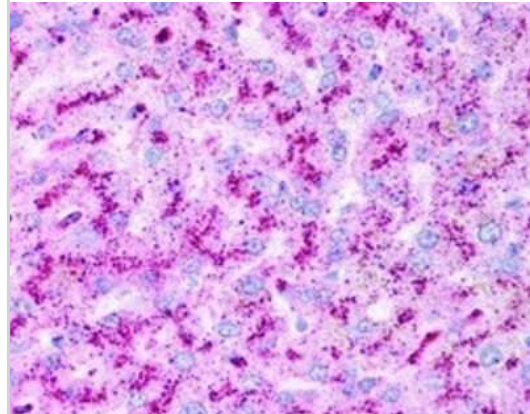


Immunocytochemistry/Immunofluorescence: ATG5 Antibody - BSA Free [NB110-53818] - ERK2 utilizes kinase-docking domains to interact with ATG5-ATG12 and LC3-II. (a) Mutations in FRS on ERK2 decrease colocalization of ERK2 with ATG5-ATG12, LC3 and WIPI1.

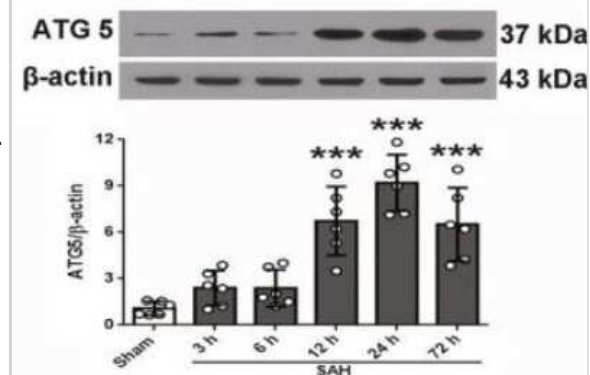
Immunofluorescence (IF) showing colocalization (depicted as white pixels by colocalization finder application) of WT-ERK2-HA, FRS ERK2 mutants (L198A-, L232A-, L198A/L232A-, Y261A-ERK2-HA) or common docking (CD) mutant (D319N-ERK2-HA) with ATG5-ATG12 (panels 1-6), LC3 (panels 7-12) or WIPI1 (panels 13-18) in EGF-treated NIH/3T3 cells. ERK2 is stained in red, and autophagy proteins are stained in green. Scale bar, 10 μ m. The bars represent mean \pm s.e.m. ***P* < 0.01, *****P* < 0.0001 compared with WT-ERK2-transfected cells; Student's t-test, 50 cells analysed from *n* = 2. Image collected and cropped by CiteAb from the following publication (<https://www.nature.com/articles/ncomms3799>), licensed under a CC-BY license.



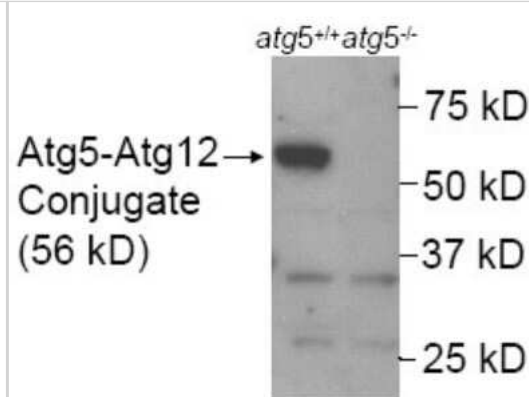
Immunohistochemistry: ATG5 Antibody - BSA Free [NB110-53818] - Immunohistochemistry analysis of human liver hepatocytes at 2.5 μ g/mL. 40X magnification.



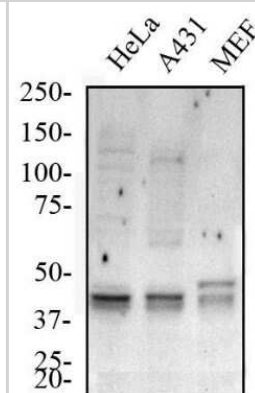
Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - Representative Western blot images and quantitative analyses of ATG5 from the left hemisphere of rat brains at different time points after SAH. Sample size is 36, *n* = 6 per group. Data were presented as mean \pm SD. *F* = 22.14 for ATG5. **P* < .05, ** *P* < .01, ****P* < .001 vs Sham group. SAH, subarachnoid hemorrhage; Vehicle, phosphate-buffered saline; rOPN, recombinant OPN. NS, not significant. Image collected and cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/31436915/>) licensed under a CC-BY license.



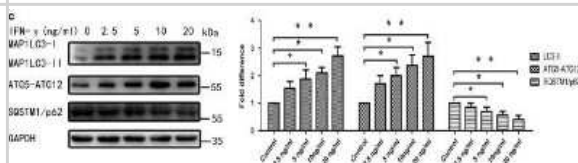
Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - Western blot analysis in mouse wildtype ES cell lysate (Lane 1) using NB110-53818. Lane 2 is a mouse ATG5 KO ES cell lysate (negative control). Atg5^{-/-} ES cells from Dr. Noboru Mizushima [Mizushima, N. et al. J. Cell Biol. 152 (2001)] Photo courtesy of Dr. Beth Levine, UT Southwestern Medical Center.



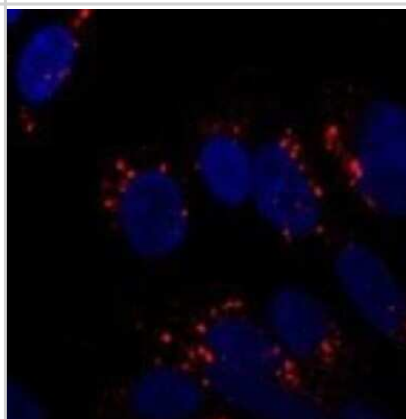
Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - Western blot analysis of total protein from Human HeLa and A431 and Mouse MEF cells. Lysates were separated on a 7.5% gel by SDS-PAGE, transferred to PVDF membrane and blocked in 5% non-fat milk in TBST. The membrane was probed with 2.0 ug/mL anti-ATG5 in 1% non-fat milk in TBST and detected with an anti-rabbit HRP secondary antibody using chemiluminescence.



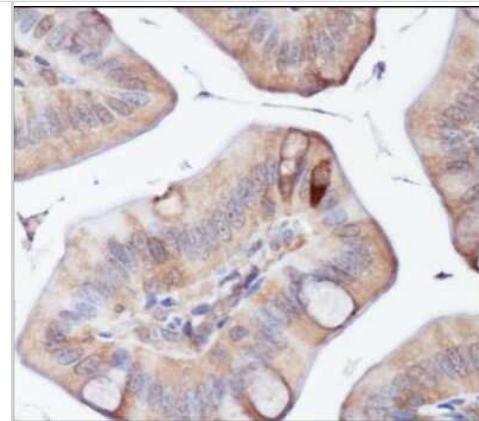
Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - IFN-gamma increased rates of autophagy in primary BMECs in vitro. BMECs were treated with IFN-gamma at 2.5, 5, 10 or 20 ng/mL. At the end of the treatment, levels of MAP1LC3, ATG12-ATG5, SQSTM1/p62 and GAPDH were analyzed using Western blot analysis with specific antibodies as described in the Materials and Methods section. The data represent the mean \pm S.E.M. of three independent experiments. Error bars are \pm S.E.M. One-way ANOVA; *P<0.05; **P<0.01. Image collected and cropped by CiteAb from the following publication (<https://www.nature.com/articles/cddiscovery201565>) licensed under a CC-BY license.



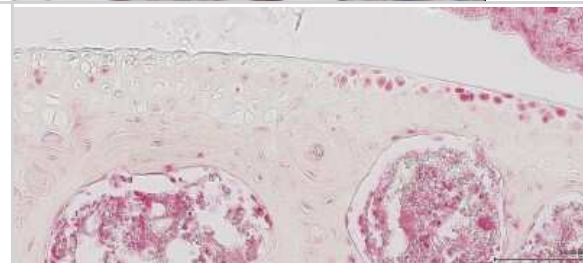
Immunocytochemistry/Immunofluorescence: ATG5 Antibody - BSA Free [NB110-53818] - ICC/IF analysis of SY5Y cells at 1:250. Incubated overnight at 4 degrees. Photo courtesy of an anonymous collaborator.



Immunohistochemistry: ATG5 Antibody - BSA Free [NB110-53818] - Immunohistochemistry analysis of mouse intestine using DAB with hematoxylin counterstain.



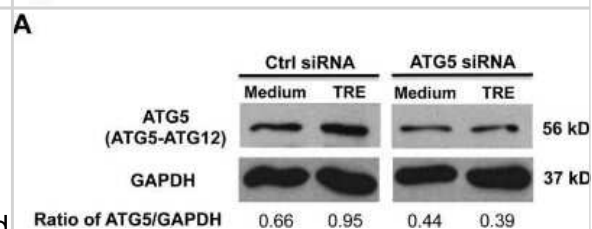
Immunohistochemistry-Paraffin: ATG5 Antibody - BSA Free [NB110-53818] - Immunohistochemistry analysis of mouse knee. IHC-P image submitted by a verified customer review.



Simple Western: ATG5 Antibody - BSA Free [NB110-53818] - Simple Western lane view shows a specific band for ATG5 in 0.5 mg/mL of HeLa lysate. This experiment was performed under reducing conditions using the 12-230 kDa separation system.

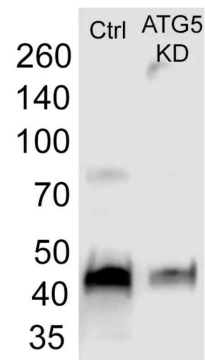


Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - Knockdown of autophagy-related gene 5 (ATG5) inhibits trehalose-induced autophagy in normal human primary airway epithelial cells. Normal human tracheobronchial epithelial cells were transfected with Naito1 chimera RNAi (control siRNA) or ATG5 chimera siRNA (ATG5 siRNA). Twenty-four hours after siRNA transfection, cells were treated with medium or trehalose (TRE, 100 mM) for 48 h. ATG5 protein as examined by Western blot analysis with GAPDH protein used as loading control. The representative Western blot picture was shown from 2 independent experiments with each being performed in triplicate wells.

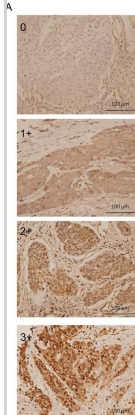


Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - Western Blot of the lysate of DU145 cells - control and ATG5 KD. Image from verified customer review.

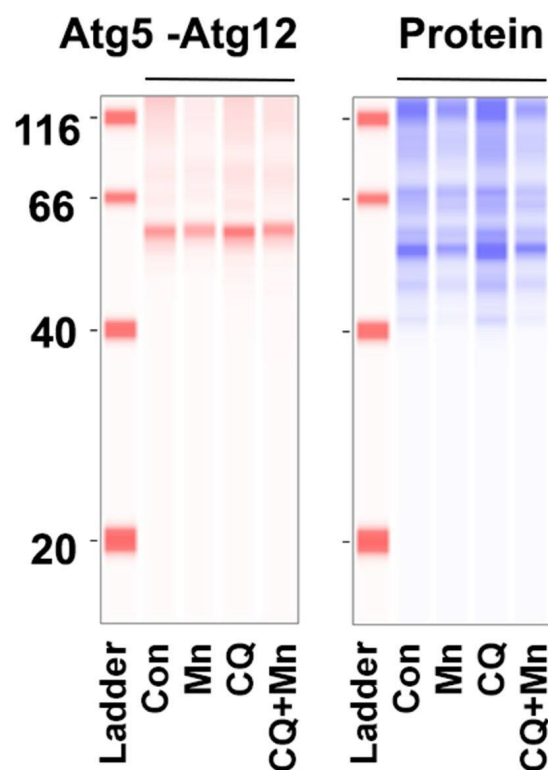
MW, KDa



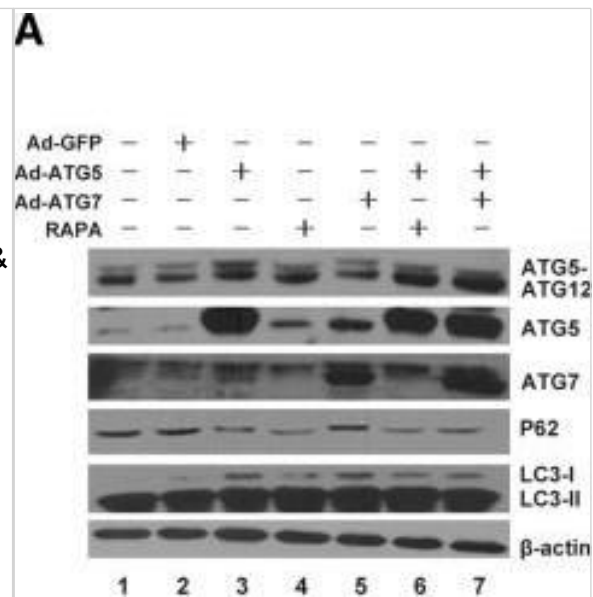
(A) ATG5 expression in ESCC tissue was analyzed by IHC and scored as 0, 1+, 2+, and 3+. (B) Expression level of ATG5 in adjacent non-cancerous (normal) and early-stage ESCC tissues by IHC. (C-D) Kaplan–Meier estimates of OS (C) and PFS (D) by the expression levels of ATG5 (low and high) of adjacent normal tissue from early-stage ESCC patients. MST: median survival time. Low, expression score 0 or 1+; High, expression score 2+ or 3+.



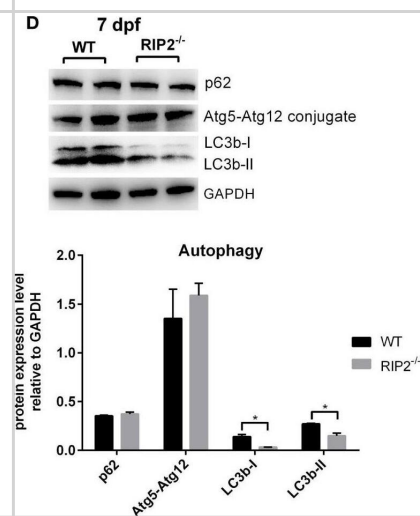
Simple Western: ATG5 Antibody - BSA Free [NB110-53818] - Dose-responses of Mn on cell viability and autophagy flux. (A) Stable HeLa cells overexpressing mRFP-GFP-LC3, and immortalized rat dopaminergic neuronal cells N27 were exposed to MnCl₂ (62.5 uM to 2mM) for 24h, or (B) 48h. Cell viability was evaluated using Calcein AM dye. Data represent mean \pm SEM, n = 3 independent experiments with 4 replicates per experiment. (C) Schematic diagram illustrating the autophagy flux pathway and the construct used to create the mRFP-GFP-LC3 stable HeLa reporter cells. With this cell model, autophagosomes appear yellow due to the colocalization of RFP and GFP signals. Red signal indicates the flux is functional because the green signal is quenched by the acidic environment of the lysosomes, which fuse with autophagosomes. These stable cells were treated with vehicle control or Mn (15.6-250 uM) for 24h. (D) Representative images of cells treated with different doses of Mn were captured using confocal microscopy. (E) Green and red vesicles per cell were quantified using Fiji. Green vesicles represent autophagosomes. The number of autolysosomes was calculated by subtracting the number of green from the red puncta per cell. Scale bar = 20um. (F) Stable HeLa cells were treated with MnCl₂ (125 uM), chloroquine (CQ) or both. (G) The number of autophagosomes and autolysosomes were quantified. The combination of Mn with CQ did not significantly alter the numbers of autophagosome (P = 0.9941) and autolysosome (P > 0.9999), as compared to CQ (50 uM) only, a dose that did not completely inhibit autophagy. Data represent mean \pm SEM. (H) N27 cells were treated with 62.5 or 125 uM of MnCl₂ for 24h, then incubated with LysoSensor Yellow/Blue. The ratio of acidic (yellow) vs. neutral (blue) pH of lysosomes were quantified using a plate reader. (I) N27 cells were treated with Mn 125 uM, CQ50 uM or both for 24h, followed by immunoblotting for Atg5, a marker for autophagosomes, using Jess (ProteinSimple, Inc.) (J) Total protein per capillary was used as loading control to normalize the levels of Atg5. Mn did not further increase the accumulation of autophagosomes when combined with CQ versus CQ alone (P = 0.9998). All data represent mean \pm SEM (n = 3-4 independent experiments with approximately 30 cells per experiment for E and G) and groups were compared by using Kruskal-Wallis ANOVA test with Dunn's post-hoc test; ns: non-significant. Image collected and cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/24957904>), licensed under a CC-BY license. Not internally tested by R&D Systems. Image collected and cropped by CiteAb from the following publication (<https://molecularneurodegeneration.biomedcentral.com/articles/10.1186/s13024-024-00708-w>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



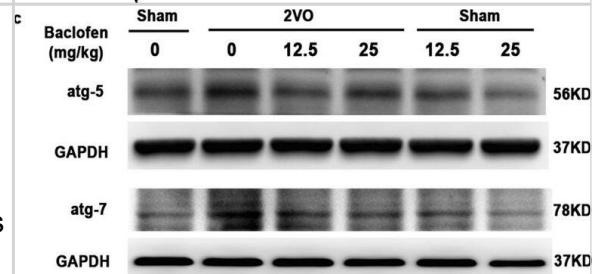
ATG5 & ATG7 enhanced autophagy & inhibited ER stress in chondrocyte. a Western blotting analysis of LC3, P62, ATG5, ATG7 & ATG5-ATG12 expression after infected with Ad-ATG5, Ad-ATG7 & Ad-ATG5 + Ad-ATG7 in the C28I2 cells. β -actin is served as an internal control. b Qualitative analysis of ATG5, ATG7, ATG5-ATG12, LC3 & P62. The values were normalized to β -actin. c C28I2 cells were double stained with LC3 (red) & DAPI (blue) & visualized by confocal microscopy (400X) after treated with Rapamycine, Ad-ATG5, Ad-ATG7 & Ad-ATG5 + Ad-ATG7 24 h. * $P < 0.05$, ** $P < 0.01$ compared with the controls. Values are means \pm SD $n = 3$). d Qualitative analysis of LC3 fluorescence intensity of chondrocytes. The values were normalized to the NC group. e Western blotting analysis of PERK, p-PERK & Nrf2 expression after infected with Ad-ATG5, Ad-ATG7 & Ad-ATG5 + Ad-ATG7 in the C28I2 cells. β -actin is served as an internal control. f Qualitative analysis of PERK, p-PERK & Nrf2 were normalized to β -actin. (1:NC, 2:Ad-GFP, 3:Ad-ATG5, 4:RAPA, 5:Ad-ATG7, 6:Ad-ATG5 + RAPA, 7:Ad-ATG5 + Ad-ATG7). Rapamycin (25 μ M) used as a positive control Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/31060556>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



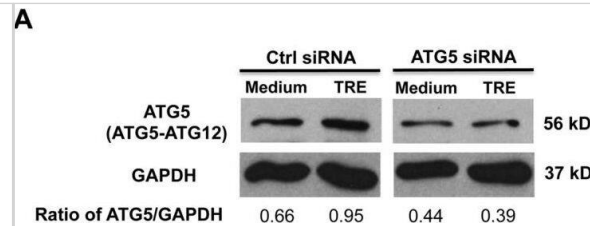
Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - RIP2 is not essential for MAPK pathways in the early ontogenesis. (A) Immunoblot analysis of phospho-p44/42 MAPK, p44/42 MAPK, & p38 MAPK in larvae homogenate from WT & NOD1-1IS^{-/-} zebrafish at 7 days post-fertilization (dpf). (B) Immunoblot analysis of phospho-p44/42 MAPK, p44/42 MAPK, & p38 MAPK in larvae homogenate from WT & RIP2^{-/-} zebrafish at 7 dpf. (C) Immunoblot analysis of Atg5, p62, & LC3b in larvae homogenate from WT & NOD1-1IS^{-/-} zebrafish at 7 dpf. (D) Immunoblot analysis of Atg5, p62, & LC3b in larvae homogenate from WT & RIP2^{-/-} zebrafish at 7 dpf. Western blotting results were quantified using Quantity One software. Data represent the average of two independent experiments. * $p < 0.05$, ** $p < 0.01$. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/29692779>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



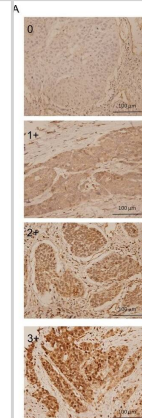
Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - Baclofen reversed the changes of protein markers characteristic for autophagy in hippocampal CA1 area under chronic cerebral hypoperfusion. (a-c) Five weeks after induction of hypoperfusion, p-mTOR was significantly decreased, & LC3-II, Beclin 1, atg5 & atg7 were significantly increased, & baclofen could reverse the changes of these proteins expression. Treatment with baclofen at 12.5 mg/kg & 25 mg/kg in sham-operated rats did not change the expression of LC3-II, mTOR, p-mTOR, Beclin 1, atg5 & atg7 compared with sham-operated rats ($n = 4$ in each group). Blots shown have been cropped to fit space requirements & run under the same experimental conditions. * $P < 0.05$ & ** $P < 0.01$ vs sham-operated rats; ### $P < 0.01$ vs 2VO rats. Image collected & cropped by CiteAb from the following publication (<https://www.nature.com/articles/srep14474>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



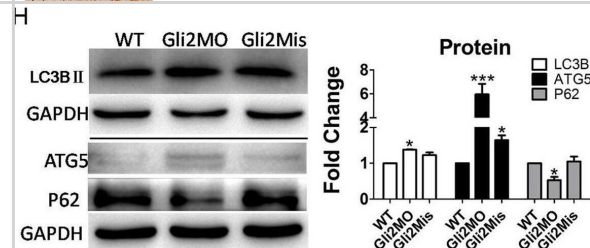
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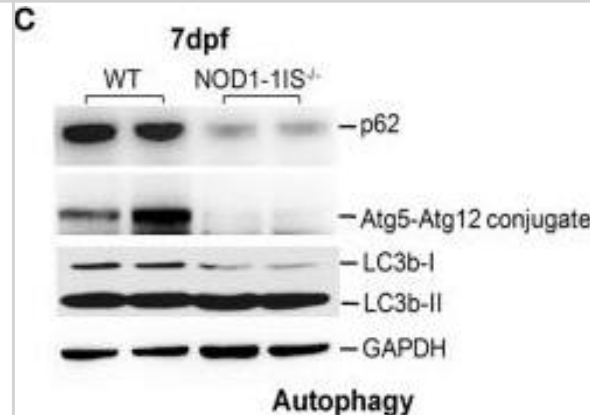
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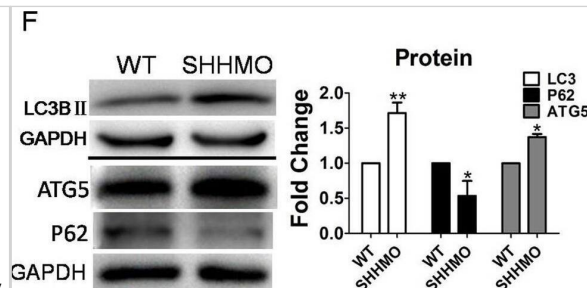
Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - Autophagy level was promoted by downregulation of the Hh signaling pathway in zebrafish larvae. (A–C) Larvae were exposed to 10 μ M cyA & 0.1% DMSO at 3 dpf for 3 days. (D–H) Embryos were injected with SHHMO, Gli2MO & Gli2Mis at the 1–4 cell stage; at 6 dpf, larvae were collected for assays. GAPDH served as a loading control. The means & s.d. were derived from triple duplicates. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ versus the WT groups or DMSO control groups (one-way ANOVA). The data were from three independent experiments. (A,D) SHH protein was detected by western blot assay. (B,E,G) Transcriptional level of autophagy-related genes was tested by qPCR assays. (C,F,H) Autophagy-related proteins LC3B II, ATG5 & P62 level were determined by western blot. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/30992325>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



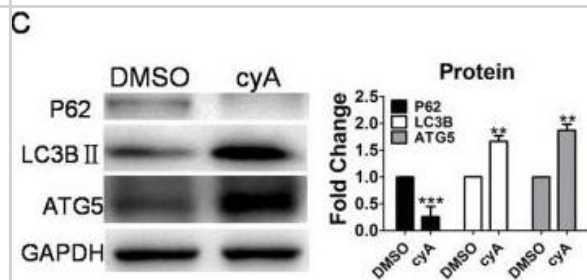
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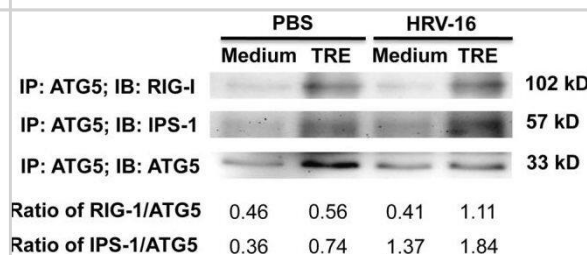
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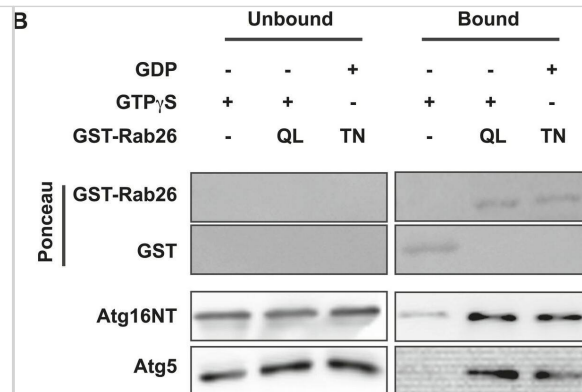
Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - Autophagy level was promoted by downregulation of the Hh signaling pathway in zebrafish larvae. (A–C) Larvae were exposed to 10 μ M cyA & 0.1% DMSO at 3 dpf for 3 days. (D–H) Embryos were injected with SHHMO, Gli2MO & Gli2Mis at the 1–4 cell stage; at 6 dpf, larvae were collected for assays. GAPDH served as a loading control. The means & s.d. were derived from triple duplicates. * P <0.05, ** P <0.01, *** P <0.001 versus the WT groups or DMSO control groups (one-way ANOVA). The data were from three independent experiments. (A,D) SHH protein was detected by western blot assay. (B,E,G) Transcriptional level of autophagy-related genes was tested by qPCR assays. (C,F,H) Autophagy-related proteins LC3B II, ATG5 & P62 level were determined by western blot. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/30992325>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



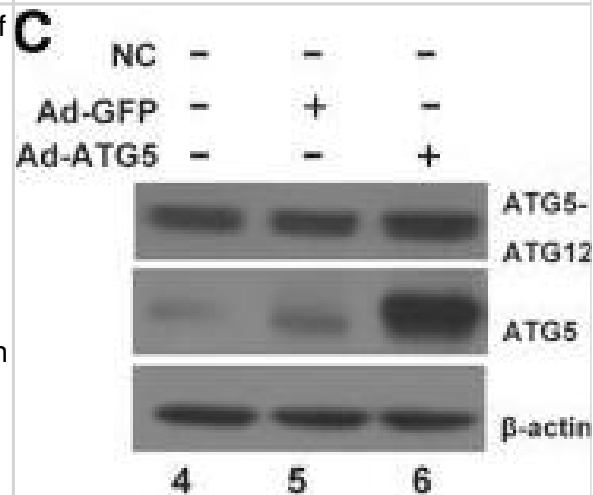
Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - Autophagy-related gene 5 (ATG5) protein interacts with retinoic acid-inducible gene I (RIG-I) & IFN- β promoter stimulator 1 (IPS-1) in normal human primary airway epithelial cells. Normal human tracheobronchial epithelial cells were treated with medium or trehalose (TRE, 100 mM) for 48 h & then infected with HRV-16 (104 TCID₅₀/well) for 2 h. After removing the free viruses, cells were incubated with medium or trehalose for additional 6 h. Pre-cleared cell lysates were incubated with a mouse anti-human ATG5 antibody, & immunoprecipitated proteins were separated on 10% SDS—PAGE for immunoblotting of RIG-I, IPS-1 & ATG5. The representative Western blot picture was shown from 2 independent experiments with each being performed in triplicate wells. Image collected & cropped by CiteAb from the following publication (<https://dx.plos.org/10.1371/journal.pone.0124524>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



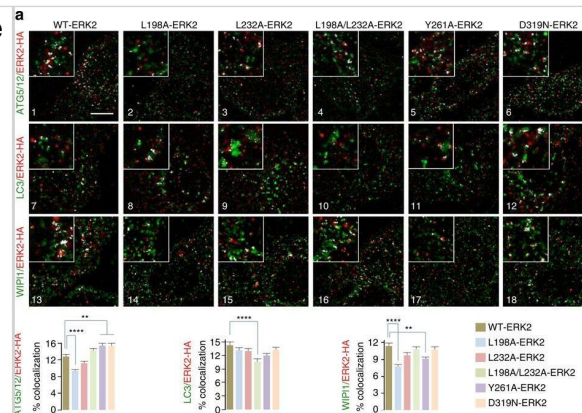
Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - Atg16 is an effector of GTP-Rab26. (A) Co-Immunoprecipitation of FLAG-tagged Rab26 variants expressed in HeLa cells with endogenous Atg16L1 protein. Immunoprecipitation was carried out following lysis in detergent-containing buffer & clearance by centrifugation to remove cell debris. Note that only the GTP-preferring QL variant of Rab26 showed significant binding to Atg16L1 (arrow). (B) GST pull-down of purified recombinantly expressed GST-Rab26 variants with a pre-formed complex of His-tagged versions of Atg5 & the N-terminal domain of Atg16L1 (Atg16NT). Note that Atg16NT selectively interacted with the GTP-preferring QL-variant of Rab26. DOI: <http://dx.doi.org/10.7554/eLife.05597.015> Analyses of Rab26 & ATG16L1 interaction by analytical gel filtration. To analyze complexes formed between Atg16L1 & Rab26 or Rab33, samples containing either Rab protein were pre-incubated with Atg16NT/Atg5 & loaded on a Superdex 200 10/30 GL column. As controls, Rab26, Rab33, or Atg16NT/Atg5 alone were separately loaded & analyzed. Eluted fractions were collected, analyzed by SDS-PAGE & protein bands visualized using Coomassie staining. (A) The elution profiles for various samples analyzed. (B–F) Representative fractions were separated on SDS PAGE gels & stained with Coomassie to visualize the protein bands. (B) Co-migration of Rab26QL with Atg16NT/Atg5 was not detected when pre-incubated samples of these proteins were tested. (C) Complexes of Rab33QL with Atg16NT/Atg5 can be observed to co-migrated (fractions 5–7, arrows). (D–F) Control runs were performed using only Rab26QL, Rab33QL or Atg16NT/Atg5. DOI: <http://dx.doi.org/10.7554/eLife.05597.016> Image collected & cropped by CiteAb from the following publication (<https://elifesciences.org/articles/05597>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - Expression of ER stress associated protein & autophagy related protein in the C28I2 chondrocyte infected with Ad-ATG5 or Ad-ATG7. a The ATG5 & ATG7 mRNA level were analysed by RT-PCR in C28I2 cells. b Analysis of ATG5 & ATG7 mRNA level with Q-PCR in C28I2 cells. Ad-ATG5(a), Ad-ATG7(b). c The ATG5 protein expression was analysed by western blotting in the Ad-ATG5 infected C28I2 cells. The level of ATG5 was normalized to β -actin. d The ATG7 protein expression was analysed by western blotting in the Ad-ATG7 infected C28I2 cells. The level of ATG7 was normalized to β -actin. e The expression levels of LC3, P62, PERK, p-PERK, Nrf2 proteins after infected with Ad-ATG5 in C28I2 cells for 24 h by western blotting. The levels of related proteins were normalized to β -actin. f The expression levels of LC3, P62, PERK, p-PERK, Nrf2 proteins after infected with Ad-ATG7 in C28I2 cells for 24 h by western blotting. The levels of related proteins were normalized to β -actin. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ compared with the controls. Values are means \pm SD $n = 3$). (1:NC, 2: Ad-GFP, 3:Ad-ATG5(a) & Ad-ATG7(b), 4:NC, 5:Ad-GFP, 6:Ad-ATG5, 7:Ad-ATG7) Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/31060556>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



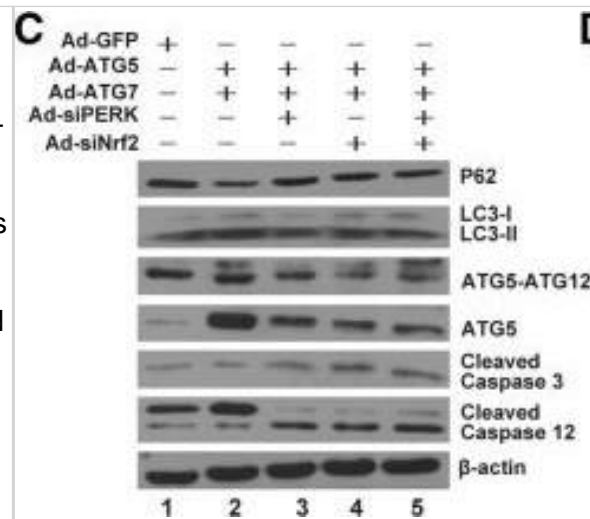
Immunocytochemistry/ Immunofluorescence: ATG5 Antibody - BSA Free [NB110-53818] - ERK2 utilizes kinase-docking domains to interact with ATG5–ATG12 & LC3-II. (a) Mutations in FRS on ERK2 decrease colocalization of ERK2 with ATG5–ATG12, LC3 & WIPI1. Immunofluorescence (IF) showing colocalization (depicted as white pixels by ‘colocalization finder application’) of WT-ERK2-HA, FRS ERK2 mutants (L198A-, L232A-, L198A/L232A-, Y261A-ERK2-HA) or common docking (CD) mutant (D319N-ERK2-HA) with ATG5–ATG12 (panels 1–6), LC3 (panels 7–12) or WIPI1 (panels 13–18) in EGF-treated NIH/3T3 cells. ERK2 is stained in red, & autophagy proteins are stained in green. Scale bar, 10 μ m. The bars represent mean \pm s.e.m. ** $P < 0.01$, **** $P < 0.0001$ compared with WT-ERK2-transfected cells; Student’s t-test, 50 cells analysed from $n=2$. (b) Mutations in FRS on ERK2 decrease colocalization of ERK2 with nuclear ATG5–ATG12 or LC3. IF showing colocalization (white pixels) of WT-ERK2-HA (red), FRS ERK2 mutants (L198A-, L232A-, L198A/L232A-, Y261A-ERK2-HA) or common docking (CD) mutant (D319N-ERK2-HA) with ATG5–ATG12 (panels 1–6) or LC3-II (panels 7–12) in EGF-treated NIH/3T3 cells. Scale bar, 5 μ m. The bars represent mean \pm s.e.m. ** $P < 0.01$, **** $P < 0.0001$ compared with WT-ERK2-transfected cells; Student’s t-test, 50 cells analysed from $n=2$. Arrows depict colocalization. (c) ATG5–ATG12 conjugation is required for ERK phosphorylation. Immunoblots for P-ERK, total ERK & β -actin in NIH/3T3 cells transfected with WT ATG5 or the conjugation-defective ATG5 K130R mutant & treated with EGF (10 min). The bars represent mean \pm s.e.m. * $P < 0.05$ compared with WT ATG5-transfected cells; Student’s t-test, $n=3$. Image collected & cropped by CiteAb from the following publication (<https://www.nature.com/articles/ncomms3799>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



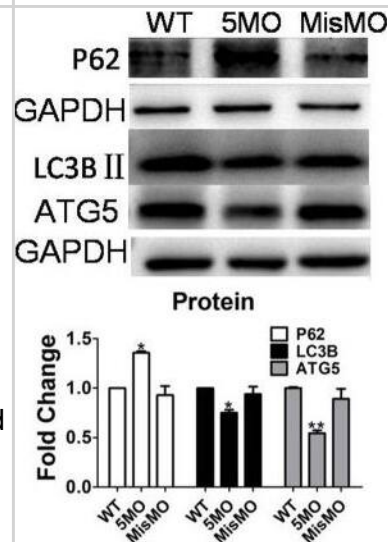
Immunocytochemistry/ Immunofluorescence: ATG5 Antibody - BSA Free [NB110-53818] - Components of the ERK signalling cascade associate with APh. (a) EGF enhances colocalization of phosphorylated (P)-bRAF, P-MEK & P-ERK with LC3. Immunofluorescence (IF) showing colocalization of P-bRAF (green), P-MEK (green) & P-ERK (green) with LC3 (red) in NIH/3T3 cells in presence or absence of EGF (10 min as shown in scheme). The bars represent mean \pm s.e.m. ** $P < 0.01$, **** $P < 0.0001$; Student’s t-test, 60 cells analysed from two experiments. Scale bars, 10 μ m. (b) P-ERK colocalizes with pre-autophagosomal & autophagosomal structures. IF depicting colocalization of P-ERK (green or red) with LC3 (red), ATG5–ATG12 (red), ATG16 (green), vps34 (red), WIPI1 (red), WIPI2 (red) & P-ULK1 (red) in EGF-treated NIH/3T3 cells. Scale bars, 10 μ m. The bars represent mean \pm s.e.m. 50 cells analysed from two experiments. (c) Growth factors in serum maintain P-ERK/LC3 colocalization. IF depicting colocalization of P-ERK (green) with LC3 (red) in 2 h serum-fed NIH/3T3 cells. (a–c) Native merged images or images with colocalization highlighted in white pixels are shown. Nuclei are blue (DAPI). Scale bar, 10 μ m. Image collected & cropped by CiteAb from the following publication (<https://www.nature.com/articles/ncomms3799>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



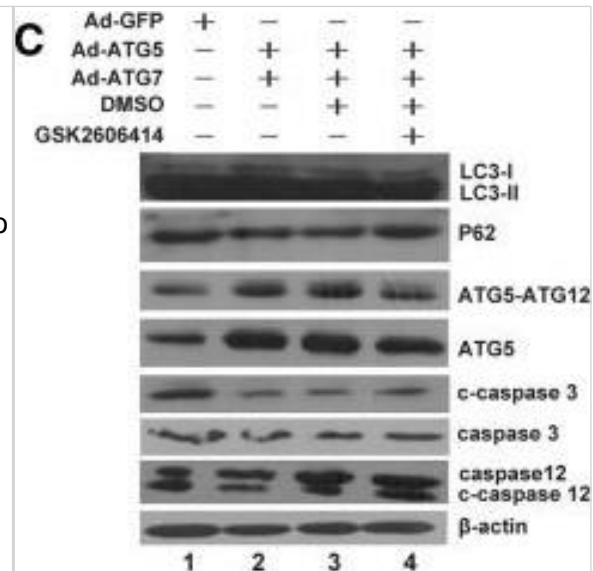
Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - ATG5 & ATG7 influenced on autophagy, apoptosis & cell cycle through PERK/Nrf2 signaling. a The ER stress proteins were analysed by western blotting in Ad-GFP, Ad-ATG5 + Ad-ATG7, Ad-ATG5 + Ad-ATG7 + siPERK, Ad-ATG5 + Ad-ATG7 + siNrf2 & Ad-ATG5 + Ad-ATG7 + siPERK + siNrf2 induced chondrocytes. b The levels of ER stress proteins were normalized to β -actin. c Determination of autophagy & apoptosis proteins expression by western blotting in Ad-GFP, Ad-ATG5 + Ad-ATG7, Ad-ATG5 + Ad-ATG7 + siPERK, Ad-ATG5 + Ad-ATG7 + siNrf2 & Ad-ATG5 + Ad-ATG7 + siPERK + siNrf2 induced chondrocytes. d The levels of related proteins were normalized to β -actin. e FCM analysis was used to calculate the percentage of apoptotic cells at the time point of 24 h. The apoptosis rate were increased when combined treatment with silencing of PERK or Nrf2. Experiments were repeated 3 times, Representative images are shown. f Analysis of cell apoptosis. Data come from 3 independent experiments. g FCM analysis indicated that the S phase percentage were decreased compared to that of the control groups when combine infected with silencing of PERK or Nrf2 in C2812 cells. Experiments were repeated 3 times, Representative images are shown. h Percentage of cells at each phase in different groups. * $P < 0.05$, ** $P < 0.01$ compared with the controls. Values are means \pm SD $n = 3$). (1:Ad-GFP, 2:Ad-ATG5 + Ad-ATG7, 3:Ad-ATG5 + Ad-ATG7 + siPERK, 4:Ad-ATG5 + Ad-ATG7 + siNrf2, 5:Ad-ATG5 + Ad-ATG7 + siPERK + siNrf2) Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/31060556>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



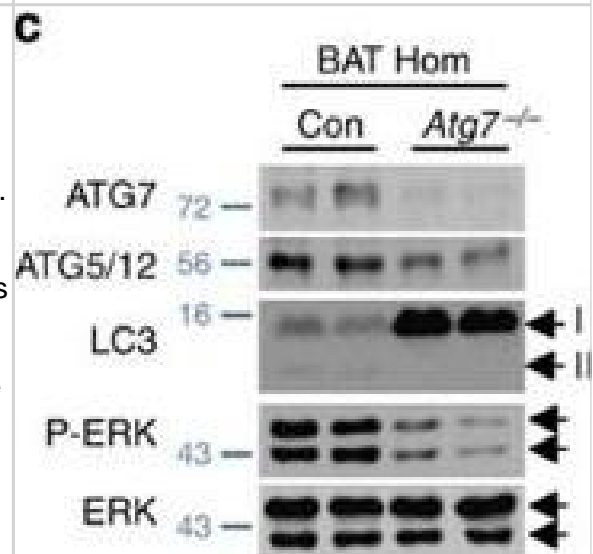
Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - Autophagy did not regulate the Hh signaling pathway. ATG5MO & MisMO were injected into embryos at the 1–4 cell stage. MisMO-injected group as a negative control. The means & s.d. were derived from triple duplicates. (A) Expression of LC3BII & ATG5 proteins was determined by a western blot assay. GAPDH served as a loading control. (B) Heat map of mRNA expression of Hh signaling pathway-related genes Smo, Gli family number Gli1, Gli2a, Gli2b, ptch1 & Shh mRNA from wild-type & ATG5MO-injected embryos was determined by RNAseq assay. (C) Embryos at 1–4 cell stage were either injected with ATG5MO or MisMO or mock-treated & treated with 10 μ M of RAPA or 2 mM of 3MA for 3 days from 3 dpf. Expression of SHH protein was determined by a western blot assay. GAPDH served as a loading control. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/30992325>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



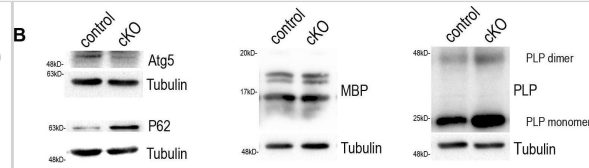
Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - The effect of ATG5 & ATG7 on autophagy, ER stress, apoptosis & cell cycle depends on PERK. a The ER stress proteins were analysed by western blotting in the GSK2606414 treated chondrocytes. b The levels of ER stress proteins were normalized to β -actin. c Determination of autophagy & apoptosis proteins expression by western blotting in the GSK2606414 treated chondrocytes. d The levels of related proteins were normalized to β -actin. e FCM analysis was used to calculate the percentage of apoptotic cells at the time point of 24 h. The apoptosis rate were increased when combined treated with GSK2606414. Experiments were repeated 3 times, Representative images are shown. f Analysis of cell apoptosis. Data come from 3 independent experiments. g FCM analysis indicated that the S phase percentage were decreased compared to that of the control groups when combine treated with GSK2606414 in C28I2 cells. Experiments were repeated 3 times, Representative images are shown. h Percentage of cells at each phase in different groups. * $P < 0.05$, ** $P < 0.01$ compared with the controls. Values are means \pm SD n = 3). (1:Ad-GFP, 2:Ad-ATG5 + Ad-ATG7, 3:Ad-ATG5 + Ad-ATG7 + DMSO, 4:Ad-ATG5 + Ad-ATG7+ GSK2606414) Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/31060556>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



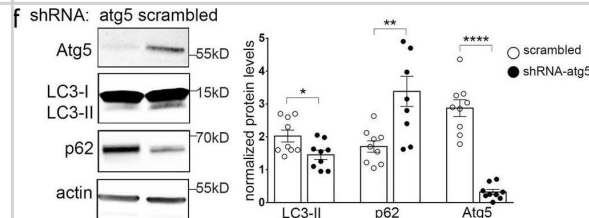
Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - ATG7/LC3-II regulates ERK phosphorylation.(a) *Atg7*^{-/-} livers display decreased ERK phosphorylation. Immunoblots for the indicated proteins in liver homogenate (Hom) & cytosolic (Cyt) fractions of control (Con) & liver-specific *Atg7*^{-/-} mice are shown. The bars represent mean \pm s.e.m. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ compared with Con; Student's t-test, n=4. (b) *Atg7*^{-/-} livers display decreased ERK2 dimers. Immunoblots show the indicated proteins in Hom from Con & *Atg7*^{-/-} livers. The bars represent mean \pm s.e.m. * $P < 0.05$, *** $P < 0.001$ compared to Con; Student's t-test, n=4-5. Arrows indicate p42 monomers & dimers. (c) *Atg7*^{-/-} brown adipose tissues (BAT) display decreased ERK phosphorylation. Immunoblots for indicated proteins in Hom from Con & *Atg7*^{-/-}-BAT. The bars represent mean \pm s.e.m. ** $P < 0.01$ compared with Con; Student's t-test, n=4-6. (d) *Atg7*^{-/-} livers exhibit decreased nuclear ERK phosphorylation. Immunoblots show the indicated proteins in homogenate (Hom, lane 1) & nuclear fractions (lanes 2-5) from Con & *Atg7*^{-/-} livers. The bars represent mean \pm s.e.m. *** $P < 0.001$ compared with Con; Student's t-test, n=4-5. (e) LC3 C terminus glycine-deleted (Δ G) mutants exhibit decreased ERK phosphorylation. Immunoblots for indicated proteins in total lysates from CFP-LC3- & CFP-LC3 Δ G-transfected NIH/3T3 cells exposed or not to EGF (10 min). The bars represent mean \pm s.e.m. *** $P < 0.001$ compared with Con; Student's t-test, n=8. Image collected & cropped by CiteAb from the following publication (<https://www.nature.com/articles/ncomms3799>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



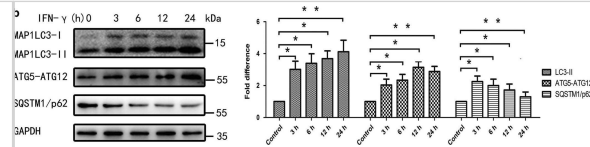
Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - FIGURE 2: Blockage of autophagy leads to increased plp mRNA & protein levels. (A) Schematic illustration of the experimental protocol used for tamoxifen (TMX) induction & analysis in the plpCreERT2; atg5f/f mice. All animals were injected i.p. with 1 mg of TMX per day at the age of 2.5 months for 10 days with two days break in between. The analysis was performed at 6 mo. (B) Western blot analysis with antibodies against Atg5, P62, PLP & MBP proteins in optic nerve lysates of 6mo control & cKO mice. Representative images of a single experiment are depicted. (C) Quantification of normalized protein levels of "B". (D) Quantification of quantitative real-time PCR in mouse optic nerves of 6mo control & cKO mice. Data information: Data are shown as mean \pm SEM. N=3-4 animals per genotype. Student's t-test was used to determine statistical significance between control & cKO protein & mRNA levels. * $p < 0.05$, ** $p \leq 0.01$. Image collected & cropped by CiteAb from the following publication (<http://www.cell-stress.com/researcharticles/2022a-ktena-cell-stress/>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



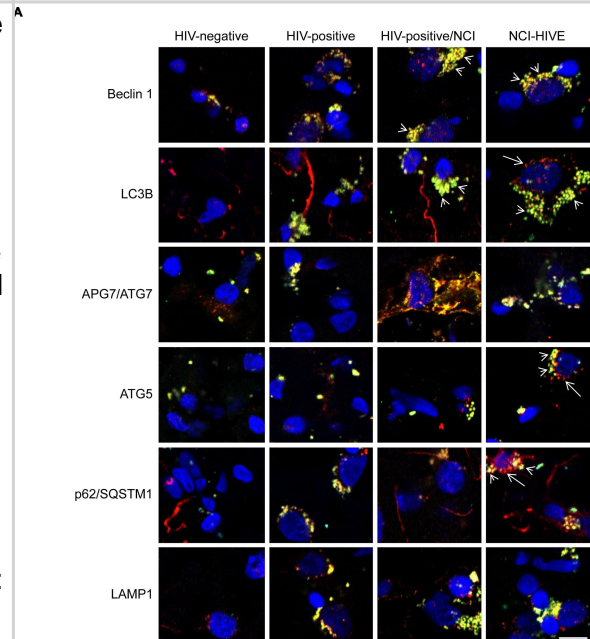
Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - Autophagy is required cell-autonomously in postsynaptic pyramidal neurons for LTD. a–c Time-plots of normalized fEPSP slope & bar graphs of average normalized fEPSP slope during last 15 min of recording after LFS (a) or NMDA (b) or DHPG (c) bath application in P22 hippocampal slices from control (atg5f/f, white circles & bars) & thy1-cKO (black circles & bars) animals. N = 7 animals per genotype. Representative traces of fEPSPs before (black) & after (red) LTD induction for two genotypes in every LTD-triggering protocol. d Schematic representation of protocol used to knockdown atg5 selectively in excitatory neurons of CA1 area of hippocampus in adult (P60) animals. e Images showing representative slices expressing AAV-CamK2a-shatg5-eGFP or AAV-CamK2a-shscrambled-mCherry in CA1 area, stained w/ Hoechst dye (nuclei). f Western blot analyses of Atg5, LC3, p62, & actin in lysates from shRNA-atg5 & control hippocampal slices to validate efficacy of knockdown protocol. Graph showing normalized protein levels of LC3, p62, & Atg5 in indicated condition. N = 9 animals. g Time-plot of normalized fEPSP slope & bar graphs of average normalized fEPSP slope during last 15 min of recording following LFS in slices expressing AAV-CamK2a-shscrambled-mCherry (white circles & bars) or AAV-CamK2a-shatg5-eGFP (black circles & bars) in CA1 area. N = 9 animals per condition. Representative traces of fEPSPs before (black) & after (red) LTD induction for two conditions. a–c, f, g Bars represent mean values \pm SEM. Statistical analyses performed using unpaired, two-tailed Student's t-test. a, $P = 0.0007$, b, $P < 0.0001$, c, $P = 0.0001$. In f, $P = 0.0231$ for LC3, $P = 0.0034$ for p62, & $P < 0.0001$ for ATG5. For g, $P < 0.0001$. Image collected & cropped by CiteAb from following publication (<https://pubmed.ncbi.nlm.nih.gov/35115539>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



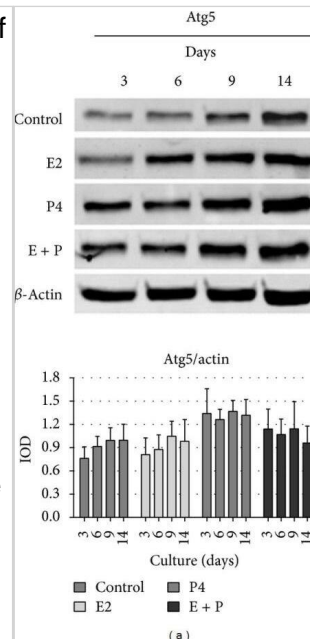
Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - IFN- γ increased rates of autophagy in primary BMECs in vitro. (a) The expression of IFNGRs in BMECs. BMECs treated w/ IFN- γ for 6, 24 or 48 h. At the end of the treatment period, the levels of IFNGR1 & IFNGR2 analysed using western blot analysis w/ specific antibodies as described in the Materials & Methods section. The data represent the mean \pm S.E.M. of three independent experiments. Error bars are \pm S.E.M. One-way ANOVA; * P <0.05; ** P <0.01. (b) BMECs treated w/ IFN- γ for 3, 6, 12 or 24 h. At the end of the treatment, levels of MAP1LC3, ATG12-ATG5, SQSTM1/p62 & GAPDH analysed using western blot analysis w/ specific antibodies as described in the Materials & Methods section. The data represent the mean \pm S.E.M. of three independent experiments. Error bars are \pm S.E.M. One-way ANOVA; * P <0.05; ** P <0.01. (c) BMECs treated w/ IFN- γ at 2.5, 5, 10 or 20 ng/ml. At the end of the treatment, levels of MAP1LC3, ATG12-ATG5, SQSTM1/p62 & GAPDH analysed using western blot analysis w/ specific antibodies as described in the Materials & Methods section. The data represent the mean \pm S.E.M. of three independent experiments. Error bars are \pm S.E.M. One-way ANOVA; * P <0.05; ** P <0.01. (d) Direct effect of IFN- γ on induction of autophagy. BMECs cultured for 24 h in supernatants from either mock-treated BMECs or cells treated w/ 10 ng/ml of IFN- γ for 24 h in the presence or absence of anti-IFNGR1, IFNGR2 or IFNGR1/2. At the end of the treatment, levels of MAP1LC3, ATG12-ATG5, SQSTM1/p62 & GAPDH analysed using western blot analysis w/ specific antibodies as described in the Materials & Methods section. The data represent the mean \pm S.E.M. of three independent experiments. Error bars are \pm S.E.M. One-way ANOVA; * P <0.05; ** P <0.01. Image collected & cropped by CiteAb from the following publication (<https://www.nature.com/articles/cddiscovery201565>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



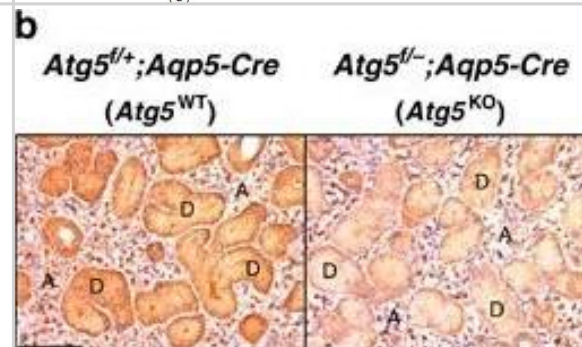
Immunocytochemistry/ Immunofluorescence: ATG5 Antibody - BSA Free [NB110-53818] - Autophagy associated protein immunoreactivity in HIV-infected brain tissue. (A) Representative images from five randomly selected fields of cells each examined in duplicate frontal lobe white matter sections for the indicated subject groups. The indicated proteins were labeled red & microglia with the cell-type-specific marker Iba1 (green). Blue staining indicates cell nuclei. Arrow heads indicate examples of higher Iba1 immunoreactivity whereas arrows indicate more focal (punctal) vs. diffuse (filamentous) patterns of autophagy associated protein expression. Scale bar = 10 μ m. (B) Quantification of relative Iba1 immunoreactivity from (A). $F(3,20) = 6.450$, $p = 0.0031$; $\square p < 0.05$ when compared to all other subject groups. Error bars show the SEM for the average values of 2–6 regions from each subject group across the six autophagy associated proteins examined. (C) Quantification of the indicated autophagy associated protein relative immunoreactivity from (A). Beclin 1: $F(3,12) = 11.29$, $p = 0.0008$; LC3B: $F(3,12) = 1.994$, $p = 0.1687$; APG7/ATG7: $F(3,12) = 84.20$, $p = < 0.0001$; ATG5: $F(3,12) = 6.218$, $p = 0.0086$; p62/SQSTM1: $F(3,12) = 87.04$, $p = < 0.0001$; LAMP1: $F(3,12) = 8.317$, $p = 0.0029$. $\square p < 0.05$ when compared to HIV-negative; $\#p < 0.05$ when compared to HIV-positive; & $\Omega p < 0.05$ when compared to HIV-positive/NCI subjects. Error bars show the SEM for four regions from each subject group. Image collected & cropped by CiteAb from the following publication (<http://journal.frontiersin.org/Article/10.3389/fmicb.2015.00653/abstract>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - Expression of autophagic proteins (Atg5, Atg3, LC3, & beclin-1) & phosphorylated Bcl-2 (Ser70) in BME-UV1 cells forming acinar structures on Matrigel. Representative images of Western blot analysis of Atg5 (a), Atg3 (b), LC3 (c), beclin-1 (d), & phosphorylated Bcl-2 (Ser70) (f) in bovine MECs grown in 3D culture for 3, 6, 9, & 14 days in differentiation medium (control), enriched with 17 β -estradiol (E2, 1 nM), progesterone (P4, 5 ng/mL), or both (E + P); expression of β -actin was used as a loading control; graphs below the images show the results of densitometric analysis, in which IOD of each band was measured, & the values were normalized to IOD of β -actin; the IOD results are presented as means \pm SEM from at least three separate experiments. (e) Confocal images of immunofluorescence staining of beclin-1 (green fluorescence) in cells grown in 3D culture for 3 or 9 days in control conditions: DNA was counterstained with 7AAD (red fluorescence) & white arrows indicate the nuclear localization of beclin-1. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/24895572>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



Immunohistochemistry: ATG5 Antibody - BSA Free [NB110-53818] - Elevated basal expression of proinflammatory cytokines genes in Atg5-knockout SMGs. (a) Schematic diagram of generation of experimental mice. Mice developed a spontaneous heterologous Atg5 deletion after five generations of crossing between Atg5f/f & Atg5f/f;Aqp5-Cre mice, resulting in Atg5f/-;Aqp5-Cre mice. Atg5f/+;Aqp5-Cre (Atg5WT) & Atg5f/-;Aqp5-Cre (Atg5KO) mice were used in studies herein. (b) Immunohistochemical analyses show decreased ATG5 protein in both SMG granular convoluted ducts (GCDs; labeled D) & acinar cells (labeled A) of Atg5KO mice, compared with that of Atg5WT mice. Bar: 100 μ m. (c) Correlation of decreased ATG5 expression & impaired MAP1LC3 lipidation in SMGs among different genotypes. Equal amounts of whole SMG lysates from two individual mouse of the indicated genotype were analyzed for ATG5 & MAP1LC3 levels by western blots. ATG5 expression was greatly reduced in the SMGs of Atg5F/F than that in Atg5+/+ mice. This hypomorphic phenotype in floxed mouse line has been reported previously in the loxP mouse gene targeting system.18+/+, Atg5+/+, +/-, Atg5+/-, +/-C, Atg5+/-;Aqp5-Cre; F/F, Atg5F/F; F/+, Atg5F/+; F/+C, Atg5F/+;Aqp5-Cre (or Atg5WT); F/-, Atg5F/-; F/-C, Atg5F/-;Aqp5-Cre (or Atg5KO). Relative ATG5 levels were determined by setting average level of ATG5 in Atg5+/+ mice as 1. (d) Quantitative RT-PCR analyses show elevated basal expression of selected proinflammatory cytokine genes in SMGs from Atg5KO mice. Respective mean expression level of the indicated message from SMGs of Atg5WT mice was set as 1. Results are shown as mean \pm S.D.; N=3; *P<0.05 Image collected & cropped by CiteAb from the following publication (<https://www.nature.com/articles/cddis2014428>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

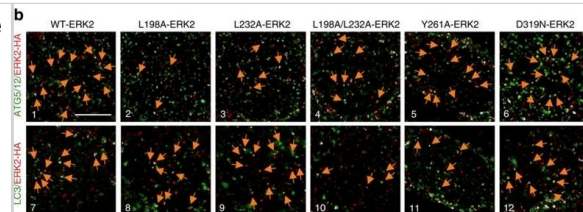


Immunocytochemistry/ Immunofluorescence: ATG5 Antibody - BSA Free [NB110-53818] - ERK2 utilizes kinase-docking domains to interact with ATG5–ATG12 & LC3-II. (a) Mutations in FRS on ERK2 decrease colocalization of ERK2 with ATG5–ATG12, LC3 & WIPI1.

Immunofluorescence (IF) showing colocalization (depicted as white pixels by 'colocalization finder application') of WT-ERK2-HA, FRS ERK2 mutants (L198A-, L232A-, L198A/L232A-, Y261A-ERK2-HA) or common docking (CD) mutant (D319N-ERK2-HA) with ATG5–ATG12 (panels 1–6), LC3 (panels 7–12) or WIPI1 (panels 13–18) in EGF-treated NIH/3T3 cells. ERK2 is stained in red, & autophagy proteins are stained in green. Scale bar, 10 μ m. The bars represent mean \pm s.e.m. ** P <0.01, **** P <0.0001 compared with WT-ERK2-transfected cells; Student's t-test, 50 cells analysed from $n=2$.

(b) Mutations in FRS on ERK2 decrease colocalization of ERK2 with nuclear ATG5–ATG12 or LC3. IF showing colocalization (white pixels) of WT-ERK2-HA (red), FRS ERK2 mutants (L198A-, L232A-, L198A/L232A-, Y261A-ERK2-HA) or common docking (CD) mutant (D319N-ERK2-HA) with ATG5–ATG12 (panels 1–6) or LC3-II (panels 7–12) in EGF-treated NIH/3T3 cells. Scale bar, 5 μ m. The bars represent mean \pm s.e.m. ** P <0.01, *** P <0.001, **** P <0.0001 compared with WT-ERK2-transfected cells; Student's t-test, 50 cells analysed from $n=2$. Arrows depict colocalization.

(c) ATG5–ATG12 conjugation is required for ERK phosphorylation. Immunoblots for P-ERK, total ERK & β -actin in NIH/3T3 cells transfected with WT ATG5 or the conjugation-defective ATG5 K130R mutant & treated with EGF (10 min). The bars represent mean \pm s.e.m. * P <0.05 compared with WT ATG5-transfected cells; Student's t-test, $n=3$. Image collected & cropped by CiteAb from the following publication (<https://www.nature.com/articles/ncomms3799>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.

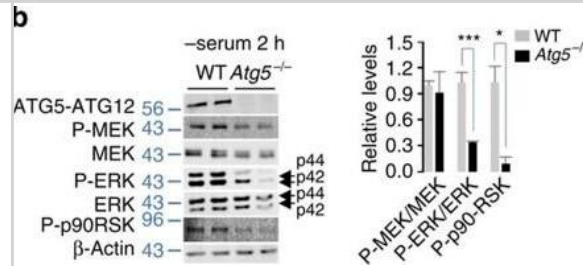


Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - ERK phosphorylation in *Atg5*^{-/-} cells depends on nutrient availability. (a) *Atg5*^{-/-} MEFs display reduced ERK phosphorylation during nutrient deprivation. Quantification of P-ERK levels normalized to total ERK (detected by immunoblotting) in WT & *Atg5*^{-/-} MEFs cultured in the absence of serum for indicated times. The bars represent mean \pm s.e.m. * P <0.05 compared with corresponding WT value; *** P <0.001, **** P <0.0001 compared with 10 min serum-starved *Atg5*^{-/-} MEFs; ANOVA-Bonferroni post hoc test, $n=3$.

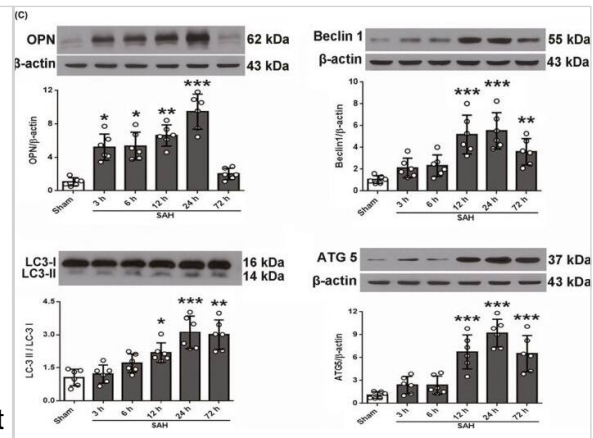
(b) Serum-deprived *Atg5*^{-/-} MEFs display decreased ERK & p90RSK phosphorylation. Immunoblots for the indicated proteins in total lysates from 2 h serum-deprived WT & *Atg5*^{-/-} MEFs. The bars represent mean \pm s.e.m. * P <0.05, *** P <0.001 compared with Con; Student's t-test, $n=3$.

(c) *Atg5*^{-/-} MEFs display decreased nuclear ERK activity. Immunoblots for phosphorylated HA (haemagglutinin)-tagged Elk1 in total lysates from 2 h serum-deprived WT & *Atg5*^{-/-} MEFs cells exposed or not to EGF (10 min). (d) *Atg5*^{-/-} MEFs display decreased Elk1-driven gene expression. Elk1-driven ZFP36 mRNA levels from 2 h serum-deprived WT & *Atg5*^{-/-} MEFs cells exposed or not to EGF (10 min). The bars represent mean \pm s.e.m. *** P <0.001 compared with Con; Student's t-test, $n=3$.

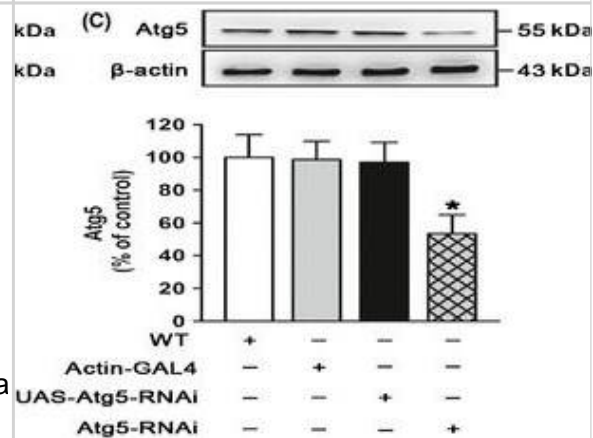
(e) Decreased ERK phosphorylation in *Atg5*^{-/-} MEFs occurs independently of changes in ERK phosphatases. Immunoblots for the indicated proteins in 2 h serum-deprived WT MEFs transfected with scrambled siRNA (scr), & *Atg5*^{-/-} MEFs transfected with scr or siRNAs against MKP3 or PP2A in the presence or absence of EGF. The bars represent mean \pm s.e.m., $n=3$. Image collected & cropped by CiteAb from the following publication (<https://www.nature.com/articles/ncomms3799>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



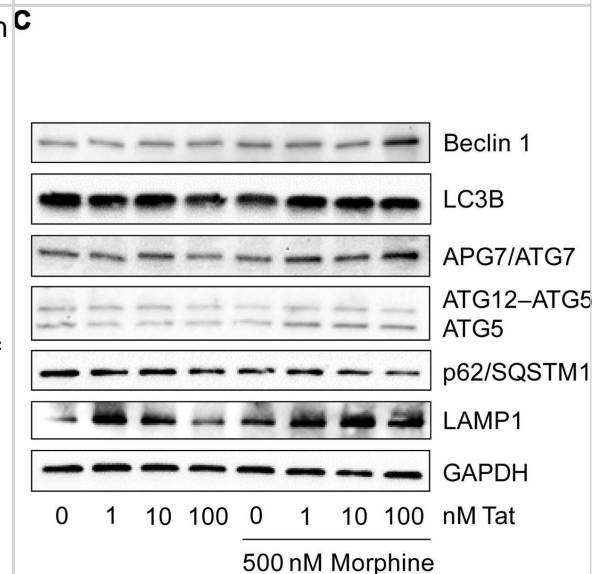
Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - Subarachnoid hemorrhage (SAH) grades & temporal expression of endogenous osteopontin (OPN) & autophagy-related proteins in rat brain after SAH. A, Representative brain images of Sham & SAH rats. B, Summary of SAH grading scores of all groups. Sample size is 27, n = 9 per group. Data were analyzed using Kruskal-Wallis test, $\chi^2 = 18.183$. C, Representative Western blot images & quantitative analyses of OPN, Beclin 1, LC3, & ATG 5 from the left hemisphere of rat brains at different time points after SAH. Sample size is 36, n = 6 per group. Data were presented as mean \pm SD. $F = 28.45$ for OPN, $F = 12.37$ for Beclin 1, $F = 18.88$ for LC3, $F = 22.14$ for ATG5. * $P < .05$, ** $P < .01$, *** $P < .001$ vs Sham group. SAH, subarachnoid hemorrhage; Vehicle, phosphate-buffered saline; rOPN, recombinant OPN. NS, not significant Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/31436915>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - β -guanidinopropionic acid (GPA) promotes autophagy that is essential for longevity in *Drosophila*. Western blot analysis of Atg8 in wild-type (WT) flies. β -GPA was found to efficiently increase the levels of Atg8 in flies (n = 6, one-way ANOVA, * $P < 0.05$ vs. control). (B) The levels of P62 were downregulated upon β -GPA administration (n = 6, $P < 0.05$ vs. control, Student's *t*-test). (C) Western blot analysis of Atg5 in actin-GAL4>UAS-Atg5-RNAi flies (Atg5-RNAi). Atg5 was reduced in Atg5-RNAi flies (n = 6, one-way ANOVA, * $P < 0.05$ vs. control). (D, E) Downregulation of autophagy abolished the β -GPA-mediated lifespan extension (n = 200, $P < 0.0001$ vs. control, log-rank test). For all Western blots, relative band intensity was estimated using IMAGEJ. Data are shown as mean \pm SEM. Image collected & cropped by CiteAb from the following publication (<https://pubmed.ncbi.nlm.nih.gov/26120775>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



Western Blot: ATG5 Antibody - BSA Free [NB110-53818] - Differences in neuronal autophagy & dendrite varicosity following HIV-1 Tat protein & morphine treatment. (A) Representative images of neurons transfected with a fluorescent reporter plasmid to monitor autophagic flux at 8 h following the indicated treatments. GFP (green) & GFP + mRFP (yellow) fluorescence are observed prior to the fusion of autophagosomes with lysosomes whereas only mRFP (red) fluorescence is present in post-fusion autolysosomes. DIC, differential interference contrast microscopy image. DAPI (blue) staining indicates cell nuclei. (B) Quantification of autolysosomes (red puncta) from (A). $F(3,13) = 8.756$, $p = 0.0019$; $p < 0.05$ when compared to all other groups. (C) Western blotting analysis of the indicated autophagy associated protein levels at 24 h following the indicated treatments. GAPDH was used as a loading control. Blots are representative of three independent experiments. (D) Quantification of dendrite beading from (A). $F(3,77) = 6.429$, $p = 0.0006$; $p < 0.05$ when compared to control cells. Error bars show the SEM. Image collected & cropped by CiteAb from the following publication (<http://journal.frontiersin.org/Article/10.3389/fmicb.2015.00653/abstract>), licensed under a CC-BY license. Not internally tested by Novus Biologicals.



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Alan P, Vandevoorde KR, Joshi B et al. Basal Gp78-dependent mitophagy promotes mitochondrial health and limits mitochondrial ROS *Cellular and Molecular Life Sciences* 2022-11-01 [PMID: 36284011]

Sekiguchi K, Miyahara H, Inoue M et al. Metabolome Characteristics of Liver Autophagy Deficiency under Starvation Conditions in Infancy *Nutrients* 2021-08-29 [PMID: 34578904]

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MKM Karuppan, CR Ojha, M Rodriguez, J Lapierre, MJ Aman, F Kashanchi, M Toborek, M Nair, N El-Hage Reduced-Beclin1-Expressing Mice Infected with Zika-R103451 and Viral-Associated Pathology during Pregnancy *Viruses*, 2020-06-02;12(6):. 2020-06-02 [PMID: 32498399]

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Inoue M, Miyahara H, Shiraishi H et al. Leucyl-tRNA synthetase deficiency systemically induces excessive autophagy in zebrafish *Scientific Reports* 2021-04-16 [PMID: 33863987]

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S Mukhopadhy, J Encarnació, EY Lin, ASW Sohn, H Zhang, JD Mancias, AC Kimmelman Autophagy supports mitochondrial metabolism through the regulation of iron homeostasis in pancreatic cancer *Science Advances*, 2023-04-19;9(16):eadf9284. 2023-04-19 [PMID: 37075122]

More publications at <http://www.novusbio.com/NB110-53818>



Procedures

Western Blot protocol for ATG5 Antibody (NB110-53818)

Protocol: Western Blot Protocol for Atg5 Antibody (NB110-53818)

Materials

- 1X PBS
- Sample buffer, 2X Laemmli buffer: 4% SDS, 5% 2-mercaptoethanol (BME), 20% glycerol, 0.004% bromophenol blue, 0.125 M Tris HCl, pH 6.8
- 1X Running Buffer: 25 mM Tris-base, 192 mM glycine, 0.1% SDS. Adjust to pH 8.3
- 1X Transfer buffer (wet): 25 mM Tris-base, 192 mM glycine, 20% methanol Adjust to pH 8.3
- TBS
- TBST, TBS and 0.1% Tween
- Blocking solution: TBST, 5% non-fat dry milk
- rabbit anti-Atg5 primary antibody (NB110-53818) in blocking buffer (1:500)

Methods

1. Grow cells (e.g. HeLa or Neuro2A) in vitro to semi-confluency (70-75%).
2. Rinse cells with ice-cold 1X PBS and lyse cells with sample buffer.
3. Sonicate and incubate cells for 5 minutes at 95°C.

Tip: Cells are lysed directly in sample buffer.

4. Load 10-40 µg/lane of sample on a 12% polyacrylamide gel (SDS-PAGE).
5. Transfer proteins to a PVDF membrane for 60 minutes at 100V.

Tip: For more information on Western Blotting, see our Western Blot handbook:

6. After transfer, rinse the membrane with dH₂O and stain with Ponceau S for 1-2 minutes to confirm efficiency of protein transfer.
7. Rinse the membrane in dH₂O to remove excess stain and mark the loaded lanes and molecular weight markers using a pencil.
8. Block the membrane using blocking buffer solution (5% BSA in TBST) for 1 hour at room temperature.
9. Rinse the membrane with TBST for 5 minutes.
10. Dilute anti-Atg5 primary antibody (NB110-53818) in blocking buffer (1:500) and incubate the membrane for 1 hour at room temperature.
11. Rinse the membrane with dH₂O.
12. Rinse the membrane with TBST, 3 times for 10 minutes each.
13. Incubate the membrane with diluted secondary antibody, according with product's specification, (e.g. anti-rabbit-IgG HRP-conjugated) in blocking buffer for 1 hour at room temperature.

Note: Tween-20 may be added to the blocking or antibody dilution buffer at a final concentration of 0.05-0.2%, provided it does not interfere with antibody-antigen binding.

14. Rinse the membrane with TBST, 3 times for 10 minutes each.
15. Apply the detection reagent of choice (e.g. BioFX Super Plus ECL) in accordance with the manufacturer's instructions.

Deparaffinization:

1. Treat slides with Xylene: 3 changes for 5 minutes each. Drain slides for 10 seconds between changes.
2. Treat slides with 100% Reagent Alcohol: 3 changes for 5 minutes each. Drain slides for 10 seconds between changes.

Quench Endogenous Peroxidase:

1. Place slides in peroxidase quenching solution: 15-30 minutes. To Prepare 200ml of Quenching Solution: Add 3ml of 30% Hydrogen Peroxide to 200ml of Methanol. Use within 4 hours of preparation.
2. Place slides in distilled water: 2 changes for 2 minutes each.

Retrieve Epitopes:

1. Preheat Citrate Buffer. Place 200ml of Citrate Buffer Working Solution into container, cover and place into steamer. Heat to 90-96 degrees C.
2. Place rack of slides into hot Citrate Buffer for 20 minutes. Cover.
3. Carefully remove container with slides from steamer and cool on bench, uncovered, for 20 minutes.
4. Slowly add distilled water to further cool for 5 minutes.
5. Rinse slides with distilled water. 2 changes for 2 minutes each.

Immunostaining Procedure:

1. Remove each slide from rack and circle tissue section with a hydrophobic barrier pen (e.g. Liquid Blocker-Super Pap-Pen).
2. Flood slide with wash solution. Do not allow tissue sections to dry for the rest of the procedure.
3. Drain wash solution and apply 4 drops of blocking reagent to each slide and incubate for 15 minutes.
4. Drain Blocking Reagent (do not wash off the Blocking Reagent), apply 200ul of primary antibody solution to each slide, and incubate for 1 hour.
5. Wash slides with wash solution: 3 changes for 5 minutes each.
6. Drain wash solution, apply 4 drops of secondary antibody to each slide and incubate for 1 hour.
7. Wash slides with wash solution: 3 changes for 5 minutes each.
8. Drain wash solution, apply 4 drops of DAB substrate to each slide and develop for 5-10 minutes. Check development with microscope.
9. Wash slides with wash solution: 3 changes for 5 minutes each.
10. Drain wash solution, apply 4 drops of Hematoxylin to each slide and stain for 1-3 minutes. Increase time if darker counterstaining is desired.
11. Wash slides with wash solution: 2-3 changes for 2 minutes each.
12. Drain wash solution and apply 4 drops of Bluing Solution to each slide for 1-2 minutes.
13. Rinse slides in distilled water.
14. Soak slides in 70% reagent alcohol: 3 minutes with intermittent agitation.
15. Soak slides in 95% reagent alcohol: 2 changes for 3 minutes each with intermittent agitation.
16. Soak slides in 100% reagent alcohol: 3 changes for 3 minutes each with intermittent agitation. Drain slides for 10 seconds between each change.
17. Soak slides in Xylene: 3 changes for 3 minutes each with intermittent agitation. Drain slides for 10 seconds between each change.
18. Apply 2-3 drops of non-aqueous mounting media to each slide.
19. Lay slides on a flat surface to dry prior to viewing under microscope.

NOTES:

- Use treated slides (e.g. HistoBond) to ensure adherence of FFPE sections to slide.
- Prior to deparaffinization, heat slides overnight in a 60 degrees C oven.
- All steps in which Xylene is used should be performed in a fume hood.
- For Epitope Retrieval, a microwave or pressure cooker may be substituted for the steamer method. Adjust times as necessary depending on conditions.
- For the initial IHC run with a new primary antibody, test tissues with and without Epitope Retrieval. In some instances, Epitope Retrieval may not be necessary.
- 200ul is the recommended maximum volume to apply to a slide for full coverage. Using more than 200ul may allow solutions to wick off the slide and create drying artifacts. For small tissue sections, less than 200ul may be used.
- 5 minutes of development with DAB substrate should be sufficient. Do not develop for more than 10 minutes. If 5 minutes of development causes background staining, further dilution of the primary antibody may be necessary.
- Hematoxylin should produce a light nuclear counterstain so as not to obscure the DAB staining. Counterstain for 1-2 minutes for nuclear antigens. Counterstain for 2-3 minutes for cytoplasmic and membranous antigens. If darker counterstaining is desired, increase the time (up to 10 minutes).



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Limitations

This product is for research use only and is not approved for use in humans or in clinical diagnosis. Primary Antibodies are guaranteed for 1 year from date of receipt.

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